# Stem cells with potential to generate insulin-producing cells in man

### Henryk Zulewski

Division of Endocrinology, Diabetes and Clinical Nutrition and Department of Research, University Hospital, Basel, Switzerland

# Summary

Replacement of insulin-producing cells represents an almost ideal treatment for patients with diabetes mellitus type 1. Transplantation of pancreatic islets of Langerhans – although successful in experienced centres – is limited by the lack of donor organs. Generation of insulin-producing cells from stem cells represents an attractive alternative. Stem cells with the potential to differentiate into insulin-producing cells include embryonic stem cells (ESC) as well as adult stem cells from various tissues including the pancreas, liver, central nervous system, bone marrow and adipose tissue. The use of human ESC is hampered by ethical concerns and the inability to create patient specific ESC with therapeutic cloning. Among adult stem cells mesenchymal stem cells appear to have a particular developmental plasticity ex vivo that include their ability to adopt a pancreatic endocrine phenotype. The present review summarises the current knowledge on the development of insulin-producing cells from stem cells with special emphasis on human mesenchymal stem cells isolated from the pancreas and adipose tissue.

Key words: stem cells; diabetes mellitus; pancreas

## Introduction

Diabetes mellitus is a debilitating metabolic disease caused by absent (juvenile or type 1) or insufficient (type 2) insulin production from pancreatic  $\beta$  cells. With an actual prevalence of 5.9% diabetes is affecting 15.7 millions people in the USA and approximately 400,000 in Switzerland (data for CH calculated). Diabetes is associated with serious long-term complications, such as cardiovascular disorders, kidney disease, and blindness. The actual treatment strategies for type 1 diabetes (5–10% of all diabetics) are based on insulin replacement by several injections daily combined with meticulous blood glucose monitoring and life style adaptations. However, even under best cir-

cumstances type 1 diabetic patients are faced with a therapeutic dilemma. A good metabolic control that is prerequisite in order to avoid long-term complications is associated with a high frequency of severe hypoglycaemia (DCCT) [1]. Replacement of pancreatic  $\beta$ -cells would represent an almost ideal treatment that could overcome this therapeutic dilemma. Transplantation of islets of Langerhans was shown to be successful in experienced centres but, due to shortage of organs and life long immunosuppression this therapy can be offered to a very limited number of patients only [2, 3]. Insulin secreting cells generated from stem cells could represent an attractive alternative [4].

This work was supported by grants of the Juvenile Diabetes Research Foundation International (grant number 5 – 2001 – 857) and the Swiss National Science Foundation (NFP 46 Stem Cell Transplants 4046-101232).

# Embryonic stem cells

Embryonic stem cells (ESC) have the potential for almost unlimited supply of  $\beta$ -cells [5] but there are considerable ethical concerns regarding the use of human ESC. The generation of individual, patient specific stem cell-derived  $\beta$ -cells for transplantation purposes requires therapeutic cloning of human ESC. Cloning of embryonic stem cells is already accomplished in mice and other mammals [6]. The only paper so far that claimed to have cloned human ESC reported fabricated data [7]. Another problem with ESC is related to their enormous proliferative capacity and the risk for development of teratocarcinoma. Therefore, many scientists continue to evaluate the not yet exhausted potential of adult stem/progenitor cells. Nevertheless, research with human ESC may help to decipher some crucial steps in development of pancreatic endocrine cells in vitro, since almost all data available on pancreas development were obtained from animal models. Recently, one important step towards differentiation and isolation of endodermal cells, the presumed precursors for pancreatic cells, was achieved in mouse and human ESC [8, 9].

# Potential sources for adult stem cells within the pancreas

Stem/progenitor cells with the potential to differentiate into insulin-producing cells in vitro and/or in vivo were described in pancreatic islets [10–12], pancreatic ducts [13], among the population of pancreatic acinar cells [14-16] and within adult or foetal pancreas without further specification [17–19]. In some instances progenitor cells were postulated in pancreatic ducts that would expand and differentiate into insulin-producing cells in response to specific stimuli [13]. Whereas in the case of acinar cells a de-differentiation appears to be the first step followed by re-differentiation into β-cells. The origin of the bona fide stem cell however, remains somewhat elusive and a recent report questioned the entire concept of  $\beta$ -cell stem/progenitor cells with studies using genetic lineage tracing experiments [20]. With this approach it has

been shown that pre-existing  $\beta$ -cells rather than adult stem/progenitor cells retained a proliferative capacity and may thus represent the major source of new  $\beta$ -cells in adult life, at least in mice [20]. In this study the authors almost excluded the possibility of stem or progenitor cells to play a role in β-cell replacement in adult life. This extreme position however may not be justified by the data, given the fact that the study was not designed to identify precursor cells per se but rather to provide evidence for or against their participation in  $\beta$ -cell regeneration. And, this study is in conflict with numerous recent in vivo and in vitro studies suggesting the existence of pancreatic stem/precursor cells [14–16, 19, 21]. Today, it is not evident which of the concepts will pass the test of time.

# Nestin-expressing mesenchymal stem cells from human islets of Langerhans

The development of normal pancreas is the result of close interaction between mesenchymal and epithelial cells that form the initial buds. Signals from mesenchymal cells direct pancreatic development towards endocrine or exocrine fate ([22] for review). Mesenchymal cells of the developing pancreas express the transcriptional factor islet 1 (Isl-1) [23] that is also expressed in pancreatic epithelial cells, as well as nestin [24], a neural stem cell marker [25]. Nestin-positive cells have been also described in human and rat islets of Langerhans including the hypothesis that these cells may represent a stem cell population [11]. It has been shown that cultured nestin-expressing cells from adult human islets can be differentiated into insulinexpressing cells [11, 12], and nestin-positive cells isolated from human foetal pancreas can be expanded and differentiated into insulin-expressing islet-like clusters that reversed hyperglycaemia in diabetic mice [18]. A recent study described clonal multipotential precursors from adult mouse pancreas that were able to generate neural and pancreatic lineages [19]. The respective precursor cells were generated initially from both nestin<sup>+</sup> and nestin<sup>-</sup> cells, but all of them expressed nestin during the expansion period indicating that nestin may be a marker of proliferating stem cells. Nestin is believed to play an important role in the selec-

tive, unequal partitioning of cytoplasmic components during the division of stem cells so as to maintain one daughter cell as a stem cell and the other daughter cell as a "differentiated" cell [26]. Others described that nestin-expressing cells in the pancreas may be part of vascular endothelial cells or pancreatic stellate cells [27, 28]. In addition, studies including cell lineage analyses indicate that during development nestin-expressing cells may give rise to exocrine pancreatic cells [29, 30] but not endocrine lineages [29–32]. A very recent report however, described expression of exocrine as well as endocrine markers in nestinpositive cells isolated from the developing pancreas of nestin/EGFP transgenic mice [33]. Thus, the developmental potential of nestin-positive cells may also include endocrine cells.

Even though it has been shown that nestin-expressing cells are part of mesenchymal cells of the developing pancreas [24, 34] their fate or function in postnatal life are unknown. Stem cells with a mesenchymal phenotype have been recently shown to develop in human islet cultures *in vitro* by de-differentiation of epithelial  $\beta$  cells induced by powerful growth factors like epidermal growth factor (EGF) or fibroblast growth factor (FGF) [35]. This phenomenon is named epithelial to mesenchymal transition (EMT) [35]. The isletderived mesenchymal cells express nestin and vimentin and were able to re-differentiate into insulin-producing cells given the appropriate stimuli [35].

In our current research project we isolated and immortalised on a single cell basis nestin-positive cells from cultured human islets. Interestingly, these cells were also positive for Isl-1 that is known for its critical role in the development of pancreatic endocrine cells. In addition, the isolated nestin- and Isl-1-positive cells displayed a mesenchymal phenotype as mirrored by their ability to differentiate into adipocytic and osteocytic phenotypes given the appropriate stimuli [36]. These cells were negative for insulin and the insulin promoter factor 1 (Ipf-1). Upon differentiation with serum free medium supplemented with a cocktail of differentiation stimulating factors they could be induced to form islet-like cluster and to express several pancreatic developmental genes. This includes the transcription factors Ipf-1, Isl-1, Pax-4, Pax-6, Ngn-3, Nkx-2.2 and Nkx-6.1, as well as the pancreatic endocrine genes insulin, glucagon and somatostatin [36] (figure 1). Glucagon and c-peptide-positive cells were also identified by immunocytochemistry and electron microscopy (figure 2).

40 bp

69 bp

73 bp

299 bp

50 bp

54 bp

84 bu

248 bp

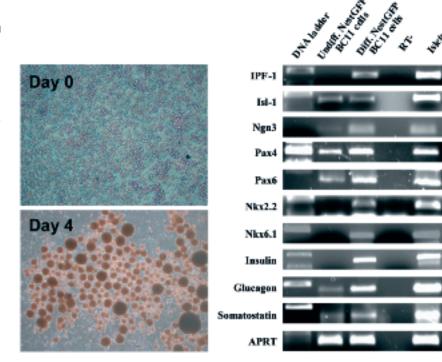
80 bp

82 bp

84 bp

#### Figure 1

Differentiation of reversibly immortalised islet-derived nestinpositive stem cells (Nest GFP-BC11 cells) into an endocrine phenotype. After 4 days of culture in differentiation medium the cells formed isletlike clusters (left panels). Expression of pancreatic endocrine transcription factors including lpf-1, lsl-1, Ngn3, Pax4, Pax6, Nkx2.2, and Nkx6.1 as well as the mRNA transcripts of the islet genes insulin, alucagon and somatostatin (right panel). After Eberhardt et al. [75]. Reproduction with permission of the publisher.

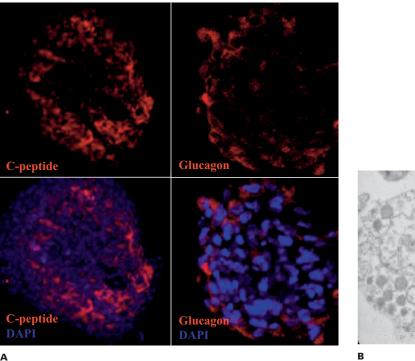


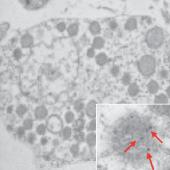
#### Figure 2

Immunofluoresence staining of differentiated reversibly immortalised islet-derived mesenchymal stem cells.

A. Islet-like clusters were stained with specific antibodies against C-peptide or glucagon. Nuclei staining was performed with DAPI. Image was obtained with a laser scanning confocal microscope (Zeiss LSM 510). Original magnification x 200 for C-peptide image and, x 400 for glucagon. B. Electron microscopy studies revealed formation of granules in differentiated cells in contrast to undifferentiated cells. Red arrows indicate immunogold labelling for C-peptide (inset). After Eberhardt et al. [75]

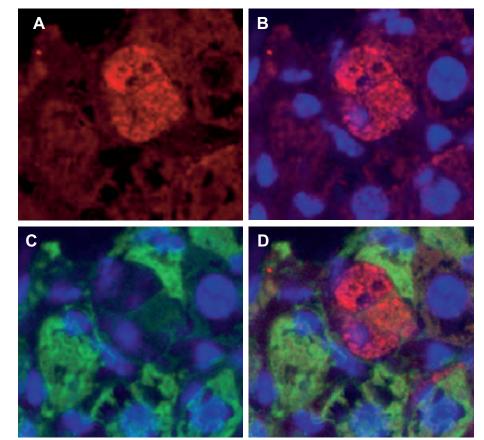
Α





# Figure 3

Engraftment of cultured human isletderived cells into mouse liver. Fluorescence-immunohistochemistry with human and mouse specific antibodies against albumin using confocal microscopy with 630 fold magnification. Panel (A) shows one cell stained with antibodies against human albumin, (B) the same cell with additional DAPI staning for cell nuclei, (C) albumin staining with antibodies against mouse albumin, (D) digital overlay of human and mouse albumin staining showing no co-staining for mouse albumin in the human albumin-positive cell. From von Mach et al. [37], reproduction with permission of the publisher.



C-peptide is part of the proinsulin peptide and thus indicator of de-novo insulin synthesis. Although these results are encouraging and indicate that such cells may have the potential to become functioning  $\beta$ -cells the actual differentiation efficacy is limited and the cells are not yet able to secrete insulin in response to glucose.

There is however evidence that cultured nestin-positive cells from human islets are multipotent as they can adopt a hepatic phenotype *in vivo*. We have recently shown that nestin-positive

islet-derived cells differentiate in human albuminproducing cells if grafted directly into mouse liver (figure 3) [37]. Similar results were obtained with our immortalised nestin- and Isl-1-positive cells [36]. Interestingly, human bone marrow-derived mesenchymal stem cells (MSC) were also recently described to differentiate into a hepatic phenotype *in vivo* without evidence for cell fusion [38]. These data suggest that cells with a mesenchymal phenotype may share properties of hepatic precursor/ stem cells.

# Stem cells outside the pancreas as potential source for $\beta$ -cell replacement

Stem cells with the potential to differentiate into insulin-producing cells have been also described in the liver [39, 40], the central nervous system [41], the spleen [42] and bone marrow [43–46]. Some of the in vivo reports however are controversial and were not confirmed by others [47-51]. Another hypothesis generated by recent data suggests that at least in vivo bone marrow-derived cells could play a supportive role in pancreas regeneration rather than participate in the differentiation of endocrine cells themselves [48]. Such developmental potential of bone marrow-derived MSC however was described in vitro. MSC from mouse and rat bone marrow were shown to harbour the potential to differentiate into insulin-secreting cells in vitro and to reverse hyperglycaemia in an animal model of diabetes [44, 46]. Recently human bone marrow-derived MSC were described to express at low level the islet transcription factor Nkx-6.1 and to differentiate into insulin-expressing cells upon adenoviral transduction with vectors encoding the transcription factors Ipf-1, Hlxb-9 or Foxa-2 [52]. The mechanisms underlying this apparent developmental plasticity of MSC are unknown. Interestingly, MSC were also shown to bear the potential to adopt a neural phenotype in vitro and in vivo [53-56] in rodents and humans [55] suggesting a neuro-endocrine developmental capacity of these cells. Expanding MSC express several stem cell marker-like stem cell factors (SCF) and Thy-1 [57, 58] but also nestin [55, 56, 59]. Neural precursor cells express besides nestin the side population stem cell marker ABCG2 [60, 61]. ABCG2 expression was described in nestin-positive islet-derived precursor cells [62]. Nestin and possibly ABCG2 expression

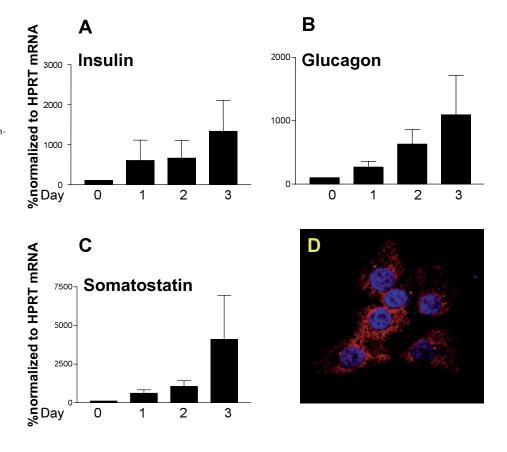
could therefore represent a possible link between MSC and their ability to differentiate into neuroendocrine cells. In addition to bone marrowderived MSC another cell type of haematopoietic origin was recently discovered as potential source for pluripotent stem cells. Human peripheral blood monocyte-derived subset of proliferating cells appear to have the potential to differentiate *in*  *vitro* in different cell lineages including neural, hepatic [63] and pancreatic phenotype with induction of insulin production [64]. There is an increasing body of evidence suggesting that adult stem cells may indeed be converted into insulin-producing cells although the efficacy needs substantial improvements and the mechanisms responsible for this phenomenon are poorly understood.

# Mesenchymal stem cells from human adipose tissue

MSC from human bone marrow and adipose tissue represent a very similar cell population with comparable phenotypes [65–68]. Thus, MSC with the potential to adopt a pancreatic endocrine phenotype could also exist in human adipose tissue. We isolated human adipose tissue-derived MSC (adMSC) from four donors and expanded the cells in FGF containing culture medium. Proliferating adMSC expressed the stem cell markers ABCG2, nestin, SCF and Thy-1 but also Isl-1 mRNA. Using immunocytochemistry we found Isl-1 protein in the nuclei of approximately 10% of cultured adMSC [69]. When subjected to serum-free differentiation medium an up-regulation of the transcription factors Ipf-1, Isl-1 and Ngn-3 and the islet genes insulin, glucagon and somatostatin was observed (figure 4a–c). In parallel, c-peptidepositive cells were found after differentiation (figure 4d) [69].

Figure 4

A-C: Induction of the islet genes insulin, glucagon and somatostatin in adipose tissuederived MSC in response to defined culture conditions. D: Immunocytochemistry for C-peptide adipose tissue-derived MSC (magnification x 200). After Timper et al. [69], reproduction with permission of the publisher.



# Expression of Isl-1 in mesenchymal stem cells

The phenotype of mesenchymal stem cells that are able to turn on Isl-1 and other early transcription factors in response to defined culture conditions is unknown and needs further studies. Expression of Isl-1 is together with Ipf-1 one of the earliest pancreatic markers detected in the pancreatic anlage at embryonic day E9 in the mouse [23]. Isl-1 is crucial for the generation of pancreatic endocrine cells and explants of the pancreatic anlage from Isl-1 knock-out mice were unable to give rise to insulin- and glucagon-positive cells as did the wild-type controls [23]. Isl-1 expression in our MSC could thus reflect the first crucial step towards induction of other developmental transcription factors and pancreatic islet genes. But, Isl-1 is also involved in the development of the central nervous system, especially motorneurons [70, 71] and it has been recently also shown to play a role in heart development [72, 73]. Therefore, at least some of the Isl-1-positive MSC may equally represent potential precursors for these organs.

# Potential and limitations of adult mesenchymal stem cells

It has been shown that human bone marrowderived MSC can be differentiated into hepatocytes in vivo without evidence for cell fusion if xenografted directly into the rat liver [38]. This demonstrates an unexpected developmental potential of these cells. Liver and pancreas are believed to origin from similar endodermal precursors during development. Pancreatic stem/precursor cells can give rise to hepatocyte and vice versa [37, 39, 74, 75]. It is therefore tempting to speculate that human adipose tissue- as well as bone marrow-derived MSC harbour the potential to adopt a pancreatic endocrine phenotype and give rise to functioning insulin-secreting cells. We have shown that human MSC from pancreatic islets as well as adipose tissue can be induced to activate pancreatic developmental genes including Isl-1, Ipf-1, Ngn-3, Pax-4, Pax-6, Nkx-2.2, Nkx-6.1, as well as the islet genes insulin, glucagon and somatostatin. This differentiation was achieved without genetic modifications of the cells. However, we are still far away from production of clinically meaningful amounts of insulin.

Although, impressive knowledge accumulated in recent years regarding pancreas development and especially the role of crucial transcription factors required for proper differentiation of pancreatic  $\beta$ -cells [76], there is considerable lack of information on factors needed for in vitro differentiation of stem cells; ESC and adult stem cells alike. There is no unequivocally accepted protocol

for differentiation of stem cells into insulin-producing cells and there is little general agreement on the cell type that should be studied. The overall potential of adult cells to change their fate in vitro and differentiate into almost every tissue was again demonstrated recently, using adult fibroblasts that were converted into cells with ESC-like phenotype by manipulating just 4 important factors [77]. There is substantial circumstantial evidence suggesting that MSC may have the potential to differentiate into various tissues including insulin-producing cells in vitro. But, the candidate cells among the MSC that harbour this potential need to be identified together with a considerable improvement of current differentiation protocols in order to achieve significant advancements in the search for stem cell-based therapies for diabetes mellitus type 1.

Correspondence: Dr. Henryk Zulewski Departement Forschung Klinik für Endokrinologie, Diabetologie und klin. Ernährung Universitätskliniken Hebelstrasse 20 CH-4031 Basel Switzerland henryk.zulewski@unibas.ch

# References

- 1 The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. The Diabetes Control and Complications Trial Research Group. N Engl J Med. 1993;329(14): 977–86.
- 2 Shapiro AMJ, Lakey JRT, Ryan EA, et al. Islet transplantation in seven patients with type 1 diabetes mellitus using a glucocorticoid-free immunosuppressive regimen. N Engl J Med. 2000; 343(4):230–8.
- 3 Shapiro AM, Ricordi C, Hering B. Edmonton's islet success has indeed been replicated elsewhere. Lancet. 2003;362(9391): 1242.
- 4 Scharfmann R. Alternative sources of beta cells for cell therapy of diabetes. Eur J Clin Invest. 2003;33(7):595–600.
- 5 Soria B, Roche E, Berna G, Leon-Quinto T, Reig JA, Martin F. Insulin-secreting cells derived from embryonic stem cells normalize glycemia in streptozotocin-induced diabetic mice. Diabetes. 2000;49(2):157–62.
- 6 Jaenisch R. Human cloning the science and ethics of nuclear transplantation. N Engl J Med. 2004;351(27):2787–91.

- 7 Chong S, Normile D. Stem cells. How young Korean researchers helped unearth a scandal. Science. 2006;311(5757): 22–5.
- 8 D'Amour KA, Agulnick AD, Eliazer S, Kelly OG, Kroon E, Baetge EE. Efficient differentiation of human embryonic stem cells to definitive endoderm. Nat Biotechnol. 2005;23(12): 1534–41.
- 9 Yasunaga M, Tada S, Torikai-Nishikawa S, et al. Induction and monitoring of definitive and visceral endoderm differentiation of mouse ES cells. Nat Biotechnol. 2005;23(12):1542–50.
- 10 Guz Y, Nasir I, Teitelman G. Regeneration of pancreatic beta cells from intra-islet precursor cells in an experimental model of diabetes. Endocrinology. 2001;142(11):4956–68.
- 11 Zulewski H, Abraham EJ, Gerlach MJ, et al. Multipotential nestin-positive stem cells isolated from adult pancreatic islets differentiate exvivo into pancreatic endocrine, exocrine, and hepatic phenotypes. Diabetes. 2001;50(3):521–33.
- 12 Abraham EJ, Leech CA, Lin JC, Zulewski H, Habener JF. Insulinotropic hormone glucagon-like peptide-1 differentiation of human pancreatic islet-derived progenitor cells into insulinproducing cells. Endocrinology. 2002;143(8):3152–61.

- 13 Bonner-Weir S, Sharma A. Pancreatic stem cells. J Pathol. 2002; 197(4):519–26.
- 14 Baeyens L, De Breuck S, Lardon J, Mfopou JK, Rooman I, Bouwens L. In vitro generation of insulin-producing beta cells from adult exocrine pancreatic cells. Diabetologia. 2005;48(1): 49–57.
- 15 Lardon J, Huyens N, Rooman I, Bouwens L. Exocrine cell transdifferentiation in dexamethasone-treated rat pancreas. Virchows Arch. 2004;444(1):61–5.
- 16 Minami K, Okuno M, Miyawaki K, et al. Lineage tracing and characterization of insulin-secreting cells generated from adult pancreatic acinar cells. Proc Natl Acad Sci U S A. 2005;102(42): 15116–21.
- 17 Ramiya VK, Maraist M, Arfors KE, Schatz DA, Peck AB, Cornelius JG. Reversal of insulin-dependent diabetes using islets generated in vitro from pancreatic stem cells [see comments]. Nat Med. 2000;6(3):278–82.
- 18 Huang H, Tang X. Phenotypic determination and characterization of nestin-positive precursors derived from human fetal pancreas. Lab Invest. 2003;83(4):539–47.
- 19 Seaberg RM, Smukler SR, Kieffer TJ, et al. Clonal identification of multipotent precursors from adult mouse pancreas that generate neural and pancreatic lineages. Nat Biotechnol. 2004;22(9):1115–24.
- 20 Dor Y, Brown J, Martinez OI, Melton DA. Adult pancreatic beta-cells are formed by self-duplication rather than stem-cell differentiation. Nature. 2004;429(6987):41–6.
- 21 Lee CS, De Leon DD, Kaestner KH, Stoffers DA. Regeneration of pancreatic islets after partial pancreatectomy in mice does not involve the reactivation of neurogenin-3. Diabetes. 2006;55(2):269–72.
- 22 Scharfmann R. Control of early development of the pancreas in rodents and humans: implications of signals from the mesenchyme. Diabetologia. 2000;43(9):1083–92.
- 23 Ahlgren U, Pfaff SL, Jessell TM, Edlund T, Edlund H. Independent requirement for ISL1 in formation of pancreatic mesenchyme and islet cells. Nature. 1997;385(6613):257–60.
- 24 Selander L, Edlund H. Nestin is expressed in mesenchymal and not epithelial cells of the developing mouse pancreas. Mech Dev. 2002;113(2):189–92.
- 25 Lendahl U, Zimmerman LB, McKay RD. CNS stem cells express a new class of intermediate filament protein. Cell. 1990; 60(4):585–95.
- 26 Chou YH, Khuon S, Herrmann H, Goldman RD. Nestin promotes the phosphorylation-dependent disassembly of vimentin intermediate filaments during mitosis. Mol Biol Cell. 2003; 14(4):1468–78.
- 27 Lardon J, Rooman I, Bouwens L. Nestin expression in pancreatic stellate cells and angiogenic endothelial cells. Histochem Cell Biol. 2002;117(6):535–40.
- 28 Klein T, Ling Z, Heimberg H, Madsen OD, Heller RS, Serup P. Nestin is expressed in vascular endothelial cells in the adult human pancreas. J Histochem Cytochem. 2003;51(6):697–706.
- 29 Esni F, Stoffers DA, Takeuchi T, Leach SD. Origin of exocrine pancreatic cells from nestin-positive precursors in developing mouse pancreas. Mech Dev. 2004;121(1):15–25.
- 30 Delacour A, Nepote V, Trumpp A, Herrera PL. Nestin expression in pancreatic exocrine cell lineages. Mech Dev. 2004;121 (1):3–14.
- 31 Treutelaar MK, Skidmore JM, Dias-Leme CL, et al. Nestin-Lineage Cells Contribute to the Microvasculature but Not Endocrine Cells of the Islet. Diabetes. 2003;52(10):2503–12.
- 32 Humphrey RK, Bucay N, Beattie GM, et al. Characterization and Isolation of Promoter-Defined Nestin-Positive Cells from the Human Fetal Pancreas. Diabetes. 2003;52(10):2519–25.
- 33 Bernardo AS, Barrow J, Hay CW, et al. Presence of endocrine and exocrine markers in EGFP-positive cells from the developing pancreas of a nestin/EGFP mouse. Mol Cell Endocrinol. 2006;253(1-2):14–21.
- 34 Edlund H. Pancreatic organogenesis developmental mechanisms and implications for therapy. Nat Rev Genet. 2002; 3(7):524–32.
- 35 Gershengorn MC, Hardikar AA, Hardikar A, et al. Epithelialto-Mesenchymal Transition Generates Proliferative Human Islet Precursor Cells. Science. 2004.
- 36 Eberhardt M SP, von Mach MA, HengstlerJG, Brulport M, Linscheid P, Seboek D, Oberholzer J, Barbero A, Martin I, Müller B, Trono D, Zulewski H. Multipotential nestin and Isl-1 positive mesenchymal stem cells isolated from human pancreatic islets Biochem Biophys Res Commun. 2006: in press.

- 37 von Mach MA, Hengstler JG, Brulport M, et al. In vitro cultured islet-derived progenitor cells of human origin express human albumin in severe combined immunodeficiency mouse liver in vivo. Stem Cells. 2004;22(7):1134–41.
- 38 Sato Y, Araki H, Kato J, et al. Human mesenchymal stem cells xenografted directly to rat liver are differentiated into human hepatocytes without fusion. Blood. 2005;106(2):756–63.
- 39 Zalzman M, Gupta S, Giri RK, et al. Reversal of hyperglycemia in mice by using human expandable insulin-producing cells differentiated from fetal liver progenitor cells. Proc Natl Acad Sci U S A. 2003;100(12):7253–8.
- 40 Sapir T, Shternhall K, Meivar-Levy I, et al. Cell-replacement therapy for diabetes: Generating functional insulin-producing tissue from adult human liver cells. Proc Natl Acad Sci U S A. 2005;102(22):7964–9.
- 41 Hori Y, Gu X, Xie X, Kim SK. Differentiation of insulin-producing cells from human neural progenitor cells. PLoS Med. 2005;2(4):e103.
- 42 Kodama S, Kuhtreiber W, Fujimura S, Dale EA, Faustman DL. Islet regeneration during the reversal of autoimmune diabetes in NOD mice. Science. 2003;302(5648):1223–7.
- 43 Ianus A, Holz GG, Theise ND, Hussain MA. In vivo derivation of glucose-competent pancreatic endocrine cells from bone marrow without evidence of cell fusion. J Clin Invest. 2003;111 (6):843–50.
- 44 Tang DQ, Cao LZ, Burkhardt BR, et al. In vivo and in vitro characterization of insulin-producing cells obtained from murine bone marrow. Diabetes. 2004;53(7):1721–32.
- 45 D'Ippolito G, Diabira S, Howard GA, Menei P, Roos BA, Schiller PC. Marrow-isolated adult multilineage inducible (MIAMI) cells, a unique population of postnatal young and old human cells with extensive expansion and differentiation potential. J Cell Sci. 2004;117(Pt 14):2971–81.
- 46 Oh SH, Muzzonigro TM, Bae SH, LaPlante JM, Hatch HM, Petersen BE. Adult bone marrow-derived cells trans-differentiating into insulin-producing cells for the treatment of type I diabetes. Lab Invest. 2004;84(5):607–17.
- 47 Lechner A, Yang YG, Blacken RA, Wang L, Nolan AL, Habener JF. No evidence for significant transdifferentiation of bone marrow into pancreatic beta-cells in vivo. Diabetes. 2004;53(3): 616–23.
- 48 Hess D, Li L, Martin M, et al. Bone marrow-derived stem cells initiate pancreatic regeneration. Nat Biotechnol. 2003;21(7): 763–70.
- 49 Chong AS, Shen J, Tao J, et al. Reversal of diabetes in non-obese diabetic mice without spleen cell-derived beta cell regeneration. Science. 2006;311(5768):1774–5.
- 50 Nishio J, Gaglia JL, Turvey SE, Campbell C, Benoist C, Mathis D. Islet recovery and reversal of murine type 1 diabetes in the absence of any infused spleen cell contribution. Science. 2006; 311(5768):1775–8.
- 51 Suri A, Calderon B, Esparza TJ, Frederick K, Bittner P, Unanue ER. Immunological reversal of autoimmune diabetes without hematopoietic replacement of beta cells. Science. 2006;311 (5768):1778–80.
- 52 Moriscot C, de Fraipont F, Richard MJ, et al. Human bone marrow mesenchymal stem cells can express insulin and key transcription factors of the endocrine pancreas developmental pathway upon genetic and/or microenvironmental manipulation in vitro. Stem Cells. 2005;23(4):594–603.
- 53 Mezey E, Chandross KJ, Harta G, Maki RA, McKercher SR. Turning blood into brain: cells bearing neuronal antigens generated in vivo from bone marrow. Science. 2000;290(5497): 1779–82.
- 54 Azizi SA, Stokes D, Augelli BJ, DiGirolamo C, Prockop DJ. Engraftment and migration of human bone marrow stromal cells implanted in the brains of albino rats – similarities to astrocyte grafts. Proc Natl Acad Sci U S A. 1998;95(7):3908–13.
- 55 Woodbury D, Schwarz EJ, Prockop DJ, Black IB. Adult rat and human bone marrow stromal cells differentiate into neurons. J Neurosci Res. 2000;61(4):364–70.
- 56 Munoz-Elias G, Marcus AJ, Coyne TM, Woodbury D, Black IB. Adult bone marrow stromal cells in the embryonic brain: engraftment, migration, differentiation, and long-term survival. J Neurosci. 2004;24(19):4585–95.
- 57 Ashman LK. The biology of stem cell factor and its receptor C-kit. Int J Biochem Cell Biol. 1999;31(10):1037–51.
- 58 Mareschi K, Ferrero I, Rustichelli D, et al. Expansion of mesenchymal stem cells isolated from pediatric and adult donor bone marrow. J Cell Biochem. 2005.

- 59 Vogel W, Grunebach F, Messam CA, Kanz L, Brugger W, Buhring HJ. Heterogeneity among human bone marrow-derived mesenchymal stem cells and neural progenitor cells. Haematologica. 2003;88(2):126–33.
- 60 Cai J, Cheng A, Luo Y, et al. Membrane properties of rat embryonic multipotent neural stem cells. J Neurochem. 2004;88 (1):212–26.
- 61 Zhou S, Schuetz JD, Bunting KD, et al. The ABC transporter Bcrp1/ABCG2 is expressed in a wide variety of stem cells and is a molecular determinant of the side-population phenotype. Nat Med. 2001;7(9):1028–34.
- 62 Lechner A, Leech CA, Abraham EJ, Nolan AL, Habener JF. Nestin-positive progenitor cells derived from adult human pancreatic islets of Langerhans contain side population (SP) cells defined by expression of the ABCG2 (BCRP1) ATP-binding cassette transporter. Biochem Biophys Res Commun. 2002;293 (2):670–4.
- 63 Zhao Y, Glesne D, Huberman E. A human peripheral blood monocyte-derived subset acts as pluripotent stem cells. Proc Natl Acad Sci U S A. 2003;100(5):2426–31.
- 64 Ruhnke M, Ungefroren H, Nussler A, et al. Differentiation of in vitro-modified human peripheral blood monocytes into hepatocyte-like and pancreatic islet-like cells. Gastroenterology. 2005;128(7):1774–86.
- 65 Zuk PA, Zhu M, Ashjian P, et al. Human adipose tissue is a source of multipotent stem cells. Mol Biol Cell. 2002;13(12): 4279–95.
- 66 De Ugarte DA, Morizono K, Elbarbary A, et al. Comparison of multi-lineage cells from human adipose tissue and bone marrow. Cells Tissues Organs. 2003;174(3):101–9.
- 67 Lee RH, Kim B, Choi I, et al. Characterization and expression analysis of mesenchymal stem cells from human bone marrow and adipose tissue. Cell Physiol Biochem. 2004;14(4-6):311–24.

- 68 Dicker A, Le Blanc K, Astrom G, et al. Functional studies of mesenchymal stem cells derived from adult human adipose tissue. Exp Cell Res. 2005;308(2):283–90.
- 69 Timper K, Seboek D, Eberhardt M, et al. Human adipose tissue-derived mesenchymal stem cells differentiate into insulin, somatostatin, and glucagon expressing cells. Biochem Biophys Res Commun. 2006;341(4):1135–40.
- 70 Ericson J, Thor S, Edlund T, Jessell TM, Yamada T. Early stages of motor neuron differentiation revealed by expression of homeobox gene Islet-1. Science. 1992;256(5063):1555–60.
- 71 Tsuchida T, Ensini M, Morton SB, et al. Topographic organization of embryonic motor neurons defined by expression of LIM homeobox genes. Cell. 1994;79(6):957–70.
- 72 Cai CL, Liang X, Shi Y, et al. Isl1 identifies a cardiac progenitor population that proliferates prior to differentiation and contributes a majority of cells to the heart. Dev Cell. 2003;5(6): 877–89.
- 73 Laugwitz KL, Moretti A, Lam J, et al. Postnatal isl1<sup>+</sup> cardioblasts enter fully differentiated cardiomyocyte lineages. Nature. 2005;433(7026):647–53.
- 74 Grompe M. Pancreatic-hepatic switches in vivo. Mech Dev. 2003;120(1):99–106.
- 75 Eberhardt M, Salmon P, von Mach M, et al. Multipotential nestin and Isl-1 positive mesenchymal stem cells isolated from human pancreatic islets Biochem Biophys Res Commun. 2006: in press.
- 76 Habener JF, Kemp DM, Thomas MK. Minireview: transcriptional regulation in pancreatic development. Endocrinology. 2005;146(3):1025–34.
- 77 Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell. 2006;126(4):663–76.