Peer reviewed article

Restorative neuroscience: concepts and perspectives

Robert H. Andresa, Morten Meyerb, Angélique D. Ducraya, Hans R. Widmera

- ^a Department of Neurosurgery, University of Berne, Inselspital, Berne, Switzerland
- b Department of Anatomy and Neurobiology, Institute of Medical Biology, University of Southern Denmark, DK-Odense, Denmark

Summary

There is increasing interest in the search for therapeutic options for diseases and injuries of the central nervous system (CNS), for which currently no effective treatment strategies are available. Replacement of damaged cells and restoration of function can be accomplished by transplantation of cells derived from different sources, such as human foetal tissue, genetically modified cell lines, embryonic or somatic stem cells. Preclinical and clinical trials have shown promising results in neurodegenerative disorders, like Parkinson's and Huntington's disease, but also ischaemic stroke, intracerebral haemorrhage, demyelinating disorders, epilepsy and traumatic lesions of the brain and spinal cord. Other studies

have focused on finding new ways to activate and direct endogenous repair mechanisms in the CNS, eg, by exposure to specific neuronal growth factors or by inactivating inhibitory molecules. Neuroprotective drugs may offer an additional tool for improving neuronal survival in acute or chronic CNS diseases. Importantly however, a number of scientific issues need to be addressed in order to permit the introduction of these experimental techniques in the wider clinical setting.

Key words: neural transplantation; stem cells; neurotrophic factors; neuroprotection; regenerative medicine

List of Abbreviations

AAV	adeno-associated virus		
AD	Alzheimer's disease		
ALS	amyotrophic lateral sclerosis		
APP	amyloid precursor protein		
ARTN	artemin		
beta-Gal	beta-galactosidase		
BDNF	brain-derived neurotrophic factor		
BFGF	basic fibroblast growth factor		
BMP	bone morphogenetic protein		
CDNF	conserved dopamine neurotrophic factor		
CNTF	Ciliary neurotrophic factor		
CK	creatine kinase		
CNS	central nervous system		
EGF	epidermal growth factor		
ES	embryonic stem cell		
FDG	fluorodopa deoxyglucose		
FGF	fibroblast growth factor		
GABA	gamma-aminobutyric acid		
GDNF	glial cell line-derived neurotrophic factor		
GFL	GDNF family ligand		

GFP	green fluorescent protein		
HD	Huntington's disease		
ICH	intracerebral haemorrhage		
IGF	insulin-like growth factor		
IL	interleukin		
MS	multiple sclerosis		
NGF	nerve growth factor		
NT	neurotrophin		
NTN	neurturin		
OEC	olfactory ensheathing cell		
PCr	phosphocreatine		
PERV	porcine endogenous retrovirus		
PET	positron emission tomography		
PD	Parkinson's disease		
PSP	persephin		
SOD1	superoxid dismutase 1		
SVZ	subventricular zone		
SV40	simian virus 40		
TGF	transforming growth factor		
TH	tyrosine hydroxylase		
VM	vental mesencephalon		

No financial support declared.

Introduction

The diseased or damaged brain has limited regenerative capacity, which is mainly of a functional and not of a structural nature. There are a number of neurodegenerative processes, neurovascular pathologies and traumatic lesions of the central nervous system (CNS) for which there are currently no effective treatment options available. These are usually devastating diseases with a major impact on quality of life, showing a chronic course and are associated with high socioeconomic costs. Due to increasing life expectancy and a higher prevalence of neurodegenerative and neurovascular pathologies in the elderly population, these disorders will become even more important for our society in the future and there is need for the development of new, adequate treatment options.

To develop strategies for repair of the impaired brain and spinal cord, extensive research efforts have been implemented particularly during the last two decades. Effective *in vitro* and *in vivo* models have been developed in order to replicate the core pathology of the underlying disorders. The concept of neural transplantation has evolved as an instrument for replacing the neurons lost in degenerative processes, trauma, and vascular lesions, as well as for replacing glial cells in the context of demyelinating lesions. Major advances in basic research have enabled first clinical trials, which have proved that this ap-

proach is feasible and effective. In addition, strategies have been developed to influence endogenous stem cell proliferation, migration and differentiation in the brain.

Nevertheless, there are still a number of major limitations to overcome, such as lack of sufficient and well-characterised donor tissue, suboptimal survival and functional integration of transplanted cells, the presence of side effects, and the missing knowledge on factors influencing migration, growth and differentiation of transplanted stem and progenitor cells. In addition, and importantly also a number of ethical issues needs to be addressed. In this article we discuss the possibilities and limitations in cell replacement strategies and also address current research in restorative neuroscience. Notably, the current review deals with a rather specific part of current plasticity research and does not cover the entire field of neuronal plasticity, like for example, mechanisms of synaptic plasticity [1, 2], activity dependent modulation of axonal motility [3], effects of dietary restriction [4], potential of environmental enrichment [5], the impact of adaptive learning for rehabilitation [6], assessment of structural plasticity by transcranial magnetic stimulation [7], multimodal imaging of brain reorganisation [8], and the importance of nuclear medicine imaging in rehabilitative treatment evaluation [9].

Neural transplantation for Parkinson's and Huntington's disease

Allogenic neuronal tissue harvested within certain developmental windows has been reported to survive, extend axons and make connections with the surrounding host brain after transplantation. Parkinson's disease (PD) and Huntington's disease (HD), common neurodegenerative disorders with relatively selective loss of certain subpopulations of neurons, have received the most attention with respect to therapies designed to replace the missing neurons [10, 11]. While there are effective symptomatic treatments for PD, the

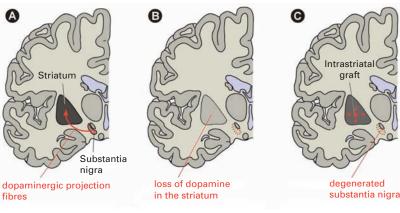
applied drugs become less effective with the progression of the disease and produce significant side effects. In HD, there are at present only few symptomatic treatments available, which are most effective in controlling the psychiatric abnormalities associated with this disease. Both PD and HD can be mimicked in experimental *in vitro* and *in vivo* models to replicate the underlying pathological processes permitting study of cell replacement strategies under laboratory conditions.

Transplantation of foetal neuronal tissue

Parkinson's Disease

Idiopathic PD is the second most common neurodegenerative disorder and affects more than 1% of all individuals over the age of fifty years [12]. In Switzerland, about 10 000 to 12 000 persons suffer from this disease. As the incidence of PD rises with age, it is expected that this number will increase significantly because of the aging

character of our society [13]. Clinical symptoms are resting tremor, bradykinesia, rigidity and postural imbalance [14, 15]. PD is characterised by a predominant and progressive loss of dopaminergic neurons in the substantia nigra pars compacta in the upper brain stem, which leads to a profound loss of dopaminergic input into the striatum. It has been shown that complex I activity is defective



Normal function of the dopaminergic nigrostriatal pathway in the healthy brain

Normal brain

Degeneration of nigral dopaminergic neurons in PD resulting in depletion of dopamine in the striatum

Parkinson's disease

Re-innervation of the striatum by grafted dopaminergic cells resulting in restoration of dopaminergic input After transplantation

Figure 1

Schematic drawing of transplantation of dopaminergic cells in a patient suffering from Parkinson's disease (PD). In the healthy brain, dopaminergic input to the striatum is provided by the nigrostriatal projection system (A). In PD, degeneration of the dopaminergic neurons in the substantia nigra leads to dopamine depletion in the striatum and thus dysfunction of the extrapyramidal system (B). After intrastriatal transplantation of dopaminergic neuronal precursor cells, these differentiate into neurons, establish functional connections to the surrounding striatal cells and restore the dopaminergic input to the striatum (C).

in multiple tissues from PD patients [16], and that there is a maternal association in the hereditary form of PD [17], suggesting a mitochondrial basis for this disease.

Due to the fact that PD represents a rather selective degenerative process of mainly dopaminergic neurons of the nigrostriatal pathway, this pathology has been considered as particularly suitable for the application of cell replacement therapies (fig. 1). Extensive *in vivo* studies have shown that foetal ventral mesencephalic (VM) allografts display long-term survival in the host brain, making and receiving connections from host neurons [18, 19] (fig. 2). The transplanted tissue releases dopamine in a regulated

Figure 2

Photomicrograph of rat embryonic dopaminergic precursor cells transplanted in a rat model of Parkinson's disease and immunohistochemically stained for tyrosine hydroxylase (TH). TH-immunoreactive grafted cells (arrows) survive in the host brain and extend TH-immunoreactive axons into the denervated striatum (asterisk). Scale bar: 200 μm.



fashion and reverses many of the behavioural deficits seen in animal models of PD [20, 21]. Based on the experimental data, first clinical trials with neuronal transplantation in patients were started in the late 1980s. Foetal nigral tissue can be transplanted safely into the caudate and putamen bilaterally in patients with PD and with little post-operative complications [22, 23]. The function of the grafted cells can be assessed by fluorodopa deoxyglucose positron emission tomography (FDG-PET). Neuropathological evidence has been provided that human foetal VM grafts survived and reinnervated the host striatum of a PD patient who had shown significant clinical improvement as well as enhanced fluorodopa uptake on PET scans [23-26]. Long-term graft survival was confirmed in two patients with persisting high FDG uptake when investigated 6 and 12 years after surgery [27, 28]. Significant clinical improvement associated with graft survival has been reported by several groups, mainly characterised by reduced rigidity and bradykinesia, with the ability to completely withdraw L-DOPA treatment after surgery in the most successful cases.

Notably, less than 20% of the transplanted dopaminergic cells survive the transplantation procedure [24, 29]. Studies in rats indicate that most of these cells die within one week posttransplantation [30], predominantly by apoptosis [31]. Hence it has become evident from the clinical trials performed so far that significant clinical improvement is only achieved after grafting of a sufficient amount of VM tissue (cells from 3 to 4 human embryos per side) [32, 22] followed by favourable integration of the grafted dopaminergic neurons into the host brain. In addition, the position of the graft has been shown to play a major role in the pathogenesis of novel dyskinetic behaviour after transplantation in PD rats, and widespread grafting could be an option to overcome this problem [33].

The recently reported double-blind studies by Freed and coworkers and Olanow and colleagues both failed to meet their primary end points [34, 35]. Moreover, several patients developed severe side effects including dystonia and dyskinesias [34, 36,37]. This might have been due to insufficient dopamine release by the transplants [35, 38]. Both studies, however, included subjects who had failed drug therapy and it has been suggested that patient selection may explain the negative results reported by Olanow and colleagues [39]. It is likely that better results can be expected with improved transplant protocols [35, 40, 41]. The complexity of these problems should not be underestimated and clinical applications should be planned with great care [37, 42]. These notions ask for improved understanding of the transplantation approaches and require extended experimental studies. Winkler and co-authors concluded that with further improvement and refinement of the grafting procedure there is every

reason to believe that cell transplantation can be developed into a safe and efficacious restorative therapy for advanced not too seriously affected PD patients [41].

Taken together, neural transplantation therapy in PD using tissue from aborted foetuses is likely to remain experimental due to questions regarding standardisation and purity of cell material [37, 42]. Due to the above mentioned limitations, transplantation of human foetal tissue has therefore been largely abandoned at present and the basic research focuses on alternatives to human foetal tissue as a graft source, eg, transplantation of immortalised neuronal cell lines, embryonic and neural stem cells, porcine tissue and genetically modified cells [43].

Huntington's disease

HD is a chronic autosomal dominant inherited neurodegenerative disorder with full penetrance by mid-adult life and clinically presents with progressive choreoathetotic movements in combination with severe cognitive and emotional dysfunction [44, 45]. The symptoms typically appear between the age of 30 and 40 years and deteriorate over a 10-20 year period, finally leading to death [46]. HD has an overall prevalence rate in Europe of approximately 10 per 100 000 people [47]. The main pathological finding is a selective loss of the GABA-ergic medium sized spiny projection neurons in the striatum. The gene (IT15) located on the short arm of chromosome 4 [48] responsible for the expression of HD and its associated protein, huntingtin [49], have been identified through genetic research. The gene has been shown to contain a multiplication of CAG trinucleotide repeats responsible for different grades of severity in the course of the disorder, while the function of huntingtin has still to be elucidated. A defect in energy metabolism has been proposed as one of the potential pathogenetic mechanisms [50]. Nevertheless, a wide gap still remains between the knowledge about the neuronal substrates of HD and the ability to prevent or alleviate the progression of the disorder.

Transplantation of embryonic striatal precursor cells has been shown to restore deficits in experimental animal models of HD [51-53]. Similarly to the situation in PD, clinical trials for cell replacement strategies in HD using human foetal tissue as cell source revealed survival of grafts in the host brain. Most of the trials on cell transplantation in HD published so far were dealing with feasibility and safety issues. On the basis of theses results, one can assume that cell replacement for HD using foetal tissue is safe for the patients [54, 55]. Particularly interesting is the fact that in the study of Hauser et al. surviving graft tissue did not contain mutant huntingtin, indicating that the transplanted tissue was not affected by the pathology of the host brain [56, 57]. In a recent study, Keene and co-workers reported on long-term survival and neuronal differentiation of transplanted foetal tissue, however, the host brain was found to be poorly innervated. In line with this observation, the clinical benefit was found to be rather poor in transplanted patients [58]. Based on the outcome of this study, it was highlighted that it is mandatory to perform clinical trials in HD with great care and by means of blinded, well-designed and controlled studies [59]. Nevertheless, the outcome of the French HD trial showed minor but significant clinical improvement in some of the transplanted patients [60, 61]. A multi-centre trial was recently initiated to evaluate the best protocols for cell replacement in HD in order to make this technique available for a broader range of non-specialist centres and to verify these initial promising results (for review see: [62]). As stated for cell transplantation approaches in PD, foetal tissue will not provide a sufficient and standardised cell source for grafting a large number of HD patients. Importantly to note, cell transplantation is considered at present to be the only choice for restorative treatment of HD patients, hence research in this area as well as searching for alternative cell sources should be pursued [62].

Xenotransplantation

Xenotransplantation means that tissue is transplanted across the species barrier. Using organs from other species has long been considered for overcoming a shortage of human donor organs, such as heart, kidney and liver [63].

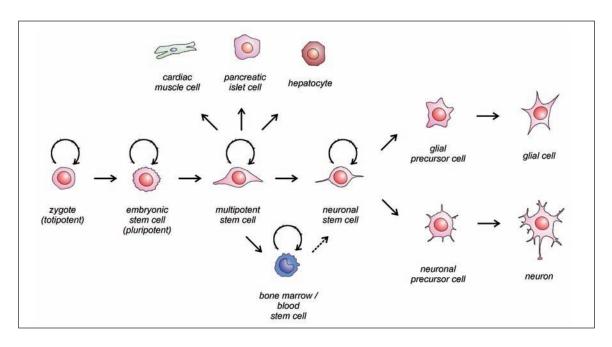
As a non-endangered species, pigs are considered a suitable source of donor tissue allowing sterile dissection of large quantities of pathogen-free tissue of the optimal embryonic age. A relatively large brain size combined with a protracted gestational period may provide the basis for long-distance axonal growth after grafting, facilitating efficient innervation of the host striatum. To what extent grafted dopaminergic neurons are affected by the on-going disease is still not known, but natural species differences, including genetic resistance in response to the disease process, could provide an important advantage promoting long-term survival and function of grafted porcine cells.

The transplantation of embryonic porcine xenografts has been established after extensive studies in Sweden and in the USA, both in PD and HD [64]. A clinical safety trial with porcine tissue including 12 Parkinson patients has been published [65, 66]. Moderate clinical improvements in some of the patients were reported. In the brain of a patient who died for unrelated causes a very limited survival rate of the dopaminergic neurons, not accompanied by major rejection processes, was observed [65]. This finding may point to lack of trophic support of grafted tissue after transplantations similar to the suggestions made in respect to allografts.

The risk of spread of infection across the species barrier (xenozoonosis), particularly by

Figure 3

Diagram illustrating the potential for differentiation of stem cells. Embryonic stem cells can differentiate into almost all cells of our body. With ongoing maturation, their ability to differentiate into different types of cells becomes more and more restricted. The differentiation capability of neural stem cells is restricted to the neuronal and glial lineage. Bone marrow stem cells have the capability to transdifferentiate into neural stem cells (dashed arrow). Stem cells can proliferate indefinitely (selfrenewal; circular arrows).



porcine endogenous retroviruses (PERVs), remains a major obstacle that hampers further clinical studies. However, in several studies investigating the possibility of cross-species infectivity, including a retrospective analysis of 160 human transplant recipients exposed to porcine tissue, no evidence for such transmission has been found [67, 68]. Cross-species rejection issues requiring immunosuppression of the host constitute another unsolved problem. The ability to genetically modify species such as the pig to express human genes and silence those provoking immune response has enabled interesting perspectives for genetically engineered cells in this context. The production of a gal-α-1,3-galtransferase transgenic pig [69] represents a significant advance towards eliminating hyperacute and acute vascular rejection.

Stem cells

Stem cells are undifferentiated cells without mature, tissue-specific characteristics that in response to proper stimuli are able to proliferate, to reproduce themselves and to produce generations of progenitor cells, which can differentiate into one or more cell types.

Several strategies are currently being investigated aiming at transplanting cells derived from a variety of different stem cells, including embryonic stem (ES) cells, neural stem cells, bone marrow or mesenchymal stem cells as well as umbilical cord blood stem cells (fig. 3).

Human ES are derived from preimplantation embryos generated for *in vitro* fertilisation. Within a few days after fertilisation, they can be removed from the inner cell mass of the blastocyst, dissociated and propagated in specialised cell culture media, where they can proliferate indefinitely [70]. Differentiation of these cells can be induced by changing culture conditions and exposure to specific growth factors. Due to their pluripotency, ES cells can potentially become any cell in the body, which offers a huge potential for cell replacement therapies [71].

Neural stem cells are found in already developed tissues of the foetus or the newborn, juvenile

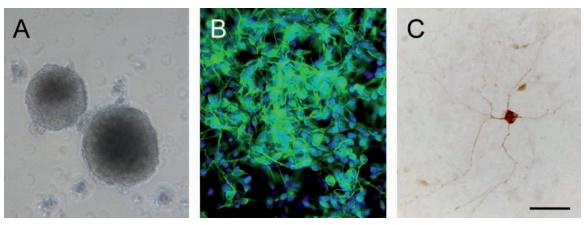


Figure 4

Photomicrographs of rat embryonic subventricular zone stem cells cultured as neurospheres (A). Differentiation of the cultured cells (Hoechst staining, blue) towards the neuronal phenotype is demonstrated by microtubule associated protein 2 immunostaining (green) and neurite outgrowth (B). Further differentiation towards specific neuronal subpopulations, eg, dopaminergic cells, is demonstrated using immunohistochemistry for the catecholaminergic marker tyrosine hydroxylase (C). Scale bar: 200 μ m (A), 50 μ m (B/C).

and adult organism. They represent a source of immature cells with the potential to renew themselves (immortality), and give rise to cells restricted to the neuronal and glial lineage. These cells are found in specific brain regions such as the subventricular zone (SVZ), hippocampus, cortex and spinal cord [72-74], in the developing and in the adult brain [75–77]. Isolated neuronal stem cells are able to proliferate in response to different growth factors, such as basic fibroblast growth factor (bFGF) or epidermal growth factor (EGF), and to differentiate towards specific neuronal and glial phenotypes when culture conditions are altered [78] (fig. 4). Hence drugs that influence the differentiation of stem cells and neuronal precursors into a specific neurochemical phenotype therefore hold a potential for improving cell replacement techniques [79]. These cells furthermore have the ability to migrate and form functional synapses with the surrounding neurons. In both HD and PD, transplantation of stem cell and progenitors have been reported to resulted in improvement of these disease states in animal models [42, 79, 80, 81]. Recent studies have suggested that bone marrow stem cells transplanted into mice are able to migrate into specific regions of the brain, including the olfactory bulb, cortex, hippocampus and cerebellum, and differentiate into cells that appear to be neurons [82, 83]. The studies suggest that bone marrow may be an easily available source of neural cells with potential for treating neurological disorders. Another study was able to show development of neuronal phenotypes after intravenous administration of previously harvested umbilical cord blood cells [84]. First experimental studies using umbilical cord stem cells have reported promising results after ischaemic insults [85], traumatic brain injury [86], amyotrophic lateral sclerosis [87] and intracerebral haemorrhage [88]. Both the usage of autologous bone marrow and umbilical cord blood stem cells offer the advantage of tolerance by the host immune system. Moreover, they can be administered by injection into a peripheral blood vessel, therefore not requiring brain surgery.

In each of these paradigms, the main problem to be solved is the missing knowledge how transplanted cells differentiate. This includes the risk of uncontrolled growth and tumour formation in the host brain. In addition, the usage of ES cells has raised severe ethical concerns, particularly in the USA, which may hamper basic research and clinical application in the future [89]. In addition, many cell lines have recently been reported to be contaminated by murine pathogens [90]. Human stem cells cultured in the presence of animal cells or sera have also been shown to incorporate foreign sugars into their surface proteins, which may provoke immune response [91]. According to these findings, most stem cell lines have to be reestablished using environments free of animal cells and sera in respect to enable a safe application in humans.

Neuronal cell lines

Immortalised neuronal precursor cell lines kept in culture might offer an additional, theoretically unlimited source of specific cells of the neuronal or glial lineage for transplantation [92]. As an example, the clonal cell line RN33b was generated from embryonic rat raphe nucleus and transduced with the temperature sensitive mutant of the Simian Virus 40 (SV40) large T-antigen [93]. The transplanted cells can be detected by reporter genes for beta-galactosidase (beta-Gal) and green fluorescent protein (GFP). Several studies have shown that transplanted RN33b cells are able to survive in the brain and spinal cord, differentiate into specific neuronal phenotypes in a region-specific fashion [94, 95], and establish electrophysiologically active axonal projections [96] (for review see: [80]). Notably, transplantation of cells derived from cell lines contains the same obstacles as transplantation of non-transformed neural stem cells.

Neural transplantation for other neurodegenerative diseases

Amyotrophic lateral sclerosis

ALS is a rare neurodegenerative disorder characterised by the loss of the large cholinergic motor neurons in the spinal cord and degeneration of the neurons in the motor cortex, resulting in progressive paralysis and ultimately death. The underlying pathological process remains enigmatic, however, mutations in the superoxid dismutase 1 gene (SOD1) have been identified, resulting in protein misfolding and toxicity on the vulnerable cholinergic cells [97]. So far, no effective treatment options are available for patients suffering from this disease.

Basic research has proven that transplanted human ES cells can be differentiated into cholinergic

motor neurons [98]. They survive in a rodent model of ALS and show functional benefits [99]. However, it has been argued that these effects are related on the differentiation of stem cells into glial cells producing trophic support for dying motor neurons rather than by direct motor neuron replacement [100]. Preliminary stem cell transplantations in patients using autologous blood- and bone marrow-derived cells have shown the absence of major side effects, but no or only slight clinical efficacy [101, 102]. It has to be concluded that the biological issues have to be clarified before further applications on patients should be performed.

Alzheimer's disease

Alzheimer's disease (AD) is the most common neurodegenerative disorder. About 5% of the population over 65 is suffering from the disease. As the overall life expectancy is prolonged with the advancement in medical science, the incidence of AD related to aging has dramatically risen [103]. The underlying pathological mechanisms are not yet understood, although aging and genetic predisposition have been identified as major risk factors. AD starts in the mesiotemporal region which in the course of the disease shows strong alterations [104, 105]. AD also severely affects neurons in the frontal and parietal association neocortex, leading to progressive dementia. The pathological hallmarks of AD are extracellular plaques and intracellular tangles constituted of amyloid, a peptide derived from amyloid precursor protein (APP). It is not clear, however, whether these pathological accumulations are the markers or the causes of AD.

Notably, Hock and co-workers reported that immunisation of patients with aggregated amyloid β -42 resulting in the production of antibodies against amyloid β slowed cognitive decline in AD [106]. The outcome of a large randomised, placebo-controlled, double-blind trial, however, did not reveal the anticipated results and was interrupted following reports of meningoencephalitis [107]. Nevertheless, there is proof of concept of this therapy for AD [108,109]. In line with this statement, a monoclonal antibody (Bapineuzumab) designed to reduce the amount of amyloid β in the brain co-developed by the companies Elan and Wyeth is currently in phase II clinical trial and foreseen charging towards phase III trial in 2008.

Importantly, many different neurotransmitter systems, in particular the cholinergic, noradrenergic and serotonergic system, are involved in the degeneration processes in AD [110]. Current drug therapies, usually based on cholinesterase inhibitors, only relieve some of the associated symptoms of the disease, if at all [111]. Due to the extensive degeneration of multiple neuronal phenotypes in widespread brain areas, establishing a cell replacement strategy in AD is considered one of the most demanding challenges in restorative neuroscience. So far, experimental studies using embryonic cholinergic transplants in animals suffering cholinergic depletion in different brain regions such as hippocampus, septal area, basal forebrain and neocortex have shown functional benefits [112, 113]. Due to the above-mentioned

problems and the nascent stage of basic research, no clinical studies of cell replacement in AD patients have been carried out so far.

Demyelinating diseases

Multiple sclerosis (MS) is the most prominent pathology in the group of demyelinating diseases, with a prevalence of 110 cases per 100 000 inhabitants in Switzerland [114]. Nowadays, potent immunosuppressive and immunomodulating treatment regimens are available and allow the prevention of severe irreversible neurological deficits in most cases. However, there are still patients with MS resistant to conventional therapy, resulting in disabling neurological sequelae. In order to offer these patients strategies for myelin repair, numerous attempts to develop cell-based therapies have been made during the past decade. In MS, the disease process is primarily directed against oligodendrocytes and/or myelin, with neuronal structures such as axons relatively spared until late disease [115]. Therefore, reparative therapy has primarily to be focused on restoring the oligodendrocytes supplying the axons with myelin, without the need to re-establish a disrupted neuronal circuitry. Interestingly, Schwann cells, which usually provide myelin and glial support in the peripheral nerve, have shown good results in myelin repair, both in the brain [116] and particularly in the spinal cord [117]. Comparable effects have been found using olfactory ensheathing cells (OECs) in spinal cord demyelination [118]. Finally, stem cells have been shown to possess a considerable remyelinating potential [119]. A recent study using transplantation of pluripotent ES cells in an antibody/complement-induced demyelination model in the rat spinal cord demonstrated survival of the cells, differentiation both in oligodendrocytes and astrocytes and formation of new glial sheets [120].

So far, first clinical studies including implantation of dissociated rat Schwann cells [121, 122], transplantation of oligodendrocyte lineage precursor cells [123, 124], transplantation of oligodendrocyte precursors derived from cell lines [125, 126], and OECs [127] have been carried out with limited success. Critical parameters that require further investigations include the developmental stage of the oligodendrocyte precursors to be transplanted, the insufficient survival of transplanted cells, particularly in the context of a systemic immunoresponse against these cells and significant differences between human and rodent oligodendrocyte progenitor cells.

Neural transplantation for specific neurological disorders

Ischaemic stroke

Cerebral ischaemic stroke is one of the leading cause of death and disability among the eld-

erly people worldwide and has an incidence of 150 per 100'000 people per year [128]. Due to the recent advances in the diagnosis, treatment and

rehabilitation in stroke, the percentage of patients suffering severe neurological deficits has constantly dropped in the last decade, and the severity of the sequelae has been reduced. However, there are still many patients suffering from irreversible brain parenchyma defects due to stroke, with about 60% of them requiring care two weeks after the insult [128]. Hence, stroke places a heavy burden on national health care systems and demands for the development of novel effective treatment options.

Importantly, multiple different neuronal phenotypes and glial cells in different brain areas, eg, cortex, basal ganglia and thalamus, are affected in cerebral stroke. Therefore this pathology poses special conditions that impact the potential success of cell replacement therapies. Because the disease affects both gray and white matter, immature cells that have the potential to differentiate into appropriate neuronal and glial phenotypes are considered as best suitable for transplantation. So far, restorative strategies have focused on the striatum, due to clear anatomical definition, good stereotactical accessibility and less degree of white matter involvement than in cortical stroke, therefore allowing a less complex approach [129]. First clinical trials of neural transplantation in stroke have been launched in the late 1990s. One study investigated the transplantation of cells from the immortalised cell line NT2, which is derived from a human testicular germ cell tumour [130]. In preclinical studies, grafted NT2 cells, which terminally differentiate into mature neuronal phenotypes after intracerebral transplantation [131], showed significant improvements in behavioural tests after focal cerebral ischaemia [132]. A first clinical study to investigate the safety of transplantation of NT2 cells after basal ganglia infarction was started in 12 patients [133]. Up to date, no adverse effects of the procedure have been reported. Subsequently, a randomised trial with observer-blinded neurological evaluation was started in 14 patients with substantial motor deficits after basal ganglia infarction. Again, no adverse effects of the transplantation were present. However, patients with NT2 grafts showed only a trend towards a better functional outcome [134]. Another group investigated xenotransplantation of porcine foetal striatal precursor cells derived from the lateral ganglionic eminence, which previously have been reported to improve deficits in animal models of HD [64]. After focal ischaemia, intrastriatal transplantation of these cells leads to graft survival and differentiation of transplanted cells into glia and neurons with a striatal phenotype. There was evidence for synaptogenesis both with the host brain and within the graft. Four weeks after transplantation, animals showed significant behavioural improvements as compared with controls. However, no effects were found at later time points [67]. In a first clinical trial transplanting foetal porcine tissue, it was reported that two out of the five treated patients showed functional improvement after four years of clinical follow-up [135].

Basic and clinical research in neuronal transplantation for stroke is still in an early stage. Particularly, using cell replacement approaches for stroke, it remains unclear if the transplanted neurons themselves promote functional recovery or if the transplants modulate the response of the brain to ischaemic neurogenesis, synaptogenesis, angiogenesis and inflammation. Actually, there are a number of new preclinical studies of neuronal transplantation in stroke carried out (for review see: [136]). At present, transplantation of bone marrow stromal cells, which can be transdifferentiated to neuronal progenitors by exposure to specific growth factors, human umbilical blood cord stem cells, human adipose stromal cells and human neural stem cells have been investigated with varying success [137]. Interestingly, recent studies have demonstrated that intravenous infusions of umbilical cord blood can ameliorate neurological deficits associated with ischaemic brain injury in rodents, but it again remains unclear whether growth factors secreted from these cells are responsible for the induced regeneration processes rather than integration of the transplanted cells in the brain [138]. It can be assumed that inducing de novo neurogenesis may provide a more effective therapeutic strategy to promote recovery from stroke rather than transplanting exogenous cells [138].

Intracerebral haemorrhage

Spontaneous intracerebral haemorrhage (ICH) represents at least 10% of strokes in the Western population [139] and constitutes one of the most devastating forms of cerebrovascular disease. No direct treatment of the brain damage caused by ICH is currently available. Like in ischaemic stroke, progress in experimental neurobiology gives hope that new brain repair strategies using stem cell transplantation could be also advantageously employed against this disease state. So far, only two preclinical studies of neuronal tissue grafting in ICH have been reported. One study did not report any functional improvement after transplantation of foetal brain tissue [140]. Another report showed integration of intravenously administrated human neuronal stem cells in the damaged brain and functional recovery in rats undergoing ICH [141]. Intense research efforts are currently being undertaken to establish a more reproducible animal model of ICH and to improve neuronal transplantation in this context [142, 143].

Trauma to the brain and spinal cord

Traumatic injury to the brain and spinal cord usually results in irreversible neurological deficits that can only partially be compensated by activation of other neural networks, which functionally replace the damaged neurons. During the last two decades, advances in both basic and clinical re-

search have markedly improved our understanding of the cellular and molecular mechanisms in brain and spinal cord trauma [144–146]. In addition, sophisticated cerebral and spinal lesion models have been developed that allow the study of traumatic events and adequate treatment strategies *in vivo* [147].

In this context, it is important to note that neutralising the inhibitory effects of NOGO-A, which is a potent inhibitor of neurite outgrowth in the adult CNS, resulted in enhanced fibre growth and functional recovery in rodent and primate models of spinal cord injury [148]. The first clinical trial assessing the therapeutic potential of antibodies to NOGO-A is currently in progress [149].

Neural transplantation has turned out to be an important therapeutic option at least in experimental studies [145, 150, 151]. Various cell types have been investigated because of their considerable potential for promoting axonal regeneration. Schwann cells and OECs are glial elements that have both the capacity to stimulate fibre outgrowth as well as to remyelinate the tissue. Studies have shown a potential benefit of the transplantation of Schwann cells [152] and OECs [153] in models of complete and incomplete transection as well as contusion of the spinal cord. While there are no published studies of Schwann cells transplantation in humans so far, first clinical trials have shown that transplantation of autologous OECs is feasible, however there were no significant effects on neurological outcome [154]. Bone marrow stromal cells, which show stem cell-like attributes and pluripotency [155] and can both

differentiate in neurons and glia [82, 83], have been found to promote functional recovery in spinal cord injury models. In addition, first studies have proven that human stem cells can differentiate into neurons and glia and promote locomotor recovery in spinal cord-injured mice [156].

Epilepsy

Medically intractable epilepsy, which means that the seizures are resistant to treatment with a combination of different anticonvulsive drugs, is usually treated surgically with ablative procedures to remove epileptogenic foci, eg, by performing selective amygdala-hippocampectomy or temporomesial lobectomy [157]. The precise underlying pathology of idiopathic epileptic seizures remains enigmatic in most cases, however, a imbalance between inhibitory and excitatory neurotransmitters in favour of the latter is usually considered to be present [158]. Cell-based therapies, such as the transplantation of inhibitory neurons like GABA-ergic cells, might therefore be useful for correcting the imbalance and thus preventing or alleviating seizures without the need for irreversible ablative surgery. First preclinical trials with transplantation of genetically engineered, GABA-releasing precursors [159] or human neural stem cells [160] in rats suffering from seizures have shown to decrease neuronal excitability and to raise the seizure threshold. With the advance of research, transplantation of inhibitory neurons therefore might become a novel, less aggressive treatment option for patients suffering from intractable epileptic seizures.

Strategies to improve cell replacement approaches

Neurotrophic Factors

Nerve growth factors or neurotrophic factors are proteins produced by glial cells and neurons during the development of the CNS that play important roles in controlling and coordinating neuronal growth, survival and differentiation. Many of them have been identified promoting survival and/or differentiation of specific neuronal subpopulations in vitro and in vivo (table 1). Among the most effective are the neurotrophin family members nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), neurotrophin-4/5 (NT-4/5) and the glial cell line-derived neurotrophic factor (GDNF) family ligands (GFLs) [161, 162]. The GFLs belong to a distant branch of the transforming growth factor-β superfamily [163, 164], which comprises GDNF [165], neurturin (NTN) [166], Persephin (PSP) [167] and artemin (ARTN) [168] and have been described as being potent survival factors for midbrain dopamine neurons, motoneurons, noradrenergic neurons, and sympathetic, parasympathetic and

sensory neurons [164, 169–173]. Neuroprotective effects of GDNF have been reported in experimental models of PD [174]. In addition, GDNF has been found to induce the expression of the dopaminergic marker tyrosine hydroxylase (TH) in late developmental stages of cultured neural progenitor cells and may therefore provide a robust tool to interfere with final cell fate specification of neural precursor cells [175]. A very recent study by Lindholm and co-workers described a novel neurotrophic factor for dopamine neurons: neurotrophic conserved dopamine (CDNF), which was at least as efficient as GDNF in their experimental settings, suggesting that CDNF might be beneficial for the treatment of PD [176].

Preliminary clinical trials with neurotrophic factors have been carried out in PD, HD, ALS and AD (table 2). Notably, the first attempts to apply factors failed to demonstrate any significant clinical benefits, despite positive preclinical data [177]. It is assumed that this was due primarily because

Table 1
Growth factors
with neurotrophic
activity*.

Baloh et al., 1998 [168]
Chen et al., 2004 [215] Harvey et al., 2005 [216]
Barde et al., 1987 [217] Leibrock et al., 1989 [218]
Lin et al., 1989 [219] Stockli et al., 1989 [220]
Lindholm et al., 2007 [176]
Morrison et al., 1988 [221]
Gospodarowicz et al., 1986 [222] Grothe and Timmer, 2007 [223]
Lin et al., 1993 [164] Beck et al., 1995 [169]
Aizenman et al., 1986 [224] Baskin et al., 1987 [225]
Spranger et al., 1990 [226] Kamegai et al., 1990 [227] Hama et al., 1990 [228]
Whittemore and Seiger, 1987 [229] Thoenen et al., 1987 [230] Hefti, 1986 [231]
Widenfalk et al., 1997 [232] Horger et al., 1998 [233] Akerud et al., 1999 [234]
Hohn et al., 1990 [235] Maisonpierre et al., 1990 [236] Rosenthal et al., 1990 [237]
Hynes et al., 1994 [238] Widmer and Hefti, 1994 [239]
Milbrandt et al., 1998 [167] Zihlmann et al., 2005 [240]
Derynck, 1988 [241] Code et al., 1987 [242]
Ren and Flanders, 1996 [243]

^{*} This table lists only a selection of growth factors and correspondingly only a selection of references.

Table 2Neurotrophic factors in clinical trials.

Neurotrophic factor	Disease	References
BDNF	ALS	Bensimon et al., 1999 [244]
		Kalra et al., 2003 [245]
CNTF	ALS / HD	Bloch et al., 2004 [186]
IGF-1	ALS	Borasio et al., 1998 [246]
		Lai et al., 1997 [247]
GDNF	PD	Nutt et al., 2003 [248]
		Lang et al., 2006 [249]
NGF	AD	Tuszynski et al., 2005 [187] CERE-110: Adeno-associated virus (AAV)-based delivery of β-NGF in subjects with mild to moderate Alzheimer's disease. Available at http://clinicaltrials.gov/ct/show/NCT 00087789?order=1.
NTN	PD	Safety of CERE-120 (AAV2-NTN) in subjects with idiopathic Parkinson's disease. Available at http://clinicaltrials.gov/ct/show/NCT00252850?order=1.

of the poor blood-brain barrier permeability of these proteins [178]. Recent developments of new delivery methods, eg, adeno-associated virus (AAV) mediated transfer [179], have revived the interest in these potential nervous system protein therapeutics [177]. A first small open-label clinical

trial provided evidence that direct infusion of GDNF into the putamen of PD patients resulted in improvement of motor scores [180]. The benefits of intracerebral delivery of GDNF, however, could not be substantiated in a recent double-blinded placebo-controlled study. Moreover,

safety concerns emerged [181]. One elegant method for delivering neurotrophic factors to the brain is the use of encapsulated cell lines engineered to produce neurotrophic factors [182-184]. Its major advantage is a continuous, almost unlimited supply of these molecules. Encapsulation in a porous polymer membrane also provides a high safety level in regard to tumour formation while allowing exchange of metabolites. In addition, it separates the xenogenic cells from the host immune system [182]. So we have previously shown that implantation of encapsulated genetically engineered fibroblasts producing GDNF resulted in better survival and host integration of transplanted dopaminergic cells in the rat [185]. A phase I study that evaluated intracerebral administration of CNTF in subjects with HD, using a device formed by a semipermeable membrane encapsulating a cell line engineered to synthesise CNTF, demonstrated the safety, feasibility, and tolerability of this gene therapy approach [186].

A new promising window of neurotrophic factor delivery to the brain has been opened by the study of Tuszynski and colleagues [187]. In a phase I human clinical trial, autologous genetically modified fibroblasts releasing NGF were transplanted in eight patients with early stage AD. The preliminary results indicate that ex vivo NGF gene delivery is safe and seems to provide trophic support to degenerating cholinergic neurons [188]. Neurotrophic factor delivery to the brain may also be achieved by means of transplanting native or modified stem cells. So it has been described that transplantation of mesenchymal stem cells results in improved functional outcome in animal models of neurological disorders. These cells have, however, generally only a limited ability to differentiate into neurons. A recent study now showed that transplanted human adult mesenchymal stem cells released neurotrophins, which offers the possibility that co-transplantation of such cells with tissue grafts results in improved functional outcome [189].

Neuroprotective drugs

Survival of neuronal cells exposed to oxidative and metabolic stress can be improved by antioxidants, which act as free radical scavengers. One of these substances is the lazaroid tirilazad mesylate, which inhibits lipid peroxidation and can be used for pretreatment of grafts and/or can be postoperatively administrated to the transplanted patients. So Brundin and colleagues were able to show improved survival of rat and human dopaminergic cells *in vitro* and *in vivo* [190], however, only discrete effects could be demonstrated in patients [191].

Compounds that inhibit neuronal apoptosis [192] have been thoroughly investigated as potential drugs for improving neuronal cell survival. Such different substances as minocycline, a tetracyclin antibiotic [193], cytokines like erythropoietin [194] and granulocyte-colony stimulating

factor [195], specific inhibitors of enzymes involved in apoptosis like the caspase inhibitor zVAD [196] and many others have been identified to possess anti-apoptotic properties on neuronal cells. However, caution must be exercised in view of possible side effects to successfully transfer therapeutic compounds to the clinic. Due to their long-established safety, minocycline and erythropoietin are the most appealing candidates for clinical trials in patients.

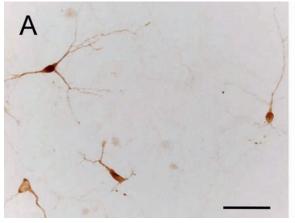
Creatine

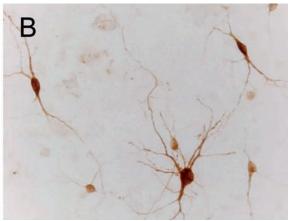
The specific functional properties of neuronal tissue make high demands on cellular energy resources. Rapid changes in ATP demands are occurring during physiological function of neurons, while cellular energy reserves are very small [197]. Widely distributed cellular processes and sites of very high energy consumption localised at remote locations from the cell body, such as synapses, require mechanisms to facilitate energy transfer within the cell. The phosphocreatine/creatine kinase (PCr/CK) system has been described as playing a key role for maintaining the cellular energy homeostasis in neurons [198, 199]. Due to its function as a temporal ATP buffer and a carrier for high energy phosphates from sites of ATP production to sites of ATP consumption, it prevents marked changes in the concentrations of ADP and ATP, which has been postulated to be crucial in neuronal cells [200].

Despite intense research activities, the aetiology of neuronal death in neurodegenerative diseases still remains widely unclear. However, there are a number of similarities in the fundamental biochemical processes involved in the pathogenesis and progression of these otherwise different pathological states. The concepts of energy depletion, oxidative stress, excitotoxicity, and mitochondrial dysfunction have been implicated in HD, PD, ALS, and several hereditary mitochondrial neuromuscular disorders [201, 202, 203]. Although these processes may be directly or indirectly involved in the pathogenesis of a given disease, they converge in final common pathways of either necrosis or apoptosis. Substantial evidence indicates that energy dysfunction plays either a primary or secondary role in cell death in neurodegenerative and neuromuscular disorders, and even in normal aging. Agents that counteract these defects may therefore be useful as novel therapeutic strategies. Therapeutic supplementation of creatine has been reported to improve cellular ATP resources, inhibit apoptosis and therefore exert neuroprotective properties [204, 205]. In respect to neurorestorative paradigms, we have shown that creatine exposure protected dopaminergic (Fig. 5) and GABA-ergic neurons in experimental in vitro models of PD [206] and HD and induced differentiation of neuronal precursors towards the GABA-ergic phenotype [207]. Furthermore, creatine provided neuroprotection on dopaminergic cells during storage in an organ-

Figure 5

Neuroprotective effects of creatine exposure on densities of dopaminergic neuronal precursors immunostained for the catecholaminergic marker tyrosine hydroxylase in mesencephalic cultures. Cells were grown for 7 days in vitro in absence (A) or presence (B) of creatine [5 mM] in the culture medium. Note the promotion of morphological differentiation of the neurons after exposure to creatine. Scale bar: 50 μm.





otypic tissue culture system [208]. Creatine seems therefore to be one of the most promising neuroprotective substances. In line with this notion, a number of clinical trials were launched or are planned for PD, HD, and ALS.

Endogenous regeneration

The presence of endogenous stem cells and persistent neuronal production in specific regions of the adult human brain [209] suggests a previously unrecognised capacity for regeneration in the CNS. In particular two brain regions, namely the subgranular zone of the hippocampal dentate gyrus, and the forebrain subventricular zones of

the lateral ventricles, have been identified as containing stem cells giving rise to neurons and glia [210]. Using 5-bromo-2'-deoxyuridine 5'-monophosphate labeling, it has been shown that experimental stroke [211] and also intermittent hypoxia [212] resulted in stimulation of neurogenesis in the adult brain. It may be speculated that activation or recruitment of these endogenous stem cells, eg, by administration of appropriate drugs and growth factors, might offer fascinating new treatment options for various neurological disease states and also means to prevent brain atrophy during normal aging [213, 214].

Future directions

The findings reported from preclinical and clinical studies demonstrate that cell replacement strategies have the potential to become novel and effective therapeutic approaches for repair of the brain and spinal cord after a variety of degenerative, vascular and traumatic lesions. However, due to the limitations described above, these techniques still remain experimental and can only be applied to small groups of patients in the context of clinical studies, if at all. There are mainly three limitations that have to be overcome in the next years if these strategies should achieve clinical significance. First, a better understanding of the treated disorders is required to further develop and improve regenerative strategies and to achieve a better survival and functional integration of the grafted cells under pathological conditions. Second, the shortage of donor tissue and ethical issues demand an improvement in transplantation efficacy and forced research for alternative tissue sources, like autologous stem cells and neuronal cell lines. Third, possible side effects, eg, dyskinesias after cell transplantation in PD, have to be elucidated and prevented by adequate means.

In our opinion, given the complexity of the field, only a multimodal approach that includes the development of reliable tissue sources, advanced techniques for tissue storage and neural transplantation, effective drugs such as growth factors to induce differentiation towards the required specific neuronal phenotype, and neuroprotective agents to improve sustained graft survival will ensure a successful clinical application of cell replacement strategies. Another potential goal of restorative neuroscience might consist of harvesting a patient's own neural stem cells, in vitro expansion and induction of differentiation to the required neuronal phenotypes and reimplantation into the damaged structures of the CNS. If we manage to gain further knowledge how stem cells are activated, differentiate, migrate and establish structural and function interaction with the surrounding cells of the host brain and the extracellular matrix, it may be possible to achieve neuronal repair by activating the endogenous stem cells without the need for any transplantation [138]. This could be accomplished by application of appropriate growth factors and other molecules that interfere with neuronal proliferation and differentiation, for example. Furthermore it may be speculated that activation of endogenous stem cells might prevent atrophy and functional loss in the aging brain, an issue that will become more and more important in the context of our increasing life expectancy.

Conclusions

The field of restorative neuroscience and regeneration in the CNS represents a research area where increasing efforts hope to provide therapeutic options for pathologies for which currently no effective treatment strategies are available. However, much more research is needed to characterise and understand the biology of different types of cells intended for cell replacement therapies both *in vitro* and *in vivo*. Existing clinical data suggest that transplantation is technically feasible and can be carried out safely, but the data on functional outcome and long-term efficiency is still rather preliminary. Taken together, we propose that cell replacement strategies in combination

with appropriate growth factors and / or neuroprotective drugs hold the potential to be effective treatment options for a variety of neuropathological conditions.

Correspondence:
Hans R. Widmer, Ph.D.
Department of Neurosurgery
University of Berne
Inselspital
Freiburgstrasse 10
CH-3010 Berne
Switzerland
E-Mail: hanswi@insel.ch

References

- 1 Kauer JA, Malenka RC. Synaptic plasticity and addiction. Nat Rev Neurosci. 2007;8:844-58.
- 2 Massey PV, Bashir ZI. Long-term depression: multiple forms and implications for brain function. Trends Neurosci. 2007;30: 176-84.
- 3 Nishiyama H, Fukaya M, Watanabe M, Linden DJ. Axonal motility and its modulation by activity are branch-type specific in the intact adult cerebellum. Neuron. 2007;56:472-87.
- 4 Prolla TA, Mattson MP. Molecular mechanisms of brain aging and neurodegenerative disorders: lessons from dietary restriction. Trends Neurosci. 2001;24:S21-S31.
- 5 McOmish CE, Hannan AJ. Environmentics: exploring gene environment interactions to identify therapeutic targets for brain disorders. Expert Opin Ther Targets. 2007;11:899-913.
- 6 Cheung ME, Broman SH. Adaptive learning: interventions for verbal and motor deficits. Neurorehabil Neural Repair. 2000; 14:159-69.
- 7 Tyc F, Boyadjian A. Cortical plasticity and motor activity studied with transcranial magnetic stimulation. Rev Neurosci. 2006;17:460-95
- 8 Gerloff C, Bushara K, Sailer A, Wassermann EM, Chen R, Matsuoka T, et al. Multimodal imaging of brain reorganization in motor areas of the contralesional hemisphere of well recovered patients after capsular stroke. Brain. 2006;129:791-808.
- 9 Mountz JM. Nuclear medicine in the rehabilitative treatment evaluation in stroke recovery. Role of diaschisis resolution and cerebral reorganization. Eura Medicophys. 2007;43:221-39.
- 10 Lindvall O, Rehncrona S, Brundin P, Gustavii B, Astedt B, Widner H, et al. Neural transplantation in Parkinson's disease: the Swedish experience. Prog Brain Res. 1990;82:729-34.
- 11 Kopyov OV, Jacques S, Lieberman A, Duma CM, Eagle KS. Safety of intrastriatal neurotransplantation for Huntington's disease patients. Exp Neurol. 1998;149:97-108.
- 12 Marttila RJ, Rinne UK. Clues from epidemiology of Parkinson's disease. Adv Neurol. 1987;45:285-8.
- 13 Fahn S, Przedborski S. Parkinsonism. In: Rowland LP (Ed.): Merritt's Neurology. New York: Lippincott Williams and Wilkins; 2000.
- 14 Lang AE, Lozano AM. Parkinson's disease. First of two parts. N Engl J Med. 1998;339:1044-53.
- 15 Lang AE, Lozano AM. Parkinson's disease. Second of two parts. N Engl J Med. 1998;339:1130-43.
- 16 Schapira AH, Cooper JM, Dexter D, Clark JB, Jenner P, Marsden CD. Mitochondrial complex I deficiency in Parkinson's disease. J Neurochem. 1990;54:823-7.
- 17 Swerdlow RH, Parks JK, Davis JN, Cassarino DS, Trimmer PA, Currie LJ, et al. Matrilineal inheritance of complex I dysfunction in a multigenerational Parkinson's disease family. Ann Neurol. 1998;44:873-81.
- 18 Nikkhah G, Cunningham MG, Cenci MA, McKay RD, Bjorklund A. Dopaminergic microtransplants into the substantia nigra of neonatal rats with bilateral 6-OHDA lesions. I. Evidence for anatomical reconstruction of the nigrostriatal pathway. J Neurosci. 1995;15:3548-61.

- 19 Arbuthnott G, Dunnett S, MacLeod N. Electrophysiological properties of single units in dopamine-rich mesencephalic transplants in rat brain. Neurosci Lett. 1985;57:205-10.
- 20 Brundin P, Isacson O, Gage FH, Prochiantz A, Bjorklund A. The rotating 6-hydroxydopamine-lesioned mouse as a model for assessing functional effects of neuronal grafting. Brain Res. 1986;366:346-9.
- 21 Brundin P, Bjorklund A. Survival, growth and function of dopaminergic neurons grafted to the brain. Prog Brain Res. 1987;71:293-308.
- 22 Brundin P, Karlsson J, Emgard M, Schierle GS, Hansson O, Petersen A, et al. Improving the survival of grafted dopaminergic neurons: a review over current approaches. Cell Transplant. 2000;9:179-95.
- 23 Hauser RA, Freeman TB, Snow BJ, Nauert M, Gauger L, Kordower JH, et al. Long-term evaluation of bilateral fetal nigral transplantation in Parkinson disease. Arch Neurol. 1999;56: 179-87
- 24 Kordower JH, Freeman TB, Snow BJ, Vingerhoets FJ, Mufson EJ, Sanberg PR, et al. Neuropathological evidence of graft survival and striatal reinnervation after the transplantation of fetal mesencephalic tissue in a patient with Parkinson's disease. N Engl J Med. 1995;332:1118-24.
- 25 Kordower JH, Rosenstein JM, Collier TJ, Burke MA, Chen EY, Li JM, et al. Functional fetal nigral grafts in a patient with Parkinson's disease: chemoanatomic, ultrastructural, and metabolic studies. J Comp Neurol. 1996;370:203-30.
- 26 Kordower JH, Freeman TB, Chen EY, Mufson EJ, Sanberg PR, Hauser RA, et al. Fetal nigral grafts survive and mediate clinical benefit in a patient with Parkinson's disease. Mov Disord. 1998;13:383-93.
- 27 Wenning GK, Odin P, Morrish P, Rehncrona S, Widner H, Brundin P, et al. Short- and long-term survival and function of unilateral intrastriatal dopaminergic grafts in Parkinson's disease. Ann Neurol. 1997;42:95-107.
- 28 Piccini P, Brooks DJ, Bjorklund A, Gunn RN, Grasby PM, Rimoldi O, et al. Dopamine release from nigral transplants visualized in vivo in a Parkinson's patient [see comments]. Nat Neurosci. 1999;2:1137-40.
- 29 Lindvall O. Update on fetal transplantation: the Swedish experience. Mov Disord. 1998;13(Suppl 1):83-7.
- 30 Barker RA, Dunnett SB, Faissner A, Fawcett JW. The time course of loss of dopaminergic neurons and the gliotic reaction surrounding grafts of embryonic mesencephalon to the striatum. Exp Neurol. 1996;141:79-93.
- 31 Zawada WM, Zastrow DJ, Clarkson ED, Adams FS, Bell KP, Freed CR. Growth factors improve immediate survival of embryonic dopamine neurons after transplantation into rats. Brain Res. 1998;786:96-103.
- 32 Bjorklund A. Neurobiology. Better cells for brain repair. Nature. 1993;362:414-5.

33 Maries E, Kordower JH, Chu Y, Collier TJ, Sortwell CE, Olaru E, et al. Focal not widespread grafts induce novel dyskinetic behavior in parkinsonian rats. Neurobiol Dis. 2006;21: 165-80.

- 34 Freed CR, Greene PE, Breeze RE, Tsai WY, DuMouchel W, Kao R, et al. Transplantation of embryonic dopamine neurons for severe Parkinson's disease. N Engl J Med. 2001;344(10): 710.0
- 35 Olanow CW, Goetz CG, Kordower JH, Stoessl AJ, Sossi V, Brin MF, et al. A double-blind controlled trial of bilateral fetal nigral transplantation in Parkinson's disease. Ann Neurol. 2003;54:403-14.
- 36 Abbott A. Trials offer way forward for Parkinson's. Nature. 2001;410:401.
- 37 Fischbach GD, McKhann GM. Cell therapy for Parkinson's disease. N Engl J Med. 2001;344:763-5.
- 38 Hurelbrink CB, Barker RA. The potential of GDNF as a treatment for Parkinson's disease. Exp Neurol. 2004;185:1-6.
- 39 Freed CR, Breeze RE, Fahn S, Eidelberg D. Preoperative response to levodopa is the best predictor of transplant outcome. Ann Neurol. 2004;55:896-7.
- 40 Redmond DE Jr. Cellular replacement therapy for Parkinson's disease where we are today? Neuroscientist. 2002;8:457-88.
- 41 Winkler C, Kirik D, Bjorklund A. Cell transplantation in Parkinson's disease: how can we make it work? Trends Neurosci. 2005;28:86-92.
- 42 Bjorklund A, Lindvall O. Parkinson disease gene therapy moves toward the clinic. Nat Med. 2000;6:1207-8.
- 43 Goldman SA, Windrem MS. Cell replacement therapy in neurological disease. Philos Trans R Soc Lond B Biol Sci. 2006; 361:1463-75
- 44 Sanberg PR, Coyle JT. Scientific approaches to Huntington's disease. CRC Crit Rev Clin Neurobiol. 1984;1:1-44.
- 45 Hefter H, Homberg V, Lange HW, Freund HJ. Impairment of rapid movement in Huntington's disease. Brain. 1987;110(Pt 3):585-612
- 46 Harper PS. Huntington's Disease. In: Harper PS (Ed.): Major Problems in Neurology. London: Saunders; 1996.
- 47 Hayden MR, Hewitt J, Stoessl AJ, Clark C, Ammann W, Martin WR. The combined use of positron emission tomography and DNA polymorphisms for preclinical detection of Huntington's disease. Neurology. 1987;37:1441-7.
- 48 Gusella JF, Wexler NS, Conneally PM, Naylor SL, Anderson MA, Tanzi RE, et al. A polymorphic DNA marker genetically linked to Huntington's disease. Nature. 1983;306:234-8.
- 49 The Huntington's Disease Collaborative Research Group. A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes. Cell. 1993;72:971-83.
- 50 Browne SE, Bowling AC, MacGarvey U, Baik MJ, Berger SC, Muqit MM, et al. Oxidative damage and metabolic dysfunction in Huntington's disease: selective vulnerability of the basal ganglia. Ann Neurol. 1997;41:646-53.
- 51 Deckel AW, Robinson RG, Coyle JT, Sanberg PR. Reversal of long-term locomotor abnormalities in the kainic acid model of Huntington's disease by day 18 fetal striatal implants. Eur J Pharmacol. 1983;93:287-8.
- 52 Dunnett SB, Hernandez TD, Summerfield A, Jones GH, Arbuthnott G. Graft-derived recovery from 6-OHDA lesions: specificity of ventral mesencephalic graft tissues. Exp Brain Res. 1988;71:411-24.
- 53 Isacson O, Dunnett SB, Bjorklund A. Graft-induced behavioral recovery in an animal model of Huntington disease. Proc Natl Acad Sci. U S A 1986;83:2728-32.
- 54 Bachoud L, Bourdet C, Brugieres P, Nguyen JP, Grandmougin T, Haddad B et al. Safety and tolerability assessment of intrastriatal neural allografts in five patients with Huntington's disease. Exp Neurol. 2000;161:194-202.
- 55 Freeman TB, Cicchetti F, Hauser RA, Deacon TW, Li XJ, Hersch SM, et al. Transplanted fetal striatum in Huntington's disease: phenotypic development and lack of pathology. Proc Natl Acad Sci. U S A 2000;97:13877-82.
- 56 Hauser RA, Furtado S, Cimino CR, Delgado H, Eichler S, Schwartz S, et al. Bilateral human fetal striatal transplantation in Huntington's disease. Neurology. 2002;58:687-95.
- 57 Ramaswamy S, Shannon KM, Kordower JH. Huntington's disease: pathological mechanisms and therapeutic strategies. Cell Transplant. 2007;16:301-12.
- 58 Keene CD, Sonnen JA, Swanson PD, Kopyov O, Leverenz JB, Bird TD, et al. Neural transplantation in Huntington disease: long-term grafts in two patients. Neurology. 2007;68:2093-8.

59 Frank S, Biglan K. Long-term fetal cell transplant in Huntington disease: stayin' alive. Neurology. 2007;68:2055-6.

- 60 Gaura V, Bachoud-Levi AC, Ribeiro MJ, Nguyen JP, Frouin V, Baudic S, et al. Striatal neural grafting improves cortical metabolism in Huntington's disease patients. Brain. 2004;127:65-72
- 61 Bachoud-Levi AC, Gaura V, Brugieres P, Lefaucheur JP, Boisse MF, Maison P, et al. Effect of fetal neural transplants in patients with Huntington's disease 6 years after surgery: a long-term follow-up study. Lancet Neurol. 2006;5:303-9.
- 62 Dunnett SB, Rosser AE. Cell transplantation for Huntington's disease Should we continue? Brain Res Bull. 2007;72:132-47.
- 63 Dwyer KM, Cowan PJ, d'Apice AJ. Xenotransplantation: Past achievements and future promise. Heart Lung Circ. 2002;11: 32-41.
- 64 Fink JS, Schumacher JM, Ellias SL, Palmer EP, Saint-Hilaire M, Shannon K, et al. Porcine xenografts in Parkinson's disease and Huntington's disease patients: preliminary results. Cell Transplant. 2000;9:273-8.
- 65 Deacon T, Schumacher J, Dinsmore J, Thomas C, Palmer P, Kott S, et al. Histological evidence of fetal pig neural cell survival after transplantation into a patient with Parkinson's disease. Nat Med. 1997;3:350-3.
- 66 Schumacher JM, Ellias SA, Palmer EP, Kott HS, Dinsmore J, Dempsey PK, et al. Transplantation of embryonic porcine mesencephalic tissue in patients with PD. Neurology. 2000;54: 1042-50.
- 67 Dinsmore JH, Manhart C, Raineri R, Jacoby DB, Moore A. No evidence for infection of human cells with porcine endogenous retrovirus (PERV) after exposure to porcine fetal neuronal cells. Transplantation. 2000;70:1382-9.
- 68 Paradis K, Langford G, Long Z, Heneine W, Sandstrom P, Switzer WM, et al. Search for cross-species transmission of porcine endogenous retrovirus in patients treated with living pig tissue. The XEN 111 Study Group. Science. 1999;285: 1236-41.
- 69 Phelps CJ, Koike C, Vaught TD, Boone J, Wells KD, Chen SH, et al. Production of alpha 1,3-galactosyltransferase-deficient pigs. Science. 2003;299:411-4.
- 70 Evans MJ, Kaufman MH. Establishment in culture of pluripotential cells from mouse embryos. Nature. 1981;292:154-6.
- 71 Nagy A, Gocza E, Diaz EM, Prideaux VR, Ivanyi E, Markkula M, et al. Embryonic stem cells alone are able to support fetal development in the mouse. Development. 1990;110:815-21.
- 72 Svendsen CN, Caldwell MA, Shen J, ter Borg MG, Rosser AE, Tyers P, et al. Long-term survival of human central nervous system progenitor cells transplanted into a rat model of Parkinson's disease. Exp Neurol. 1997;148:135-46.
- 73 Svendsen CN, ter Borg MG, Armstrong RJ, Rosser AE, Chandran S, Ostenfeld T, et al. A new method for the rapid and long term growth of human neural precursor cells. J Neurosci Methods. 1998;85:141-52.
- 74 Uchida N, Buck DW, He D, Reitsma MJ, Masek M, Phan TV, et al. Direct isolation of human central nervous system stem cells. Proc Natl Acad Sci. U S A 2000;97:14720-5.
- 75 Temple S, varez-Buylla A. Stem cells in the adult mammalian central nervous system. Curr Opin Neurobiol. 1999;9:135-41.
- 76 Nunes MC, Roy NS, Keyoung HM, Goodman RR, McKhann G, Jiang L, et al. Identification and isolation of multipotential neural progenitor cells from the subcortical white matter of the adult human brain. Nat Med. 2003;9:439-47.
- 77 Svendsen CN, Caldwell MA, Ostenfeld T. Human neural stem cells: isolation, expansion and transplantation. Brain Pathol. 1999;9:499-513.
- 78 Keyoung HM, Roy NS, Benraiss A, Louissaint A Jr, Suzuki A, Hashimoto M, et al. High-yield selection and extraction of two promoter-defined phenotypes of neural stem cells from the fetal human brain. Nat Biotechnol. 2001;19:843-50.
- 79 Sonntag KC, Simantov R, Isacson O. Stem cells may reshape the prospect of Parkinson's disease therapy. Brain Res Mol Brain Res. 2005;134:34-51.
- 80 Dunnett SB, Rosser AE. Stem cell transplantation for Huntington's disease. Exp Neurol. 2007;203:279-92.
- 81 Bjorklund A, Dunnett SB, Brundin P, Stoessl AJ, Freed CR, Breeze RE, et al. Neural transplantation for the treatment of Parkinson's disease. Lancet Neurol. 2003;2:437-45.
- 82 Mezey E, Chandross KJ, Harta G, Maki RA, McKercher SR. Turning blood into brain: cells bearing neuronal antigens generated in vivo from bone marrow. Science. 2000;290:1779-82.
- 83 Brazelton TR, Rossi FM, Keshet GI, Blau HM. From marrow to brain: expression of neuronal phenotypes in adult mice. Science. 2000;290:1775-9.

- 84 Sanchez-Ramos JR, Song S, Kamath SG, Zigova T, Willing A, Cardozo-Pelaez F, et al. Expression of neural markers in human umbilical cord blood. Exp Neurol. 2001;171:109-15.
- 85 Chen J, Sanberg PR, Li Y, Wang L, Lu M, Willing AE, et al. Intravenous administration of human umbilical cord blood reduces behavioral deficits after stroke in rats. Stroke. 2001; 32:2682-8.
- 86 Lu D, Li Y, Wang L, Chen J, Mahmood A, Chopp M. Intraarterial administration of marrow stromal cells in a rat model of traumatic brain injury. J Neurotrauma. 2001;18: 813-9
- 87 Garbuzova-Davis S, Willing AE, Zigova T, Saporta S, Justen EB, Lane JC, et al. Intravenous administration of human umbilical cord blood cells in a mouse model of amyotrophic lateral sclerosis: distribution, migration, and differentiation. J Hematother Stem Cell Res. 2003;12:255-70.
- 88 Nan Z, Grande A, Sanberg CD, Sanberg PR, Low WC. Infusion of human umbilical cord blood ameliorates neurologic deficits in rats with hemorrhagic brain injury. Ann N Y Acad Sci. 2005;1049:84-96.
- 89 Vats A, Bielby RC, Tolley NS, Nerem R, Polak JM. Stem cells. Lancet. 2005;366:592-602.
- 90 Schiff LJ. Review: production, characterization, and testing of banked mammalian cell substrates used to produce biological products. In Vitro Cell Dev Biol Anim. 2005;41:65-70.
- 91 Martin MJ, Muotri A, Gage F, Varki A. Human embryonic stem cells express an immunogenic nonhuman sialic acid. Nat Med. 2005;11:228-32.
- 92 Cacci E, Villa A, Parmar M, Cavallaro M, Mandahl N, Lindvall O, et al. Generation of human cortical neurons from a new immortal fetal neural stem cell line. Exp Cell Res. 2007; 313:588-601.
- 93 Whittemore SR, White LA. Target regulation of neuronal differentiation in a temperature-sensitive cell line derived from medullary raphe. Brain Res. 1993;615:27-40.
- 94 Onifer SM, Whittemore SR, Holets VR. Variable morphological differentiation of a raphe-derived neuronal cell line following transplantation into the adult rat CNS. Exp Neurol. 1993;122:130-42.
- 95 Shihabuddin LS, Hertz JA, Holets VR, Whittemore SR. The adult CNS retains the potential to direct region-specific differentiation of a transplanted neuronal precursor cell line. J Neurosci. 1995;15:6666-78.
- 96 Englund U, Bjorklund A, Wictorin K, Lindvall O, Kokaia M. Grafted neural stem cells develop into functional pyramidal neurons and integrate into host cortical circuitry. Proc Natl Acad Sci. U S A 2002;99:17089-94.
- 97 Hand CK, Rouleau GA. Familial amyotrophic lateral sclerosis. Muscle Nerve. 2002;25:135-59.
- 98 Wichterle H, Lieberam I, Porter JA, Jessell TM. Directed differentiation of embryonic stem cells into motor neurons. Cell. 2002;110:385-97.
- 99 Kerr DA, Llado J, Shamblott MJ, Maragakis NJ, Irani DN, Crawford TO, et al. Human embryonic germ cell derivatives facilitate motor recovery of rats with diffuse motor neuron injury. J Neurosci. 2003;23:5131-40.
- 100 Svendsen CN, Langston JW. Stem cells for Parkinson disease and ALS: replacement or protection? Nat Med. 2004;10:224-5.
- 101 Janson CG, Ramesh TM, During MJ, Leone P, Heywood J. Human intrathecal transplantation of peripheral blood stem cells in amyotrophic lateral sclerosis. J Hematother Stem Cell Res. 2001;10:913-5.
- 102 Mazzini L, Fagioli F, Boccaletti R, Mareschi K, Oliveri G, Olivieri C, et al. Stem cell therapy in amyotrophic lateral sclerosis: a methodological approach in humans. Amyotroph Lateral Scler Other Motor Neuron Disord. 2003;4:158-61.
- 103 Desai AK, Grossberg GT. Diagnosis and treatment of Alzheimer's disease. Neurology. 2005;64:S34-S39.
- 104 Braak H, Braak E. Staging of Alzheimer's disease-related neurofibrillary changes. Neurobiol Aging. 1995;16:271-8.
- 105 Henry-Feugeas MC. MRI of the 'Alzheimer syndrome'. J Neuroradiol. 2007;34:220-7.
- 106 Hock C, Konietzko U, Streffer JR, Tracy J, Signorell A, Muller-Tillmanns B, et al. Antibodies against beta-amyloid slow cognitive decline in Alzheimer's disease. Neuron. 2003; 38:547-54.
- 107 Gilman S, Koller M, Black RS, Jenkins L, Griffith SG, Fox NC, et al. Clinical effects of Abeta immunization (AN1792) in patients with AD in an interrupted trial. Neurology. 2005; 64:1553-62.
- 108 Gandy S, Heppner FL. Alzheimer's amyloid immunotherapy: quo vadis? Lancet Neurol. 2005;4:452-3.

- 109 Hock C, Nitsch RM. Clinical observations with AN-1792 using TAPIR analyses. Neurodegener Dis. 2005;2:273-6.
- 110 Reinikainen KJ, Soininen H, Riekkinen PJ. Neurotransmitter changes in Alzheimer's disease: implications to diagnostics and therapy. J Neurosci Res. 1990;27:576-86.
- 111 Kaduszkiewicz H, Zimmermann T, Beck-Bornholdt HP, van den BH. Cholinesterase inhibitors for patients with Alzheimer's disease: systematic review of randomised clinical trials. BMJ. 2005;331:321-7.
- 112 Gage FH, Bjorklund A. Cholinergic septal grafts into the hippocampal formation improve spatial learning and memory in aged rats by an atropine-sensitive mechanism. J Neurosci. 1986;6:2837-47.
- 113 Hodges H, Allen Y, Kershaw T, Lantos PL, Gray JA, Sinden J. Effects of cholinergic-rich neural grafts on radial maze performance of rats after excitotoxic lesions of the forebrain cholinergic projection system—I. Amelioration of cognitive deficits by transplants into cortex and hippocampus but not into basal forebrain. Neuroscience. 1991;45:587-607.
- 114 Beer S, Kesselring J. High prevalence of multiple sclerosis in Switzerland. Neuroepidemiology. 1994;13:14-8.
- 115 Smith KJ, McDonald WI. The pathophysiology of multiple sclerosis: the mechanisms underlying the production of symptoms and the natural history of the disease. Philos Trans R Soc Lond B Biol Sci. 1999;354:1649-73.
- 116 Gumpel M, Lachapelle F, Gansmuller A, Baulac M, Baron van EA, Baumann N. Transplantation of human embryonic oligodendrocytes into shiverer brain. Ann N Y Acad Sci. 1987;495:71-85.
- 117 Baron-Van EA, Gansmuller A, Duhamel E, Pascal F, Gumpel M. Repair of a myelin lesion by Schwann cells transplanted in the adult mouse spinal cord. J Neuroimmunol. 1992;40:2 35-42.
- 118 Kato T, Honmou O, Uede T, Hashi K, Kocsis JD. Transplantation of human olfactory ensheathing cells elicits remyelination of demyelinated rat spinal cord. Glia. 2000;30:209-18.
- 119 Zhang SC, Ge B, Duncan ID. Adult brain retains the potential to generate oligodendroglial progenitors with extensive myelination capacity. Proc Natl Acad Sci. U S A 1999;96: 4089-94.
- 120 Perez-Bouza A, Glaser T, Brustle O. ES cell-derived glial precursors contribute to remyelination in acutely demyelinated spinal cord lesions. Brain Pathol. 2005;15:208-16.
- 121 Harrison BM. Remyelination by cells introduced into a stable demyelinating lesion in the central nervous system. J Neurol Sci. 1980;46:63-81.
- 122 Baron-Van EA, vellana-Adalid V, Lachapelle F, Liblau R. Schwann cell transplantation and myelin repair of the CNS. Mult Scler. 1997;3:157-61.
- 123 Duncan ID, Paino C, Archer DR, Wood PM. Functional capacities of transplanted cell-sorted adult oligodendrocytes. Dev Neurosci. 1992;14:114-22.
- 124 Groves AK, Barnett SC, Franklin RJ, Crang AJ, Mayer M, Blakemore WF, et al. Repair of demyelinated lesions by transplantation of purified O-2A progenitor cells. Nature. 1993;362:453-5.
- 125 Barnett SC, Hutchins AM, Noble M. Purification of olfactory nerve ensheathing cells from the olfactory bulb. Dev Biol. 1993;155;337-50.
- 126 Tontsch U, Archer DR, Dubois-Dalcq M, Duncan ID. Transplantation of an oligodendrocyte cell line leading to extensive myelination. Proc Natl Acad Sci. U S A 1994;91:11616-20.
- 127 Perez-Bouza A, Wigley CB, Nacimiento W, Noth J, Brook GA. Spontaneous orientation of transplanted olfactory glia influences axonal regeneration. Neuroreport. 1998;9:2971-5.
- 128 Modan B, Wagener DK. Some epidemiological aspects of stroke: mortality/morbidity trends, age, sex, race, socioeconomic status. Stroke. 1992;23:1230-6.
- 129 Borlongan CV, Lind JG, Ilon-Carter O, Yu G, Hadman M, Cheng C, et al. Bone marrow grafts restore cerebral blood flow and blood brain barrier in stroke rats. Brain Res. 2004; 1010:108-16.
- 130 Andrews PW, Damjanov I, Simon D, Banting GS, Carlin C, Dracopoli NC, et al. Pluripotent embryonal carcinoma clones derived from the human teratocarcinoma cell line Tera-2. Differentiation in vivo and in vitro. Lab Invest. 1984;50:147-62.
- 131 Kleppner SR, Robinson KA, Trojanowski JQ, Lee VM. Transplanted human neurons derived from a teratocarcinoma cell line (NTera-2) mature, integrate, and survive for over 1 year in the nude mouse brain. J Comp Neurol. 1995;357:618-32.

132 Borlongan CV, Tajima Y, Trojanowski JQ, Lee VM, Sanberg PR. Transplantation of cryopreserved human embryonal carcinoma-derived neurons (NT2N cells) promotes functional recovery in ischemic rats. Exp Neurol. 1998;149:310-21.

- 133 Kondziolka D, Wechsler L, Goldstein S, Meltzer C, Thulborn KR, Gebel J, et al. Transplantation of cultured human neuronal cells for patients with stroke. Neurology. 2000; 55:565-9.
- 134 Kondziolka D, Steinberg GK, Wechsler L, Meltzer CC, Elder E, Gebel J, et al. Neurotransplantation for patients with subcortical motor stroke: a phase 2 randomized trial. J Neurosurg. 2005;103:38-45.
- 135 Savitz SI, Dinsmore J, Wu J, Henderson GV, Stieg P, Caplan LR. Neurotransplantation of fetal porcine cells in patients with basal ganglia infarcts: a preliminary safety and feasibility study. Cerebrovasc Dis. 2005;20:101-7.
- 136 Bliss T, Guzman R, Daadi M, Steinberg GK. Cell transplantation therapy for stroke. Stroke. 2007;38:817-26.
- 137 Vora N, Jovin T, Kondziolka D. Cell transplantation for ischemic stroke. Neurodegener Dis. 2006;3:101-5.
- 138 Abrahams JM, Gokhan S, Flamm ES, Mehler MF. De novo neurogenesis and acute stroke: are exogenous stem cells really necessary? Neurosurgery. 2004;54:150-5.
- 139 Qureshi AI, Tuhrim S, Broderick JP, Batjer HH, Hondo H, Hanley DF. Spontaneous intracerebral hemorrhage. N Engl J Med. 2001;344:1450-60.
- 140 Altumbabic M, Del Bigio MR. Transplantation of fetal brain tissue into the site of intracerebral hemorrhage in rats. Neurosci Lett. 1998;257:61-4.
- 141 Jeong SW, Chu K, Jung KH, Kim SU, Kim M, Roh JK. Human neural stem cell transplantation promotes functional recovery in rats with experimental intracerebral hemorrhage. Stroke. 2003;34:2258-63.
- 142 Barth A, Guzman R, Andres RH, Mordasini P, Barth L, Widmer HR. Experimental intracerebral hematoma in the rat. Restor Neurol Neurosci. 2007;25:1-7.
- 143 Guzman R, Uchida N, Bliss TM, He D, Christopherson KK, Stellwagen D, et al. Long-term monitoring of transplanted human neural stem cells in developmental and pathological contexts with MRI. Proc Natl Acad Sci. U S A 2007; 104:10211-6.
- 144 Royo NC, Schouten JW, Fulp CT, Shimizu S, Marklund N, Graham DI, et al. From cell death to neuronal regeneration: building a new brain after traumatic brain injury. J Neuropathol Exp Neurol. 2003;62:801-11.
- 145 Schwab ME. Repairing the injured spinal cord. Science. 2002;295:1029-31.
- 146 Bareyre FM, Schwab ME. Inflammation, degeneration and regeneration in the injured spinal cord: insights from DNA microarrays. Trends Neurosci. 2003;26:555-63.
- 147 Hulsebosch CE. Recent advances in pathophysiology and treatment of spinal cord injury. Adv Physiol Educ. 2002; 26:238-55.
- 148 Buchli AD, Rouiller E, Mueller R, Dietz V, Schwab ME. Repair of the injured spinal cord. A joint approach of basic and clinical research. Neurodegener Dis. 2007;4:51-6.
- 149 Gonzenbach RR, Schwab ME. Disinhibition of neurite growth to repair the injured adult CNS: Focusing on Nogo. Cell Mol Life Sci 2007;
- 150 Nornes H, Bjorklund A, Stenevi U. Reinnervation of the denervated adult spinal cord of rats by intraspinal transplants of embryonic brain stem neurons. Cell Tissue Res. 1983;230:15-35.
- 151 McDonald JW, Becker D, Holekamp TF, Howard M, Liu S, Lu A, et al. Repair of the injured spinal cord and the potential of embryonic stem cell transplantation. J Neurotrauma. 2004;21:383-93.
- 152 Stichel CC, Hermanns S, Lausberg F, Muller HW. Effects of schwann cell suspension grafts on axon regeneration in subacute and chronic CNS traumatic injuries. Glia. 1999;28:156-65.
- 153 Ramon-Cueto A, Cordero MI, Santos-Benito FF, Avila J. Functional recovery of paraplegic rats and motor axon regeneration in their spinal cords by olfactory ensheathing glia. Neuron. 2000;25:425-35.
- 154 Feron F, Perry C, Cochrane J, Licina P, Nowitzke A, Urquhart S, et al. Autologous olfactory ensheathing cell transplantation in human spinal cord injury. Brain. 2005; 128:2951-60.
- 155 Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, et al. Multilineage potential of adult human mesenchymal stem cells. Science. 1999;284:143-7.

156 Cummings BJ, Uchida N, Tamaki SJ, Salazar DL, Hooshmand M, Summers R, et al. Human neural stem cells differentiate and promote locomotor recovery in spinal cord-in-jured mice. Proc Natl Acad Sci. U S A 2005;102:14069-74.

- 157 Shields WD. Surgical Treatment of Refractory Epilepsy. Curr Treat Options Neurol. 2004;6:349-56.
- 158 Noe KH, Manno EM. Mechanisms underlying status epilepticus. Drugs Today. (Barc) 2005;41:257-66.
- 159 Thompson KW. Genetically engineered cells with regulatable GABA production can affect afterdischarges and behavioral seizures after transplantation into the dentate gyrus. Neuroscience. 2005;133:1029-37.
- 160 Chu K, Kim M, Jung KH, Jeon D, Lee ST, Kim J, et al. Human neural stem cell transplantation reduces spontaneous recurrent seizures following pilocarpine-induced status epilepticus in adult rats. Brain Res. 2004;1023:213-21.
- 161 Hyman C, Hofer M, Barde YA, Juhasz M, Yancopoulos GD, Squinto SP et al. BDNF is a neurotrophic factor for dopaminergic neurons of the substantia nigra. Nature. 1991;350:230-2.
- 162 Knusel B, Michel PP, Schwaber JS, Hefti F. Selective and nonselective stimulation of central cholinergic and dopaminergic development in vitro by nerve growth factor, basic fibroblast growth factor, epidermal growth factor, insulin and the insulin-like growth factors I and II. J Neurosci. 1990:10:558-70.
- 163 Beck KD, Knusel B, Hefti F. The nature of the trophic action of brain-derived neurotrophic factor, des(1-3)-insulin-like growth factor-1, and basic fibroblast growth factor on mesencephalic dopaminergic neurons developing in culture. Neuroscience. 1993;52:855-66.
- 164 Lin LF, Doherty DH, Lile JD, Bektesh S, Collins F. GDNF: a glial cell line-derived neurotrophic factor for midbrain dopaminergic neurons. Science. 1993;260:1130-2.
- 165 Airaksinen MS, Saarma M. The GDNF family: signalling, biological functions and therapeutic value. Nat Rev Neurosci. 2002;3:383-94.
- 166 Kotzbauer PT, Lampe PA, Heuckeroth RO, Golden JP, Creedon DJ, Johnson EM Jr, et al. Neurturin, a relative of glial-cell-line-derived neurotrophic factor. Nature. 1996;384: 467-70.
- 167 Milbrandt J, de Sauvage FJ, Fahrner TJ, Baloh RH, Leitner ML, Tansey MG, et al. Persephin, a novel neurotrophic factor related to GDNF and neurturin. Neuron. 1998;20: 245-53.
- 168 Baloh RH, Tansey MG, Lampe PA, Fahrner TJ, Enomoto H, Simburger KS, et al. Artemin, a novel member of the GDNF ligand family, supports peripheral and central neurons and signals through the GFRalpha3-RET receptor complex. Neuron. 1998;21:1291-302.
- 169 Beck KD, Valverde J, Alexi T, Poulsen K, Moffat B, Vandlen RA, et al. Mesencephalic dopaminergic neurons protected by GDNF from axotomy-induced degeneration in the adult brain. Nature. 1995;373:339-41.
- 170 Widenfalk J, Widmer HR, Spenger C. GDNF, RET and GFRalpha-1-3 mRNA expression in the developing human spinal cord and ganglia. Neuroreport. 1999;10:1433-9.
- 171 Schaller B, Andres RH, Huber AW, Meyer M, Perez-Bouza A, Ducray AD, et al. Effect of GDNF on differentiation of cultured ventral mesencephalic dopaminergic and non-dopaminergic calretinin-expressing neurons. Brain Res. 2005;1036:163-72.
- 172 Ducray A, Krebs SH, Schaller B, Seiler RW, Meyer M, Widmer HR. GDNF family ligands display distinct action profiles on cultured GABAergic and serotonergic neurons of rat ventral mesencephalon. Brain Res. 2005;
- 173 Reichardt LF. Neurotrophin-regulated signalling pathways. Philos Trans R Soc Lond B Biol Sci. 2006;361:1545-64.
- 174 Georgievska B, Kirik D, Rosenblad C, Lundberg C, Bjorklund A. Neuroprotection in the rat Parkinson model by intrastriatal GDNF gene transfer using a lentiviral vector. Neuroreport. 2002;13:75-82.
- 175 Sun ZH, Lai YL, Li P, Zuo HC, Xie ZP. GDNF augments survival and differentiation of TH-positive neurons in neural progenitor cells. Cell Biol Int 2004;28:323-5.
- 176 Lindholm P, Voutilainen MH, Lauren J, Peranen J, Leppanen VM, Andressoo JO, et al. Novel neurotrophic factor CDNF protects and rescues midbrain dopamine neurons in vivo. Nature. 2007;448:73-7.
- 177 Rosenblad C. Growth factor treatment of neurodegenerative disorders: new developments pave the way for clinical success. IDrugs. 2004;7:243-8.

- 178 Wu D. Neuroprotection in experimental stroke with targeted neurotrophins. NeuroRx. 2005;2:120-8.
- 179 Henry RA, Hughes SM, Connor B. AAV-mediated delivery of BDNF augments neurogenesis in the normal and quinolinic acid-lesioned adult rat brain. Eur J Neurosci. 2007; 25:3513-25.
- 180 Gill SS, Patel NK, Hotton GR, O'Sullivan K, McCarter R, Bunnage M, et al. Direct brain infusion of glial cell line-derived neurotrophic factor in Parkinson disease. Nat Med. 2003;9:589-95.
- 181 Sherer TB, Fiske BK, Svendsen CN, Lang AE, Langston JW. Crossroads in GDNF therapy for Parkinson's disease. Mov Disord. 2006;21:136-41.
- 182 Aebischer P, Kato AC. Treatment of amyotrophic lateral sclerosis using a gene therapy approach. Eur Neurol. 1995;35: 65-8
- 183 Hammang JP, Emerich DF, Winn SR, Lee A, Lindner MD, Gentile FT, et al. Delivery of neurotrophic factors to the CNS using encapsulated cells: developing treatments for neurodegenerative diseases. Cell Transplant. 1995;4(Suppl 1):S27-S28.
- 184 Zurn AD, Widmer HR, Aebischer P. Sustained delivery of GDNF: towards a treatment for Parkinson's disease. Brain Res Brain Res Rev. 2001;36:222-9.
- 185 Sautter J, Tseng JL, Braguglia D, Aebischer P, Spenger C, Seiler RW, et al. Implants of polymer-encapsulated genetically modified cells releasing glial cell line-derived neurotrophic factor improve survival, growth, and function of fetal dopaminergic grafts. Exp Neurol. 1998;149:230-6.
- 186 Bloch J, Bachoud-Levi AC, Deglon N, Lefaucheur JP, Winkel L, Palfi S, et al. Neuroprotective gene therapy for Huntington's disease, using polymer-encapsulated cells engineered to secrete human ciliary neurotrophic factor: results of a phase I study. Hum Gene Ther. 2004;15:968-75.
- 187 Tuszynski MH, Thal L, Pay M, Salmon DP, HS U, Bakay R, et al. A phase 1 clinical trial of nerve growth factor gene therapy for Alzheimer disease. Nat Med. 2005;11:551-5.
- 188 Tuszynski MH. Nerve growth factor gene therapy in Alzheimer disease. Alzheimer Dis Assoc Disord. 2007;21: 179-89.
- 189 Pisati F, Bossolasco P, Meregalli M, Cova L, Belicchi M, Gavina M, et al. Induction of neurotrophin expression via human adult mesenchymal stem cells: implication for cell therapy in neurodegenerative diseases. Cell Transplant. 2007;16:41-55.
- 190 Karlsson J, Emgard M, Brundin P. Comparison between survival of lazaroid-treated embryonic nigral neurons in cell suspensions, cultures and transplants. Brain Res. 2002;955:268-80
- 191 Brundin P, Pogarell O, Hagell P, Piccini P, Widner H, Schrag A, et al. Bilateral caudate and putamen grafts of embryonic mesencephalic tissue treated with lazaroids in Parkinson's disease. Brain. 2000;123 (Pt 7):1380-90.
- 192 Friedlander RM. Apoptosis and caspases in neurodegenerative diseases. N Engl J Med. 2003;348:1365-75.
- 193 Wang X, Zhu S, Drozda M, Zhang W, Stavrovskaya IG, Cattaneo E, et al. Minocycline inhibits caspase-independent and -dependent mitochondrial cell death pathways in models of Huntington's disease. Proc Natl Acad Sci. U S A 2003; 100:10483-7.
- 194 Bartesaghi S, Marinovich M, Corsini E, Galli CL, Viviani B. Erythropoietin: a novel neuroprotective cytokine. Neurotoxicology. 2005;26:923-8.
- 195 Schneider A, Kuhn HG, Schabitz WR. A role for G-CSF (granulocyte-colony stimulating factor) in the central nervous system. Cell Cycle. 2005;4:1753-7.
- 196 Gottron FJ, Ying HS, Choi DW. Caspase inhibition selectively reduces the apoptotic component of oxygen-glucose deprivation-induced cortical neuronal cell death. Mol Cell Neurosci. 1997;9:159-69.
- 197 Ames A. CNS energy metabolism as related to function. Brain Res Brain Res Rev. 2000;34:42-68.
- 198 Wallimann T, Wyss M, Brdiczka D, Nicolay K, Eppenberger HM. Intracellular compartmentation, structure and function of creatine kinase isoenzymes in tissues with high and fluctuating energy demands: the 'phosphocreatine circuit' for cellular energy homeostasis. Biochem J. 1992;281(Pt 1):21-40.
- 199 Wyss M, Schulze A. Health implications of creatine: can oral creatine supplementation protect against neurological and atherosclerotic disease? Neuroscience. 2002;112:243-60.
- 200 Meyer RA, Sweeney HL, Kushmerick MJ. A simple analysis of the «phosphocreatine shuttle». Am J Physiol. 1984;246: C365-C377.

- 201 Beal MF, Hyman BT, Koroshetz W. Do defects in mitochondrial energy metabolism underlie the pathology of neurodegenerative diseases? Trends Neurosci. 1993;16:125-31.
- 202 Browne SE, Beal MF. Oxidative damage and mitochondrial dysfunction in neurodegenerative diseases. Biochem Soc Trans. 1994;22:1002-6.
- 203 Beal MF. Mitochondria, free radicals, and neurodegeneration. Curr Opin Neurobiol. 1996;6:661-6.
- 204 Matthews RT, Ferrante RJ, Klivenyi P, Yang L, Klein AM, Mueller G, et al. Creatine and cyclocreatine attenuate MPTP neurotoxicity. Exp Neurol. 1999;157:142-9.
- 205 Brewer GJ, Wallimann TW. Protective effect of the energy precursor creatine against toxicity of glutamate and betaamyloid in rat hippocampal neurons. J Neurochem. 2000; 74:1968-78.
- 206 Andres RH, Huber AW, Schlattner U, Perez-Bouza A, Krebs SH, Seiler RW, et al. Effects of creatine treatment on the survival of dopaminergic neurons in cultured fetal ventral mesencephalic tissue. Neuroscience. 2005;133:701-13.
- 207 Andres RH, Ducray AD, Huber AW, Perez-Bouza A, Krebs SH, Schlattner U, et al. Effects of creatine treatment on survival and differentiation of GABA-ergic neurons in cultured striatal tissue. J Neurochem. 2005;
- 208 Andres RH, Ducray AD, Perez-Bouza A, Schlattner U, Huber AW, Krebs SH, et al. Creatine supplementation improves dopaminergic cell survival and protects against MPP+ toxicity in an organotypic tissue culture system. Cell Transplant. 2005;14:537-50.
- 209 Eriksson PS, Perfilieva E, Bjork-Eriksson T, Alborn AM, Nordborg C, Peterson DA, et al. Neurogenesis in the adult human hippocampus. Nat Med. 1998;4:1313-7.
- 210 McKay R. Stem cells in the central nervous system. Science. 1997:276:66-71.
- 211 Darsalia V, Heldmann U, Lindvall O, Kokaia Z. Stroke-induced neurogenesis in aged brain. Stroke. 2005;36:1790-5.
- 212 Zhu LL, Zhao T, Li HS, Zhao H, Wu LY, Ding AS, et al. Neurogenesis in the adult rat brain after intermittent hypoxia. Brain Res. 2005;1055:1-6.
- 213 Emsley JG, Mitchell BD, Kempermann G, Macklis JD. Adult neurogenesis and repair of the adult CNS with neural progenitors, precursors, and stem cells. Prog Neurobiol. 2005; 75:321-41.
- 214 Sohur US, Emsley JG, Mitchell BD, Macklis JD. Adult neurogenesis and cellular brain repair with neural progenitors, precursors and stem cells. Philos Trans R Soc Lond B Biol Sc.i 2006;361:1477-97.
- 215 Chen D, Zhao M, Mundy GR. Bone morphogenetic proteins. Growth Factors. 2004;22:233-41.
- 216 Harvey BK, Hoffer BJ, Wang Y. Stroke and TGF-beta proteins: glial cell line-derived neurotrophic factor and bone morphogenetic protein. Pharmacol Ther. 2005;105:113-25.
- 217 Barde YA, Davies AM, Johnson JE, Lindsay RM, Thoenen H. Brain derived neurotrophic factor. Prog Brain Res. 1987; 71:185-9.
- 218 Leibrock J, Lottspeich F, Hohn A, Hofer M, Hengerer B, Masiakowski P, et al. Molecular cloning and expression of brain-derived neurotrophic factor. Nature. 1989;341:149-52.
- 219 Lin LF, Mismer D, Lile JD, Armes LG, Butler ET, III, Vannice JL, et al. Purification, cloning, and expression of ciliary neurotrophic factor (CNTF). Science. 1989;246:1023-5.
- 220 Stockli KA, Lottspeich F, Sendtner M, Masiakowski P, Carroll P, Gotz R, et al. Molecular cloning, expression and regional distribution of rat ciliary neurotrophic factor. Nature. 1989;342:920-3.
- 221 Morrison RS, Keating RF, Moskal JR. Basic fibroblast growth factor and epidermal growth factor exert differential trophic effects on CNS neurons. J Neurosci Res. 1988;21: 71-9.
- 222 Gospodarowicz D, Baird A, Cheng J, Lui GM, Esch F, Bohlen P. Isolation of fibroblast growth factor from bovine adrenal gland: physicochemical and biological characterization. Endocrinology. 1986;118:82-90.
- 223 Grothe C, Timmer M. The physiological and pharmacological role of basic fibroblast growth factor in the dopaminergic nigrostriatal system. Brain Res Rev. 2007;54:80-91.
- 224 Aizenman Y, Weichsel ME Jr, de Vellis J. Changes in insulin and transferrin requirements of pure brain neuronal cultures during embryonic development. Proc Natl Acad Sci. U S A 1986;83:2263-6.
- 225 Baskin DG, Figlewicz DP, Woods SC, Porte D Jr, Dorsa DM. Insulin in the brain. Annu Rev Physiol. 1987;49:335-47.

226 Spranger M, Lindholm D, Bandtlow C, Heumann R, Gnahn H, Naher-Noe M, et al. Regulation of Nerve Growth Factor (NGF) Synthesis in the Rat Central Nervous System: Comparison between the Effects of Interleukin-1 and Various Growth Factors in Astrocyte Cultures and in vivo. Eur J Neurosci. 1990:2:69-76.

- 227 Kamegai M, Niijima K, Kunishita T, Nishizawa M, Ogawa M, Araki M, et al. Interleukin 3 as a trophic factor for central cholinergic neurons in vitro and in vivo. Neuron. 1990; 4:429-36.
- 228 Hama T, Kushima Y, Miyamoto M, Kubota M, Takei N, Hatanaka H. Interleukin-6 improves the survival of mesencephalic catecholaminergic and septal cholinergic neurons from postnatal, two-week-old rats in cultures. Neuroscience. 1991;40:445-52.
- 229 Whittemore SR, Seiger A. The expression, localization and functional significance of beta-nerve growth factor in the central nervous system. Brain Res. 1987;434:439-64.
- 230 Thoenen H, Bandtlow C, Heumann R. The physiological function of nerve growth factor in the central nervous system: comparison with the periphery. Rev Physiol Biochem Pharmacol. 1987;109:145-78.
- 231 Hefti F. Nerve growth factor promotes survival of septal cholinergic neurons after fimbrial transections. J Neurosci. 1986;6:2155-62.
- 232 Widenfalk J, Nosrat C, Tomac A, Westphal H, Hoffer B, Olson L. Neurturin and glial cell line-derived neurotrophic factor receptor-beta (GDNFR-beta), novel proteins related to GDNF and GDNFR-alpha with specific cellular patterns of expression suggesting roles in the developing and adult nervous system and in peripheral organs. J Neurosci. 1997; 17:8506-19.
- 233 Horger BA, Nishimura MC, Armanini MP, Wang LC, Poulsen KT, Rosenblad C et al. Neurturin exerts potent actions on survival and function of midbrain dopaminergic neurons. J Neurosci. 1998;18:4929-37.
- 234 Akerud P, Alberch J, Eketjall S, Wagner J, Arenas E. Differential effects of glial cell line-derived neurotrophic factor and neurturin on developing and adult substantia nigra dopaminergic neurons. J Neurochem. 1999;73:70-8.
- 235 Hohn A, Leibrock J, Bailey K, Barde YA. Identification and characterization of a novel member of the nerve growth factor/brain-derived neurotrophic factor family. Nature. 1990;344:339-41.
- 236 Maisonpierre PC, Belluscio L, Squinto S, Ip NY, Furth ME, Lindsay RM, et al. Neurotrophin-3: a neurotrophic factor related to NGF and BDNF. Science. 1990;247:1446-51.
- 237 Rosenthal A, Goeddel DV, Nguyen T, Lewis M, Shih A, Laramee GR, et al. Primary structure and biological activity of a novel human neurotrophic factor. Neuron. 1990;4: 767-73.

- 238 Hynes MA, Poulsen K, Armanini M, Berkemeier L, Phillips H, Rosenthal A. Neurotrophin-4/5 is a survival factor for embryonic midbrain dopaminergic neurons in enriched cultures. J Neurosci Res. 1994;37:144-54.
- 239 Widmer HR, Hefti F. Neurotrophin-4/5 promotes survival and differentiation of rat striatal neurons developing in culture. Eur J Neurosci. 1994;6:1669-79.
- 240 Zihlmann KB, Ducray AD, Schaller B, Huber AW, Krebs SH, Andres RH, et al. The GDNF family members neurturin, artemin and persephin promote the morphological differentiation of cultured ventral mesencephalic dopaminergic neurons. Brain Res Bull. 2005;68:42-53.
- 241 Derynck R. Transforming growth factor alpha. Cell. 1988; 54:593-5.
- 242 Code RA, Seroogy KB, Fallon JH. Some transforming growth factor-alpha connections and their colocalization with enkephalin in the rat central nervous system. Brain Res. 1987;421:401-5.
- 243 Ren RF, Flanders KC. Transforming growth factors-beta protect primary rat hippocampal neuronal cultures from degeneration induced by beta-amyloid peptide. Brain Res. 1996;732:16-24.
- 244 Bensimon G, Lacomblez L, Meininger V. A controlled trial of recombinant methionyl human BDNF in ALS: The BDNF Study Group (Phase III). Neurology. 1999;52: 1427-33.
- 245 Kalra S, Genge A, Arnold DL. A prospective, randomized, placebo-controlled evaluation of corticoneuronal response to intrathecal BDNF therapy in ALS using magnetic resonance spectroscopy: feasibility and results. Amyotroph Lateral Scler Other Motor Neuron Disord. 2003;4:22-6.
- 246 Borasio GD, Robberecht W, Leigh PN, Emile J, Guiloff RJ, Jerusalem F et al. A placebo-controlled trial of insulin-like growth factor-I in amyotrophic lateral sclerosis. European ALS/IGF-I Study Group. Neurology. 1998;51:583-6.
- 247 Lai EC, Felice KJ, Festoff BW, Gawel MJ, Gelinas DF, Kratz R, et al. Effect of recombinant human insulin-like growth factor-I on progression of ALS. A placebo-controlled study. The North America ALS/IGF-I Study Group. Neurology. 1997;49:1621-30.
- 248 Nutt JG, Burchiel KJ, Comella CL, Jankovic J, Lang AE, Laws ER Jr, et al. Randomized, double-blind trial of glial cell line-derived neurotrophic factor (GDNF) in PD. Neurology. 2003;60:69-73.
- 249 Lang AE, Gill S, Patel NK, Lozano A, Nutt JG, Penn R, et al. Randomized controlled trial of intraputamenal glial cell linederived neurotrophic factor infusion in Parkinson disease. Ann Neurol. 2006;59:459-66.

Formerly: Schweizerische Medizinische Wochenschrift

Swiss Medical Weekly

The European Journal of Medical Sciences

The many reasons why you should choose SMW to publish your research

What Swiss Medical Weekly has to offer:

- SMW's impact factor has been steadily rising. The 2006 impact factor is 1.346.
- Open access to the publication via the Internet, therefore wide audience and impact
- Rapid listing in Medline
- LinkOut-button from PubMed with link to the full text website http://www.smw.ch (direct link from each SMW record in PubMed)
- No-nonsense submission you submit a single copy of your manuscript by e-mail attachment
- Peer review based on a broad spectrum of international academic referees
- Assistance of professional statisticians for every article with statistical analyses
- Fast peer review, by e-mail exchange with the referees
- · Prompt decisions based on weekly conferences of the Editorial Board
- Prompt notification on the status of your manuscript by e-mail
- Professional English copy editing

Editorial Board

Prof. Jean-Michel Dayer, Geneva

Prof Paul Erne, Lucerne

Prof. Peter Gehr. Berne

Prof. André P. Perruchoud, Basel

Prof. Andreas Schaffner, Zurich

(editor in chief)

Prof. Werner Straub, Berne (senior editor)

Prof. Ludwig von Segesser, Lausanne

International Advisory Committee

Prof. K. E. Juhani Airaksinen, Turku, Finland

Prof. Anthony Bayes de Luna, Barcelona,

Prof. Hubert E. Blum, Freiburg, Germany Prof. Walter E. Haefeli, Heidelberg, Ger-

Prof. Nino Kuenzli, Los Angeles, USA Prof. René Lutter, Amsterdam, The Netherlands

Prof. Claude Martin, Marseille, France Prof. Josef Patsch, Innsbruck, Austria Prof. Luigi Tavazzi, Pavia, Italy

We evaluate manuscripts of broad clinical interest from all specialities, including experimental medicine and clinical investigation.

We look forward to receiving your paper!

Guidelines for authors: http://www.smw.ch/set authors.html

All manuscripts should be sent in electronic form, to:

EMH Swiss Medical Publishers Ltd. SMW Editorial Secretariat Farnsburgerstrasse 8 CH-4132 Muttenz

Manuscripts: Letters to the editor: letters@smw.ch **Editorial Board:** Internet:

submission@smw.ch red@smw.ch http://www.smw.ch



Official journal of the Swiss Society of Infectious Diseases, the Swiss Society of Internal Medicine and the Swiss Respiratory Society