Peer reviewed article

# Defects of airway smooth muscle cell function are important in asthma

M. Tamma, M Rotha, b

- <sup>a</sup> Pneumology, Pulmonary Cell Research, University Hospital Basel, Basel, Switzerland
- b The Woolcock Institute for Medical Research, Molecular Medicine, Camperdown, Sydney, Australia

#### **Summary**

For many years asthma has been regarded as an inflammatory disease of the airway mucosa leading to bronchial hyperreactivity. Recent studies showed marked abnormalities in airway smooth muscle behaviour in patients with asthma. The pathogenesis of asthma seems to consist of airway inflammation combined with airway smooth mus-

cle remodelling. The latter pathology is linked to a lack of the ant-proliferative transcription factor C/EBP- $\alpha$  in this specific cell type.

Key words: asthma; airway smooth muscle; gluco-corticoid receptor; C/EBP- $\alpha$ 

#### Introduction

Asthma is a chronic inflammatory disease of the airways and its incidence is increasing world wide without any known reason [1–7]. Up to the mid- nineties there was an increase of asthma incidence also in Switzerland, which seems to have reached a plateau over the last ten years. With respect to geographic variation and definition asthma affects 8–18% of children and about 8% of adults. Asthma is in most cases a life long disease, which often requires continuous therapy and therefore contributes significantly to the overall health care budget [8, 9].

Asthma is characterised by chronic airway and mucosa inflammation, tissue infiltration with eosinophils, lymphocytes and mast cells [10, 11]. In addition, the adherence of the epithelial cell layer seems to be disturbed [12–14]. This pathology of the epithelium in asthma airways has been suggested to disrupt the interaction between the epithelial cell layer and the underlying connective

tissue [10-15]. As a consequence there is increased airway remodelling reflected in the thickening of the basement membrane and neo-vascularisation [10-15]. The cellular communication and interaction between bronchial epithelial cells and fibroblast like cells (myo-fibroblasts) are the focus of current research. The thickness of the connective tissue sheet underlying the basement membrane is also increased with infiltrated myofibroblasts of unknown origin and enhanced deposition of extracellular matrix, a pathology that is often described as sub-epithelial fibrosis [12-17]. In contrast to asthma fibrotic processes in chronic obstructive pulmonary disease (COPD) include fibrosis of the bronchioli [16, 17]. There is increasing evidence that smooth muscle hyperplasia plays an important role in asthma [23–38]. The major pathological differences of the bronchial wall, comparing asthmatic to non-asthmatic tissue, is depicted in figure 1.

### Classical hypothesis of asthma as an inflammatory lung disease

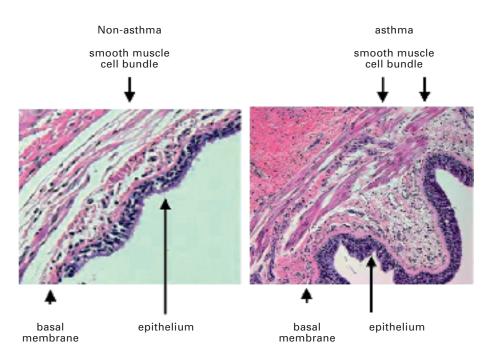
Asthma is associated with inflammation of the airway mucosa [11, 16, 19]. A significant number of asthma patients also suffer from allergic rhinoconjunctivitis [39–46]. Therefore the concept of one airway one disease was favoured [43]. The airway inflammation consists of mucosal thickening with infiltration of inflammatory cells including eosinophils and lymphocytes. Th2 cells contribute

to the aggravated inflammation in asthma since they release cytokines, which are involved in the activation of other inflammatory cells such as mast cells [47–49]. Mast cells have also been implicated in airway inflammation in asthma. Many groups have investigated their role in inflammation and tissue remodelling [50–54]. In this context it is important to note that mast cells have been shown to

Asthma research is supported by Swiss National Foundation, AstraZeneca, Fuchopharma, GlaxoSmithKline.

Figure 1

Histology of the non-asthmatic (left panel) and the asthmatic (right panel) airway wall. In the non-asthmatic airway the bundles of smooth muscle cells is thin and consists of a few cells only, while in the asthmatic airway wall the number of muscle bundles as well as the number of smooth muscle cells per bundle increases markedly. The basement membrane in the non-asthmatic airway is a think layer separating the epithelial cell layer from a thin layer of connective tissue formed by extracellular matrix and fibroblast like cells (myofibroblasts). In contrast in the asthmatic airway the basement membrane is significantly thickened as is the mass of the connective tissue layer.



interact with airway smooth muscle cells and modulate their function [55]. Two recent reviews by Norris [56] and Marrone et al. [57] summarise the contribution of mast cells to asthma in detail.

Major factors linked to mast cell activation in asthma are IgE, IL-4, and IL-5, which are all well known to be up-regulated in asthma patients. IgE together with IL-4 have long been known to be up-regulated in asthma patients and in their first degree relatives, but there is no study until today that could establish a clear genetic link of one of the factors to the inheritance of asthma. Furthermore, IgE inhibition by antibodies resulted in a marked down-regulation of the inflammatory symptoms but did not cure asthma and the symptoms re-appeared after cessation of anti-IgE therapy [58, 59]. Anti-IL-5 antibodies markedly down-regulated eosinophila in asthma but could not reduce the symptoms [60].

In view of asthma as an inflammatory disease of the lung the role of the Th1/Th2 ratio has been investigated intensively. Most studies however,

revealed that the shift towards Th2 cells in the lung and in circulating blood was rather associated with allergy than with asthma [61-69]. The early dominance of the Th2 cell type in asthma has been also demonstrated in childhood asthma [61, 62]. Additional studies investigated the consequences of Th2 cytokine dependent cellular signalling and its role in the inflammatory processes of asthma [65-68]. Interestingly Th2 cells interact directly with airway smooth muscle cells and respond to pro-inflammatory cytokines released by this bronchial cell type [66]. This fact suggests that Th2 recruitment could be caused by activated asthmatic airway smooth muscle cells. A similar interaction has been described for Th2 cells that interact with dendritic antigen presenting cells and both cell types activate each other in the presence of antigens [69]. In summary, the cause and the role of the Th1/Th2 cell imbalance in asthma as well as in atopy has to be re-defined and is likely to be a sign of allergic inflammation.

#### How to control inflammation in asthma?

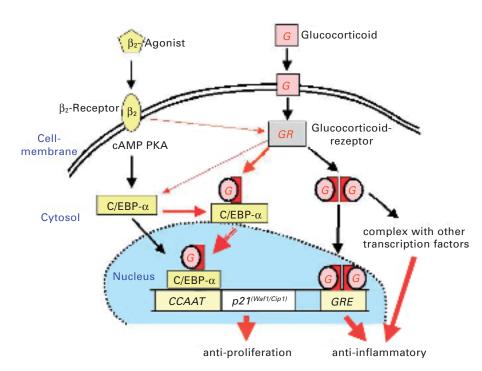
In order to control the inflammatory aspect of asthma the standard treatment includes glucocorticoids, which are among the most potent anti-inflammatory drugs available. Glucocorticoids act via their intracellular receptor, the glucocorticoid receptor, which is located in the cytosol and upon ligand binding is activated, forms a dimer and migrates into the nucleus where it acts as a transcription factor [70, 71]. The glucocorticoid receptor acts most often as an inhibitor of gene transcription but the exact conditions for this action remain to be defined [72]. In addition, the activated glucocorticoid receptor can form complexes with

other transcription factors including IkB, C/EBPs, Stats, and AP-1 [73]. Thereby glucocorticoids may affect genes that do not contain a glucocorticoid receptor binding DNA sequence in their regulatory (promoter) region. A schema of the signalling pathway is provided in figure 2.

Under glucocorticoid therapy the inflammation of the mucosa is significantly decreased and asthma symptoms are reduced. Airway hyperresponsiveness is also significantly reduced with inhaled glucocorticoids. Therefore it is believed that airway hyper-responsiveness is a direct consequence of mucosal inflammation. However, also

#### Figure 2

The interaction of the glucocorticoid and  $\beta_2$ -agonist signalling pathway and its consequences for the anti-inflammatory and the anti-proliferative effect of the two drugs. As C/EBP- $\alpha$  is missing in airway smooth muscle cells of asthma patients, the anti-proliferative effect of especially glucocorticoids cannot be effective. However, the anti-inflammatory action of glucocorticoids, mediated via the glucocorticoid receptor dimer and the GRE-promoter element as well as via the interaction of the GR with other transcription factors, is fully functional.



glucocorticoids are not capable of curing asthma and symptoms most often re-occur after cessation of treatment. From these observations it can be speculated that mucosal inflammation cannot be the only cause of asthma.

In addition to their anti-inflammatory action glucocorticoids are capable to block cell proliferation. This is of special interest since this might offer a possibility to control the increased mass of airway smooth muscle cell bundles in asthma as shown in figure 1. In an animal model it had been shown that the anti-proliferative effect of glucocorticoids involves the formation of a complex with the transcription factor C/EBP- $\alpha$  (figure 2), which we were able to confirm in human lung cells [74]. C/EBP-α belongs to a family of transcription factors that, like the glucocorticoid receptor, has been preserved during evolution, and both the glucocorticoid receptor and the C/EBP-isoproteins are central regulators for cell proliferation. Knockout animal models have been tested for both the glucocorticoid receptor and C/EBP-α, but the -/homozygous animals for either of the two factors did not survive birth longer than a few hours [75, 76]. We think that natural hormones (steroids) together with the transcription factors C/EBPs and peroxisome proliferator-activated receptor (PPAR) regulate cell homeostasis in the human body [77, 78]. Fibroblast and smooth muscle cell proliferation in the human lung is mainly controlled by the ratio of C/EBP-α to C/EBP-β. We reported recently that bronchial smooth muscle cells obtained from asthma patients lack the ex-

Th1/Th2 mucosal smooth muscle inflammation shift abnormality healthy negative negative negative atopy positive positive negative asthma positive positive positive

pression of C/EBP- $\alpha$  and therefore glucocorticoids cannot inhibit their proliferation [79]. This might explain why bronchial smooth muscle cells from asthma patients grow faster in culture than cells from COPD patients or controls [29]. The lack of C/EBP- $\alpha$  which was documented in the smooth muscle cells of all asthma patients of our cohort should not be confused with steroid resistant asthma. A mutation of the glucocorticoid receptor is usually the basis of steroid resistant asthma, which occurs in a minority of asthma patients [73, 75].

There is increasing evidence that the combination of glucocorticoids with long-acting  $\beta_2$ -agonists controls asthma symptoms better than an increase of the glucocorticoid dosage [for review see 80–82]. This clinically documented benefit of the combination of glucocorticoids with long acting β<sub>2</sub>-agonists opposes the initially feared "pro-inflammatory action" of  $\beta_2$ -agonists [83, 84]. It was previously thought that the use of long acting  $\beta_2$ agonists initially improved clinical symptoms, followed by non-compliance of patients to inhale steroids. The latter increased again the underlying inflammation of the bronchial mucosa. We postulated earlier that the clinically significant beneficial effect of the combination of glucocorticoids with long acting β<sub>2</sub>-agonists is based on their molecular biological effect. When both classes of drugs are combined, they synchronise the activation of the two transcription factors, glucocorticoid receptor and C/EBP-α [85].

We demonstrated earlier that  $\beta_2$ -agonists activate the glucocorticoid receptor in a ligand independent – yet to be explained – manner in human lung fibroblasts and smooth muscle cells [86]. This finding has recently been confirmed in patients after inhalation of either a glucocorticoid or a  $\beta_2$ -agonist alone or in combination, as shown by immuno-histochemistry in epithelial cells [87]. How-

Table 1.

The comparison of the clinical characteristics of asthma and atopic patients shows only in asthma patients the three following parameters: sub-mucosal inflammation, Th1-Th2 shift, and an abnormality of the smooth muscle cells bundles.

ever, the synchronisation of the two transcription factors might not be the only cellular signalling pathway that is modified by the combined classes of drugs [88]. The new concept of the interaction of glucocorticoids and  $\beta_2$ -agonists regarding their

effect on inflammation and proliferation is shown in figure 2. In conclusion, the combination of glucocorticoids and  $\beta_2$ -agonists shows a beneficial effect in asthma treatment, which can be explained via their molecular biological interaction.

#### The role of airway smooth muscle cells in asthma

The increased mass of smooth muscle cell bundles in the airway of asthma patients (figure 1) is an important feature, which has long been underestimated. Even in young asthma patients and in patients with mild asthma the smooth muscle cell mass is increased as recently shown by endobronchial biopsies obtained via bronchoscopy [22-28, 36, 89, 90]. This increased mass of muscle is believed to contribute to bronchial hyper-reactivity. After being able to culture for the first time bronchial smooth muscle cells of asthma patients it became evident that the smooth muscle cells of these patients behave abnormal [29, 79]. Even after long-term culture the smooth muscle cells of asthma patients proliferated at a faster rate than their controls, including COPD patients [29, 79].

Therefore inflammatory mediators present in the body can be excluded as a cause for this pathology. However, it cannot be excluded that chronic stimuli such as allergens or viruses might interfere with the C/EBP-α expression We have shown earlier that Chlamydia pneumoniae, which is known to induce asthma exacerbations, affects the above described signalling pathway by "high checking" the glucocorticoid receptor and NFkB signalling pathway [91]. Preliminary data suggest that several respiratory viruses directly alter the activity of various C/EBP-isoproteins (unpublished data). Furthermore, our group demonstrated that smooth muscle cells of asthma patients produce more connective tissue growth factor (CTGF), which could be related to the increased expression of TGF-β the increased synthesis of extracellular matrix [92, 93] and the increased synthesis of some specific extracellular matrix components [28] that contribute to airway wall thickening. In this context it is important to note that the composition of the extracellular matrix is a key regulator of airway smooth muscle cell proliferation [28, 31, 94, 95].

As described above, the ratio of the proproliferative C/EBP- $\beta$  to the anti-proliferative C/EBP- $\alpha$  controls natural cell homeostasis. The increased proliferation of the airway smooth muscle is due to the lack of C/EBP- $\alpha$ . As described above, C/EBP- $\alpha$  forms a complex with the glucocorticoid receptor and is involved in the control of cell proliferation and differentiation [74, 85]. Therefore the normal anti-proliferative action of C/EBP- $\alpha$  is missing in smooth muscle cells of asthma patients [79]. Re-introduction of C/EBP- $\alpha$  by transfection experiments restored the sensitivity of the cells for the anti-proliferative effect of glucocorticoids [79]. The family of the C/EBP tran-

scription factors consists of six members (-α, -β,  $-\gamma$ ,  $-\delta$ ,  $-\varepsilon$ ,  $-\zeta$ ) in humans and they all can interact with each other and bind to the same DNA promoter sequence, CCAAT [96]. Furthermore, at least some C/EBPs can bind to the cAMP response element (CRE), thereby affecting the expression of much more genes [97]. In other cell types C/EBPs have been identified as central regulators of cell differentiation [98–100]. We speculate that the lack of C/EBP-α in the smooth muscle cells of asthma patients leads to a dysbalance with the pro-proliferative C/EBP-β [101], which explains their faster proliferation rate [29]. Interestingly, overexpression of C/EBP-β in lung epithelial cells has recently been reported in COPD patients [86]. In contrast to C/EBP-α enhanced expression of C/EBP-β and furthermore its activation is clearly linked to proliferation and de-differentiation of several cell types [100, 101]. The recent publication of Didon et al. [102] suggests that this mechanism might play a role in the pathogenesis of small airway bronchiolitis with fibroblast proliferation and collagen deposition in the small airways. The role of C/EBPs in asthma has been discussed in details earlier [103].

Furthermore, the lack of C/EBP- $\alpha$  and its role in cell differentiation may explain the often-described trans-differentiation of fibroblast or smooth muscle cells into so called myo-fibroblasts observed in asthma [104]. If the lack of C/EBP- $\alpha$  in smooth muscle cells of asthma patients leaves the cells with an "over-activity" of the remaining C/EBP- $\beta$  the report of Hu et al. [88] showing that C/EBP- $\beta$  regulates the expression of smooth muscle cell actin becomes important; the expression of smooth muscle cell actin is often used to differentiate between smooth muscle cells and myofibroblasts [105].

Further indications that C/EBP-isoproteins control the differentiation of fibroblast like cells into more specific differentiated cell types have been provided for C/EBP-β and the differentiation of mouse fibroblasts into adipocytes [106], and for C/EBP-δ and the transdifferentiation of myofibroblasts in the kidney [107]. The role of the different C/EBP-isoforms on the differentiation of smooth muscle cells and in asthma has therefore to be studied in more detail. The function of C/EBPs is also linked to that of the transcription factor peroxisome proliferation receptor activator, PPAR, which forms complexes with the glucocorticoid receptor and C/EBP-isoforms [77, 108]. This signalling pathway and its impact on asthma and COPD is under investigation.

#### Conclusion

The new findings on abnormal smooth muscle cell behaviour in asthma have challenged the hypothesis of asthma pathogenesis. Not only inflammation must be present in the asthmatic airway but also a defect of smooth muscle cell differentiation. Current and future research has to

address the underlying molecular biological mechanisms of the smooth muscle cell defect. Of special interest are the questions, how epithelial cells can control smooth muscle cell behaviour, and how allergens and viruses can influence this interaction.

#### References

- 1 Sunyer J, Anto JM, Tobias A, Burney P. Generational increase of self-reported first attack of asthma in fifteen industrialized countries. European Community Respiratory Health Study (ECRHS). Eur Respir J 1999;14:885–91.
- 2 Huovinen E, Kaprio J, Laitinen LA, Koskenvuo M. Incidence and prevalence of asthma among adult Finnish men and women of the Finnish Twin Cohort from 1975 to 1990, and their relation to hay fever and chronic bronchitis. Chest 1999:115:928–36.
- 3 Dey AN, Schiller JS, Tai DA. Summary health statistics for U.S. children: National Health Interview Survey, 2002. Vital Health Stat 10. 2004;221:1–78.
- 4 Poulos LM, Toelle BG, Marks GB. The burden of asthma in children: an Australian perspective. Paediatr Respir Rev 2005;6:20–7.
- 5 Ellwood P, Asher MI, Beasley R, Clayton TO, Stewart AW; ISAAC Steering Committee. The international study of asthma and allergies in childhood (ISAAC): phase three rationale and methods. Int J Tuberc Lung Dis 2005;9:10–6.
- 6 Lawson JA, Senthilselvan A. Asthma epidemiology: has the crisis passed? Curr Opin Pulm Med 2005;111:79–84.
- 7 Woolcock AJ, Bastiampillai SA, Marks GB, Keena VA. The burden of asthma in Australia. Med J Aust 2001;175:141–5.
- 8 Sailly JC, Lenne X, Bercez C, Lebrun T, Tonnel AB, Tillie-Leblond I. Costs of hospitalization for severe acute asthma of patients not treated according to guidelines and recommendations French prospective study of 169 cases. Eur J Health Econ 2005,11;[Epub ahead of print] PMID:15761774.
- 9 Stock S, Redaelli M, Luengen M, Wendland G, Civello D, Lauterbach KW. Asthma: prevalence and cost of illness. Eur Respir J 2005;25:47–53.
- 10 Chen FH, Samson KT, Miura K, Ueno K, Odajima Y, Shougo T, Yoshitsugu Y, Shioda S. Airway remodeling: a comparison between fatal and nonfatal asthma. J Asthma 2004;4:631–8.
- 11 Hamid Q, Song Y, Kotsimbos TC, Minshall E, Bai TR, Hegele RG, Hogg JC. Inflammation of small airways in asthma. J Allergy Clin Immunol 1997;100:44–51.
- 12 Kuwano K, Bosken CH, Pare PD, Bai TR, Wiggs BR, Hogg JC. Small airways dimensions in asthma and in chronic obstructive pulmonary disease. Am Rev Respir Dis 1993;148: 1220–5.
- 13 Fedorov IA, Wilson SJ, Davies DE, Holgate ST. Epithelial stress and structural remodelling in childhood asthma. Thorax 2005;60:389–94.
- 14 Shebani E, Shahana S, Janson C, Roomans GM; BHR group. Attachment of columnar airway epithelial cells in asthma. Tissue Cell 2005;37:145–52.
- 15 Hashimoto M, Tanaka H, Abe S. Quantitative analysis of bronchial wall vascularity in the medium and small airways of patients with asthma and COPD. Chest 2005;127:965–72.
- 16 Martinez FJ, Standiford C, Gay SE. Is it asthma or COPD? The answer determines proper therapy for chronic airflow obstruction. Postgrad Med 2005;117:19–26.
- 17 McKnight J, Scott A, Menzies D, Bourbeau J, Blais L, Lemiere C. A cohort study showed that health insurance databases were accurate to distinguish chronic obstructive pulmonary disease from asthma and classify disease severity. J Clin Epidemiol 2005;58:206–8.
- 18 Shore SA. Airway smooth muscle in asthma not just more of the same. N Engl J Med 2004;351:531–2.
- 19 Woodruff PG, Khashayar R, Lazarus SC, Janson S, Avila P, Boushey HA, Segal M, Fahy JV. Relationship between airway inflammation, hyperresponsiveness, and obstruction in asthma. J Allergy Clin Immunol 2001;108:753–8.

- 20 Woodruff PG, Fahy JV. Airway remodeling in asthma. Semin Respir Crit Care Med 2002;23:361–8.
- 21 Kotlikoff MI, Kannan MS, Solway J, Deng KY, Deshpande DA, Dowell M, et al. Methodologic advancements in the study of airway smooth muscle. J Allergy Clin Immunol 2004;114 (2 Suppl):S18–31.
- 22 Benayoun L, Druilhe A, Dombret MC, Aubier M, Pretolani M. Airway structural alterations selectively associated with severe asthma. Am J Respir Crit Care Med 2003;167:1360–8.
- 23 Woodruff PG, Dolganov GM, Ferrando RE, Donnelly S, Hays SR, Solberg OD, et al. Hyperplasia of smooth muscle in mild to moderate asthma without changes in cell size or gene expression. Am J Respir Crit Care Med 2004;169:1001–6.
- 24 Hirst SJ, Martin JG, Bonacci JV, Chan V, Fixman ED, Hamid QA, et al. Proliferative aspects of airway smooth muscle. J Allergy Clin Immunol 2004;114(2 Suppl):S2–17.
- 25 Ammit AJ, Panettieri RAJr. Airway smooth muscle cell hyperplasia: a therapeutic target in airway remodeling in asthma? Prog Cell Cycle Res 2003;5:49–57.
- 26 Zhou L, Li J, Goldsmith AM, Newcomb DC, Giannola DM, Vosk RG, et al. Human bronchial smooth muscle cell lines show a hypertrophic phenotype typical of severe asthma. Am J Respir Crit Care Med 2004;169:703–11.
- 27 Howarth PH, Knox AJ, Amrani Y, Tliba O, Panettieri RA Jr, Johnson M. Synthetic responses in airway smooth muscle. J Allergy Clin Immunol 2004;114(2 Suppl):S32–50.
- 28 Johnson PR, Burgess JK, Underwood PA, Au W, Poniris MH, Tamm M, Ge Q, Roth M, Black JL. Extracellular matrix proteins modulate asthmatic airway smooth muscle cell proliferation via an autocrine mechanism. J Allergy Clin Immunol 2004:113:690–6.
- 29 Johnson PR, Roth M, Tamm M, Hughes M, Ge Q, King G, et al. Airway smooth muscle cell proliferation is increased in asthma. Am J Respir Crit Care Med 2001;164:474–7.
- 30 Chung KF, Sterk PJ. The airway smooth muscle cell: a major contributor to asthma? Eur Respir J 2000;15:438–9.
- 31 Halayko AJ, Amrani Y. Mechanisms of inflammation-mediated airway smooth muscle plasticity and airways remodeling in asthma. Respir Physiol Neurobiol 2003;137:209–22.
- 32 Black JL. Asthma more muscle cells or more muscular cells? Am J Respir Crit Care Med 2004;169:980–1.
- 33 Panettieri RA Jr. Airway smooth muscle: immunomodulatory cells that modulate airway remodeling? Respir Physiol Neurobiol 2003;137:277–93.
- 34 Hirst SJ. Regulation of airway smooth muscle cell immunomodulatory function: role in asthma. Respir Physiol Neurobiol 2003;137:309–26.
- 35 Stephens NL, Li W, Jiang H, Unruh H, Ma X. The biophysics of asthmatic airway smooth muscle. Respir Physiol Neurobiol 2003;137:125–40.
- 36 Aoshiba K, Nagai A. Differences in airway remodeling between asthma and chronic obstructive pulmonary disease. Clin Rev Allergy Immunol 2004;27:35–43.
- 37 Moir LM, Leung SY, Eynott PR, McVicker CG, Ward JP, Chung KF, Hirst SJ. Repeated allergen inhalation induces phenotypic modulation of smooth muscle in bronchioles of sensitized rats. Am J Physiol Lung Cell Mol Physiol 2003;284: L148–59.
- 38 Moir LM, Ward JP, Hirst SJ. Contractility and phenotype of human bronchiole smooth muscle after prolonged fetal bovine serum exposure. Exp Lung Res 2003;29:339–59.
- 39 Polosa R, Li Gotti F, Mangano G, Paolino G, Mastruzzo C, Vancheri C, et al. Effect of immunotherapy on asthma progression, BHR and sputum eosinophils in allergic rhinitis. Allergy 2004;59:1224–8.

- 40 Wadonda-Kabondo N, Sterne JA, Golding J, Kennedy CT, Archer CB, Dunnill MG; ALSPAC Study Team. Association of parental eczema, hayfever, and asthma with atopic dermatitis in infancy: birth cohort study. Arch Dis Child 2004;89: 917–21.
- 41 Ciprandi G, Cirillo I, Vizzaccaro A, Milanese M, Tosca MA. Airway function and nasal inflammation in seasonal allergic rhinitis and asthma. Clin Exp Allergy 2004;34:891–6.
- 42 Huurre TM, Aro HM, Jaakkola JJ. Incidence and prevalence of asthma and allergic rhinitis: a cohort study of Finnish adolescents. J Asthma 2004;41:311–7.
- 43 Bousquet J, Van Cauwenberge P, Khaltaev N; Aria Workshop Group; World Health Organization. Allergic rhinitis and its impact on asthma. J Allergy Clin Immunol 2001;108(5 Suppl): S147–334.
- 44 Leynaert B, Neukirch C, Kony S, Guenegou A, Bousquet J, Aubier M, Neukirch F. Association between asthma and rhinitis according to atopic sensitization in a population-based study. J Allergy Clin Immunol 2004;113:86–93.
- 45 Bousquet J, Boushey HA, Busse WW, Canonica GW, Durham SR, Irvin CG, et al. Characteristics of patients with seasonal allergic rhinitis and concomitant asthma. Clin Exp Allergy 2004;34:897–903.
- 46 Asher I, Baena-Cagnani C, Boner A, Canonica GW, Chuchalin A, Custovic A, et al.; World Allergy Organization. World Allergy Organization guidelines for prevention of allergy and allergic asthma. Int Arch Allergy Immunol 2004;135: 83–97
- 47 O'Byrne PM, Inman MD, Adelroth E. Reassessing the Th2 cytokine basis of asthma. Trends Pharmacol Sci 2004;25: 244-8
- 48 Hartl D, Griese M, Nicolai T, Zissel G, Prell C, Konstantopoulos N, et al. Pulmonary chemokines and their receptors differentiate children with asthma and chronic cough. J Allergy Clin Immunol 2005;115:728–36.
- 49 Cho SH, Stanciu LA, Holgate ST, Johnston SL. Increased interleukin-4, interleukin-5, and interferon-gamma in airway CD4<sup>+</sup> and CD8<sup>+</sup> T cells in atopic asthma. Am J Respir Crit Care Med 2005;171:224–30.
- 50 Shakoory B, Fitzgerald SM, Lee SA, Chi DS, Krishnaswamy G. The role of human mast cell-derived cytokines in eosinophil biology. J Interferon Cytokine Res 2004;24:271–81.
- 51 Cruse G, Kaur D, Yang W, Duffy SM, Brightling CE, Bradding P. Activation of human lung mast cells by monomeric immunoglobulin E. Eur Respir J 2005;25:858–63.
- 52 Balzar S, Chu HW, Strand M, Wenzel S. Relationship of small airway chymase-positive mast cells and lung function in severe asthma. Am J Respir Crit Care Med 2005;171:431–9.
- 53 Crummy F, Livingston M, Ennis M, Heaney LG. Mast cell mediator release in nonasthmatic subjects after endobronchial adenosine challenge. J Allergy Clin Immunol 2004;114:34–9.
- 54 Cruse G, Kaur D, Yang W, Duffy SM, Brightling CE, Bradding P. Activation of human lung mast cells by monomeric immunoglobulin E. Eur Respir J 2005;25:858–63.
- 55 Robinson DS. The role of the mast cell in asthma: induction of airway hyperresponsiveness by interaction with smooth muscle? J Allergy Clin Immunol 2004;114:58–65.
- 56 Norris A. Targeting mast cells. Expert Opin Investig Drugs 2004;13:739–41.
- 57 Marone G, Triggiani M, de Paulis A. Mast cells and basophils: friends as well as foes in bronchial asthma? Trends Immunol 2005;26:25–31.
- 58 Holgate ST, Djukanovic R, Casale T, Bousquet J. Anti-immunoglobulin E treatment with omalizumab in allergic diseases: an update on anti-inflammatory activity and clinical efficacy. Clin Exp Allergy 2005;35:408–16.
- 59 Brownell J, Casale TB. Anti-IgE therapy. Immunol Allergy Clin North Am 2004;24:51–68.
- 60 Kips JC, O'Connor BJ, Langley SJ, Woodcock A, Kerstjens HA, Postma DS, et al. Effect of SCH55700, a humanized antihuman interleukin-5 antibody, in severe persistent asthma: a pilot study. Am J Respir Crit Care Med 2003;167:1655–9.
- 61 Hayashi T, Gong X, Rossetto C, Shen C, Takabayashi K, Redecke V, et al. Induction and inhibition of the th2 phenotype spread: implications for childhood asthma. J Immunol 2005;174:5864–73.
- 62 Ishiura Y, Fujimura M, Nobata K, Myou S, Oribe Y, Abo M, Nakamura H. Th2 cytokine inhibition and cough in asthmatic and bronchitic patients. Ann Med 2004;36:623–9.
- 63 Caramori G, Ito K, Adcock IM. Targeting Th2 cells in asthmatic airways. Curr Drug Targets Inflamm Allergy 2004;3: 243–55.

- 64 Shi HZ, Li S, Xie ZF, Qin XJ, Qin X, Zhong XN. Regulatory CD4\*CD25\*T lymphocytes in peripheral blood from patients with atopic asthma. Clin Immunol 2004:113:172–8.
- 65 Hoshino A, Tsuji T, Matsuzaki J, Jinushi T, Ashino S, Teramura T, et al. STAT6-mediated signaling in Th2-dependent allergic asthma: critical role for the development of eosinophilia, airway hyper-responsiveness and mucus hypersecretion, distinct from its role in Th2 differentiation. Int Immunol 2004;16:1497–505.
- 66 Shore SA. Direct effects of Th2 cytokines on airway smooth muscle. Curr Opin Pharmacol 2004;4:235–40.
- 67 Zhu Y, Chen L, Huang Z, Alkan S, Bunting KD, Wen R, Wang D, Huang H. Cutting edge: IL-5 primes Th2 cytokine-producing capacity in eosinophils through a STAT5-dependent mechanism. J Immunol 2004;173:2918–22.
- 68 Ostroukhova M, Ray A. CD25\* T cells and regulation of allergen-induced responses. Curr Allergy Asthma Rep 2005;5: 35–41.
- 69 Kuipers H, Lambrecht BN. The interplay of dendritic cells, Th2 cells and regulatory T cells in asthma. Curr Opin Immunol 2004;16:702–8.
- 70 Schoneveld OJ, Gaemers IC, Lamers WH. Mechanisms of glucocorticoid signalling. Biochim Biophys Acta 2004;1680: 114–28.
- 71 Lu NZ, Cidlowski JA. The origin and functions of multiple human glucocorticoid receptor isoforms. Ann N Y Acad Sci 2004;1024:102–23.
- 72 Dostert A, Heinzel T. Negative glucocorticoid receptor response elements and their role in glucocorticoid action. Curr Pharm Des 2004;10:2807–16.
- 73 Schulz M, Eggert M. Novel ligands: fine tuning the transcriptional activity of the glucocorticoid receptor. Curr Pharm Des 2004:10:2817–26.
- 74 Rüdiger JJ, Roth M, Bihl MP, Cornelius BC, Johnson M, Ziesche R, Block LH. Interaction of C/EBPalpha and the glucocorticoid receptor in vivo and in nontransformed human cells. FASEB J 2002;16:177–84.
- 75 Cole TJ, Myles K, Purton JF, Brereton PS, Solomon NM, Godfrey DI, Funder JW. GRKO mice express an aberrant dexamethasone-binding glucocorticoid receptor, but are profoundly glucocorticoid resistant. Mol Cell Endocrinol 2001; 173:193–202.
- 76 Wang ND, Finegold MJ, Bradley A, Ou CN, Abdelsayed SV, Wilde MD, et al. Impaired energy homeostasis in C/EBP alpha knockout mice. Science 1995;269:1108–12.
- 77 Nie M, Corbett L, Knox AJ, Pang L. Differential regulation of chemokine expression by peroxisome proliferator-activated receptor gamma agonists: interactions with glucocorticoids and beta2-agonists. J Biol Chem 2005;280:2550–61.
- 78 Benayoun L, Letuve S, Druilhe A, Boczkowski J, Dombret MC, Mechighel P, et al. Regulation of peroxisome proliferator-activated receptor gamma expression in human asthmatic airways: relationship with proliferation, apoptosis, and airway remodeling. Am J Respir Crit Care Med 2001;164:1487–94.
- 79 Roth M, Johnson PR, Borger P, Bihl MP, Rudiger JJ, King GG, et al. Dysfunctional interaction of C/EBPalpha and the gluco-corticoid receptor in asthmatic bronchial smooth-muscle cells. N Engl J Med 2004;351:560–74.
- 80 Lemiere C, Becker A, Boulet LP, Bowie D, Cartier A, Cockroft D, et al. Should combination therapy with inhaled corticosteroids and long-acting beta2-agonists be prescribed as initial maintenance treatment for asthma? CMAJ 2002;167: 1008–9.
- 81 Barnes PJ. Clinical outcome of adding long-acting beta-agonists to inhaled corticosteroids. Respir Med 2001;95:S12–6.
- 82 Chung KF, Adcock IM. Combination therapy of long-acting beta2-adrenoceptor agonists and corticosteroids for asthma. Treat Respir Med 2004;3:279–89.
- 83 Shore SA, Drazen JM. Beta-agonists and asthma: too much of a good thing? J Clin Invest 2003;112:495–7.
- 84 Jackson CM, Lipworth B. Benefit-risk assessment of long-acting beta2-agonists in asthma. Drug Saf 2004;27:243–70.
- 85 Roth M, Johnson PR, Rudiger JJ, King GG, Ge Q, Burgess JK, et al. Interaction between glucocorticoids and beta2 agonists on bronchial airway smooth muscle cells through synchronised cellular signalling. Lancet 2002;360:1293–9.
- 86 Eickelberg O, Roth M, Lorx R, Bruce V, Rudiger J, Johnson M, Block LH. Ligand-independent activation of the glucocorticoid receptor by beta2-adrenergic receptor agonists in primary human lung fibroblasts and vascular smooth muscle cells. J Biol Chem 1999;274:1005–10.

- 87 Usmani OS, Ito K, Maneechotesuwan K, Ito M, Johnson M, Barnes PJ, Adcock IM. Glucocorticoid Receptor Nuclear Translocation in Airway Cells Following Inhaled Combination Therapy. Am J Respir Crit Care Med 2005 [Epub ahead of print] PMID: 15860753
- 88 Tantisira KG, Small KM, Litonjua AA, Weiss ST, Liggett SB. Molecular properties and pharmacogenetics of a polymorphism of adenylyl cyclase 9 in asthma: interaction between {beta}-agonist and corticosteroid pathways. Hum Mol Genet 2005;14:1671–7.
- 89 Niimi A, Matsumoto H, Takemura M, Ueda T, Chin K, Mishima M. Relationship of airway wall thickness to airway sensitivity and airway reactivity in asthma. Am J Respir Crit Care Med 2003;168:983–8.
- 90 Payne DN, Rogers AV, Adelroth E, Bandi V, Guntupalli KK, Bush A, Jeffery PK. Early thickening of the reticular basement membrane in children with difficult asthma. Am J Respir Crit Care Med 2003;167:78–82.
- 91 Gencay MM, Tamm M, Glanville A, Perruchoud AP, Roth M. Chlamydia pneumoniae activates epithelial cell proliferation via NF-kappaB and the glucocorticoid receptor. Infect Immun 2003;71:5814–22.
- 92 Burgess JK, Johnson PR, Ge Q, Au WW, Poniris MH, Mc-Parland BE, et al. Expression of connective tissue growth factor in asthmatic airway smooth muscle cells. Am J Respir Crit Care Med 2003;167:71–7.
- 93 Black JL, Burgess JK, Johnson PR. Airway smooth muscle its relationship to the extracellular matrix. Respir Physiol Neurobiol 2003;137:339–46.
- 94 Hirst SJ, Twort CH, Lee TH. Differential effects of extracellular matrix proteins on human airway smooth muscle cell proliferation and phenotype. Am J Respir Cell Mol Biol 2000;23: 335-44
- 95 Nguyen TT, Ward JP, Hirst SJ. beta1-Integrins mediate enhancement of airway smooth muscle proliferation by collagen and fibronectin. Am J Respir Crit Care Med 2005;171:217–23.
- 96 Ramji DP, Foka P. CCAAT/enhancer-binding proteins: structure, function and regulation. Biochem J 2002;365:561–75.
- 97 Wilson HL, Roesler WJ. CCAAT/enhancer binding proteins: do they possess intrinsic cAMP-inducible activity? Mol Cell Endocrinol 2002;188:15–20.

- 98 Gregoire FM. Adipocyte differentiation: from fibroblast to endocrine cell. Exp Biol Med 2001;226:997–1002.
- 99 Rosmarin AG, Yang Z, Resendes KK. Transcriptional regulation in myelopoiesis: Hematopoietic fate choice, myeloid differentiation, and leukemogenesis. Exp Hematol 2005;33: 131–43.
- 100 Zahnow CA. CCAAT/enhancer binding proteins in normal mammary development and breast cancer. Breast Cancer Res 2002;4:113–21.
- 101 Buck M, Chojkier M. Signal transduction in the liver: C/EBPbeta modulates cell proliferation and survival. Hepatology 2003;37:731–8.
- 102 Didon L, Qvarfordt I, Andersson O, Nord M, Riise GC. Decreased CCAAT/enhancer binding protein transcription factor activity in chronic bronchitis and COPD. Chest 2005;127:1341–6.
- 103 Borger P, Black JL, Roth M. Asthma and the CCAAT-enhancer binding proteins: a holistic view on airway inflammation and remodeling. J Allergy Clin Immunol 2002;110:841–6.
- 104 Schmidt M, Sun G, Stacey MA, Mori L, Mattoli S. Identification of circulating fibrocytes as precursors of bronchial myofibroblasts in asthma. J Immunol 2003;171:380–9.
- 105 Hu B, Wu Z, Jin H, Hashimoto N, Liu T, Phan SH. CCAAT/enhancer-binding protein beta isoforms and the regulation of alpha-smooth muscle actin gene expression by IL-1 beta. J Immunol 2004;173:4661–8.
- 106 Park BH, Qiang L, Farmer SR. Phosphorylation of C/EBPbeta at a consensus extracellular signal-regulated kinase/glycogen synthase kinase 3 site is required for the induction of adiponectin gene expression during the differentiation of mouse fibroblasts into adipocytes. Mol Cell Biol 2004;24: 8671–80.
- 107 M, Kawada N, Moriyama T, Nagatoya K, Oseto S, Akira S, Hori M, et al. CCAAT/Enhancer-binding protein delta contributes to myofibroblast transdifferentiation and renal disease progression. J Am Soc Nephrol 2004;15:2383–90.
- 108 Farmer SR. Regulation of PPARgamma activity during adipogenesis. Int J Obes Relat Metab Disord 2005;29:S13–6.



## The many reasons why you should choose SMW to publish your research

What Swiss Medical Weekly has to offer:

- SMW's impact factor has been steadily rising, to the current 1.537
- Open access to the publication via the Internet, therefore wide audience and impact
- Rapid listing in Medline
- LinkOut-button from PubMed with link to the full text website http://www.smw.ch (direct link from each SMW record in PubMed)
- No-nonsense submission you submit a single copy of your manuscript by e-mail attachment
- Peer review based on a broad spectrum of international academic referees
- Assistance of our professional statistician for every article with statistical analyses
- Fast peer review, by e-mail exchange with the referees
- Prompt decisions based on weekly conferences of the Editorial Board
- Prompt notification on the status of your manuscript by e-mail
- Professional English copy editing
- No page charges and attractive colour offprints at no extra cost

#### Editorial Board

Prof. Jean-Michel Dayer, Geneva

Prof. Peter Gehr, Berne

Prof. André P. Perruchoud, Basel

Prof. Andreas Schaffner, Zurich

(Editor in chief)

Prof. Werner Straub, Berne

Prof. Ludwig von Segesser, Lausanne

#### International Advisory Committee

Prof. K. E. Juhani Airaksinen, Turku, Finland Prof. Anthony Bayes de Luna, Barcelona, Spain

Prof. Hubert E. Blum, Freiburg, Germany

Prof. Walter E. Haefeli, Heidelberg, Germany

Prof. Nino Kuenzli, Los Angeles, USA

Prof. René Lutter, Amsterdam,

The Netherlands

Prof. Claude Martin, Marseille, France

Prof. Josef Patsch, Innsbruck, Austria

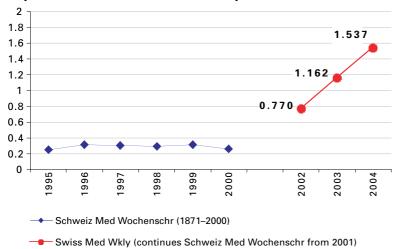
Prof. Luigi Tavazzi, Pavia, Italy

We evaluate manuscripts of broad clinical interest from all specialities, including experimental medicine and clinical investigation.

We look forward to receiving your paper!

Guidelines for authors: http://www.smw.ch/set\_authors.html

#### Impact factor Swiss Medical Weekly





All manuscripts should be sent in electronic form, to:

EMH Swiss Medical Publishers Ltd. SMW Editorial Secretariat Farnsburgerstrasse 8 CH-4132 Muttenz

Manuscripts: Letters to the editor: Editorial Board: Internet: submission@smw.ch letters@smw.ch red@smw.ch http://www.smw.ch