

# Proteinases as hormone-like signal messengers

**Proteinase-activated receptors and the pathophysiology of inflammation, pain, cardiovascular disease and cancer**

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## Summary

Proteinases like thrombin and trypsin, long known for their ability to activate the coagulation cascade or to act as hormone-processing enzymes, are now recognised as hormone-like regulators of cell function. These serine proteinases activate cell signalling by triggering a novel four-member family of G-protein-coupled receptors, termed Proteinase-Activated Receptors (PARs). This review article summarises historically the discovery of PARs as well as their unique mechanism of activation and outlines a number of different pathophysiological settings in which PARs can act to regulate cell and tissue function. PARs can be seen to

play a role in pathophysiological processes ranging from inflammation and pain to cardiovascular disease and cancer. Apart from activating PARs to cause their physiological effects in tissues, proteinases can also mediate cell signalling via a number of other mechanisms, including the activation of growth factor receptors, like the one for insulin. Therefore, this article also describes the non-PAR mechanisms whereby proteinases can have hormone-like actions in cells and tissues.

*Key words: hormone; information; PAR; protease; proteinase; receptor; signal transduction; thrombin*

## Introduction

The serine proteinase thrombin is well recognised for its role in the coagulation cascade. In addition to its ability to act as a clotting factor, thrombin has long been known to trigger signalling pathways in platelets and endothelial cells. For more than forty years, in addition to their ability to convert inactive pro-hormone precursors to their active forms (eg, pro-insulin to insulin), other serine proteinases, such as trypsin, have also been known to stimulate cellular hormone-like responses. For instance, work by the Riesers in the mid-1960s documented the insulin-like actions of proteinases like pepsin and chymotrypsin in a rat diaphragm preparation [1, 2]. Subsequent work in the early 1970s showed that like insulin, trypsin, can both stimulate glucose oxidation and inhibit lipolysis in isolated adipocyte preparations [3]. Over the past fifteen years or so, the mechanisms responsible for the cellular actions of proteinases have come into focus. In large part, the physiological actions of serine proteinases can be seen to be mediated by a novel family of G-protein-coupled receptors: the Proteinase-Activated Receptors (PARs). Other

mechanisms that mediate the actions of proteinases will also be discussed below.

### Thrombin, platelet activation and the discovery of proteinase-activated receptors

The search for the receptor on human platelets and hamster lung fibroblasts responsible for the ability of thrombin to initiate platelet aggregation and to stimulate fibroblast mitogenesis resulted in the cloning of a receptor that turned out to be a member of the G-protein-coupled receptor super-family [4-9]. It was discovered that the unique mechanism of activation of this receptor involves the proteolytic unmasking of an N-terminal receptor

#### Abbreviations:

Cha: cyclohexylalanine

Cit: Citrulline

IUPHAR: International Union of Pharmacology

PAR: Proteinase-Activated Receptor

PAR-AP: PAR-Activating Peptide

SAR: Structure-Activity Relationship

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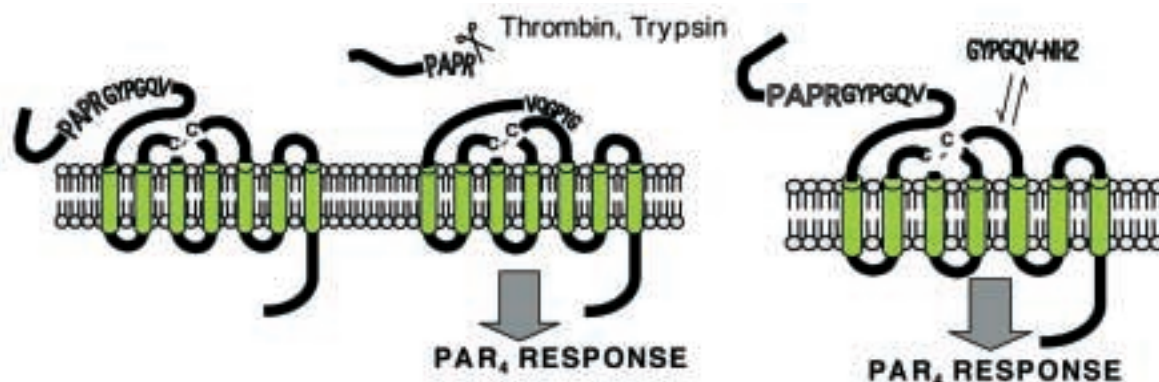
sequence that becomes a tethered ligand, which binds to the extracellular receptor domains to trigger receptor signalling [5] (figure 1). Based on this mechanism of activation, the receptor for thrombin has been referred to as a “Proteinase-Activated Receptor” and assigned the acronym “PAR” by the International Union of Pharmacology [8]. The first PAR found to be a target for thrombin has now been designated as PAR<sub>1</sub>.

Remarkably, it was also discovered that synthetic

peptides with sequences matching that of the exposed tethered ligand can also activate the receptor in the absence of proteolysis [5]. Thus, a synthetic peptide, beginning with the sequence of human PAR<sub>1</sub>, SFLLRN ..., was found to be a surrogate activator of the receptor for thrombin in a variety of settings. These peptides (initially termed Thrombin Receptor-Activating Peptides or TRAPs, which mimic the ability of thrombin to activate PAR<sub>1</sub>, soon revealed that in certain cells,

**Figure 1**

Mechanism of activation of Proteinase-Activated Receptors, as typified by human PAR<sub>4</sub>. The figure illustrates the activation of human PAR<sub>4</sub> both by the tethered ligand mechanism triggered by thrombin or trypsin (left and centre) and by a PAR<sub>4</sub>-activating peptide (GYPGQV-NH<sub>2</sub>) with a sequence based on the proteolytically-revealed tethered ligand (right).



**Table 1**

The PAR family of G-protein-coupled receptors.\*

Receptor designation (IUPHAR)	tethered ligand sequence	comment
PAR <sub>1</sub>	(h) SFLLRN...	designated: TRAP or PAR <sub>1</sub> AP
	(r, m) SFLLRN...	SFLLRN... Activates both PAR <sub>1</sub> and PAR <sub>2</sub> standard PAR <sub>1</sub> -activating peptide: TFLLR-NH <sub>2</sub> standard PAR <sub>1</sub> -inactive peptide: FTLLR-NH <sub>2</sub>
PAR <sub>2</sub>	(h) SLIGKV...	designated PAR <sub>2</sub> AP
	(r, m) SLIGRL ...	standard PAR <sub>2</sub> -activating peptide: SLIGRL-NH <sub>2</sub> standard PAR <sub>2</sub> -inactive peptide: LSIGRL-NH <sub>2</sub> selectively activates only PAR <sub>2</sub> ; murine and rat sequence more potent than human sequence
PAR <sub>3</sub>	(h) TFRGAP...	PAR <sub>3</sub> is not activated by PAR-APs
	(m) SFNGGP...	PAR <sub>3</sub> -derived sequences, eg, TFRGAP... or SFNGGP... , activate both PAR <sub>1</sub> and PAR <sub>2</sub>
PAR <sub>4</sub>	(h) GYPGQV...	designated PAR <sub>4</sub> AP
	(m) GYPGKF...	standard PAR <sub>4</sub> -activating peptide: AYPGKF-NH <sub>2</sub>
	(r) GFPGKP...	standard PAR <sub>4</sub> -inactive peptide YAPGKF-NH <sub>2</sub> PAR <sub>4</sub> AP sequences do not activate PARs 1 and 2, but are active via non-PAR <sub>4</sub> receptors in some bioassays

\* Abbreviations: h = human; m = mouse; r = rat; IUPHAR = International Union of Pharmacology. The new N-terminal sequences revealed by serine proteinase cleavage are shown as tethered ligands. These proteolytically revealed sequences activate signalling in PARs 1, 2 and 4, but not in PAR<sub>3</sub>.

**Table 2**

Unique features of PARs.

Feature	comment
Can be proteolytically activated by a number of serine proteinases	multiple circulating or local agonists are possible
Can be proteolytically cleaved downstream of the tethered ligand sequence thereby preventing receptor activation by agonist serine proteinases	multiple circulating or secreted enzymes, like neutrophil elastase can act as antagonists by disarming PARs
Susceptibility to proteinase activation can be modulated by receptor glycosylation near the tethered ligand cleavage site	tryptase does not activate fully glycosylated human PAR <sub>2</sub>
PAR-APs can mimic proteolytic activation of signalling	receptors other than PARs may be activated, even by PAR-selective PAR-APs that do not activate other PARs

such as rodent platelets, the PAR<sub>1</sub>-Activating Peptide (PAR<sub>1</sub>-AP) did not cause a thrombin response (eg, aggregation) [10]. Other structure-activity studies with peptides based on the SFLLRN sequence also pointed to subtypes of the thrombin receptor in rat vascular and gastric tissues [11].

Subsequent to the cloning of the first receptor for thrombin (PAR<sub>1</sub>), three other members of this intriguing receptor family have been identified (table 1). Each of these G-protein-coupled receptors, designated PARs 1 to 4, has a unique N-terminal tethered ligand sequence that is revealed by serine proteinase action as illustrated for PAR<sub>4</sub> in figure 1 and summarised in table 1. PARs 1, 3, and 4 have been found to be targets for thrombin, whereas PAR<sub>2</sub>, not readily activated by thrombin, can be activated by trypsin, tryptase and by other serine proteinase members of the clotting cascade apart from thrombin (eg, tissue factor-VIIa-Xa complex) [6-9, 12]. Although the signalling properties of PAR<sub>3</sub> are unclear, all of PARs 1, 2 and 4 have been found to signal via a G-protein coupled mechanism involving G $\alpha_i$  or G $\alpha_q$ . Further, based on the revealed tethered ligand sequences of PARs 1, 2 and 4, it has now been possible to design synthetic peptides (PAR-APs) that can selectively activate each receptor. Appropriate standard inactive peptides, incapable of activating the PARs, are also known (table 1).

Although PARs can be activated by a variety of serine proteinases using the tethered-ligand mechanism outlined in Figure 1, it is also the case that the cleavage of a PAR N-terminal sequence downstream of the tethered ligand portion would disarm the receptor, thus preventing its subsequent

activation by a proteinase. For instance, the elastase secreted by *Pseudomonas aeruginosa*, a complicating pathogen in the setting of cystic fibrosis, can cleave and remove the tethered ligand sequence from PAR<sub>2</sub>, thereby disabling the receptor on lung epithelial cells [13]. The disabling of PAR<sub>2</sub> in this setting may contribute to the pathophysiology of lung inflammation in this disease. Thus, PARs can be said to have a variety of circulating agonists (ie, serine proteinases that reveal the tethered ligand) as well as circulating functional antagonists that can disarm them downstream of their tethered ligands, thereby silencing the receptors. That said, the proteolytically disarmed receptors would still be sensitive to activation by the PAR-APs that do not depend on the tethered ligand sequence for receptor activation.

The unique features of PARs are summarised in table 2. One of the key features of these receptors is their ability to be activated by receptor-selective PAR-APs. These PAR-APs have proved to be of considerable utility to determine the potential consequences of activating PARs in bioassay systems *in vitro* or in inflammatory or other animal models *in vivo*. As summarised in the following sections, PARs have been found to play an important role in the pathophysiology of diseases ranging from inflammation and pain to cardiovascular disease and cancer. For a comprehensive collection of articles dealing with PARs and their potential impact on physiological function, the reader is invited to access the special issues of Drug Development Research (volumes 59 [4] and 60 [1]) to be found on the following website: <http://www.inflammation-calgary.com>.

## Discovering pathophysiological roles for PARs: a pharmacological approach

### PAR-APs trigger both PAR and non-PAR responses in target tissues: use of structure-activity studies

As alluded to above, Structure-Activity Relationship (SAR) studies using peptides with sequences based on human PAR<sub>1</sub> revealed the presence of a receptor other than PAR<sub>1</sub> in an endothelium-dependent rat aorta relaxation assay [11]. That receptor, unknown at the time, turned out to be PAR<sub>2</sub> [14, 15]. The principle that led to the discovery of functional PAR<sub>2</sub> in the rat vascular endothelium was outlined some time ago by Ahlquist [16] in defining the pharmacology of alpha- and beta-adrenoceptors. In essence, with only minor exceptions, a receptor can be typified for distinct responses in different tissues by the relative potencies (EC<sub>50</sub>s or IC<sub>50</sub>s) of a series of chemically related agonists and antagonists. The presence of distinct SAR relationships for the same set of compounds (eg, agonists) in different tissue assays points to the existence of distinct receptors.

This principle has been used to advantage in study-

ing potential PAR-mediated responses in different bioassay systems, employing, for example, a series of PAR<sub>1</sub> and PAR<sub>2</sub>APs. Thus, for PAR<sub>2</sub>-mediated calcium signalling in a PAR<sub>2</sub>-expressing KNRK cell line, the relative potencies of the PAR<sub>2</sub>-selective agonist peptides, SLIGRL-NH<sub>2</sub>, trans-cinnamoyl-LIGRLO-NH<sub>2</sub>, 2-furoyl-LIGRO-NH<sub>2</sub> and of a potent PAR<sub>1</sub>-selective PAR<sub>1</sub>AP, AparafuoroFRChaChaCitY-NH<sub>2</sub> would be: 2-furoyl-LIGRO-NH<sub>2</sub> >> trans-cinnamoyl-LIGRLO-NH<sub>2</sub>  $\approx$  SLIGRL-NH<sub>2</sub> >>> AparafuoroF-RChaChaCitY-NH<sub>2</sub> [17, 18]. A completely reversed SAR would be expected of a PAR<sub>1</sub>-mediated response, in which the three PAR<sub>2</sub>-activating peptides would be essentially inactive. Surprisingly, the SAR relationship for these PAR agonists observed in a rat jejunal ion transport assay (SLIGRL-NH<sub>2</sub> > trans-cinnamoyl-LIGRLO-NH<sub>2</sub> > AparafuoroFRChaChaCitY-NH<sub>2</sub>) was different from the SAR expected of either PAR<sub>2</sub> or PAR<sub>1</sub> [17]. A plausible conclusion was that the short-circuit current response in the jejunal Ussing chamber due to the

serosal application of the PAR-APs and trypsin was mediated by a receptor different from PAR<sub>1</sub> and PAR<sub>2</sub>. In a similar manner, some recent work with PAR<sub>4</sub>-derived agonists has been able to verify the presence of PAR<sub>4</sub> in rat platelets, using a platelet aggregation assay, while pointing to a non-PAR<sub>4</sub>-mediated response in a rat gastric longitudinal muscle assay [19].

The work with the PAR<sub>4</sub>-derived peptides illustrates that a judicious choice of standard PAR-APs as well as a standard PAR-inactive peptide is required to establish whether or not a given response can be attributed to a given PAR. For responses thought to be mediated by PAR<sub>1</sub>, use can be made of receptor antagonists (eg, RWJ56110 or SCH79797) [20, 21]. Although not yet available for PAR<sub>2</sub>, antagonists have been developed for PAR<sub>4</sub> [22, 23]. That said, although the peptide PAR<sub>4</sub> antagonists are suitable for antagonizing the receptor in platelets, these antagonists can cause responses via receptors other than PAR<sub>4</sub> in tissue assays [19]. To resolve such discrepancies, PAR-deficient mice have been used to demonstrate unequivocally the PAR-related actions of PAR-APs and to prove that a proteinase-triggered response may be due to the activation of one or more of the PARs (below). The essence of these findings with the PAR-APs is that it is now possible with reasonable confidence to use these receptor probes to assess the potential impact that PAR activation might have in a variety of physiological and pathological settings.

### PARs, inflammation, neuronal responses and nociception

Although PARs 1 and 4 were discovered primarily due to the search for the target of thrombin on mammalian platelets, the potential physiological role for PAR<sub>2</sub> was not known at the time of its discovery [24]. However, the use of selective PAR<sub>2</sub>-APs as probes for PAR<sub>2</sub> function quickly revealed a potential role for this receptor in regulating vascular and gastric smooth muscle tension

[14, 15]. It came as a surprise, however, that the administration of small doses of either a PAR<sub>1</sub> or a PAR<sub>2</sub>-AP caused marked swelling and leukocyte infiltration in a rat paw oedema model of inflammation [25, 26]. At that time, it was also observed that functional PAR<sub>2</sub> as well as PAR<sub>1</sub> could be localised on neuronal elements [27]. Putting these two sets of observations together, it has become evident that the inflammatory response triggered by PARs 1 and 2 is mediated via a neurogenic mechanism [28, 29]. The administration of a PAR<sub>4</sub>-AP also causes the formation of oedema and leukocyte recruitment in a rat paw model of inflammation [19, unpublished results]. However, in contrast with PARs 1 and 2, these PAR<sub>4</sub>-mediated events are not dependent on a neurogenic mechanism [19]. It has also become clear that in addition to triggering the inflammatory response, PARs also play a role in sensing pain [30–33]. Given the wide distribution of PARs on neurons and their associated cells, such as astrocytes, both in the central and peripheral nervous systems, it is to be expected that neuronal PARs may play a widespread physiological role. As an example, one can point to an up-regulation of PAR<sub>1</sub> in the central nervous system in the setting of HIV encephalitis [34]. Further, PAR<sub>2</sub> would appear to play a neuroprotective role in the setting of HIV infection [35].

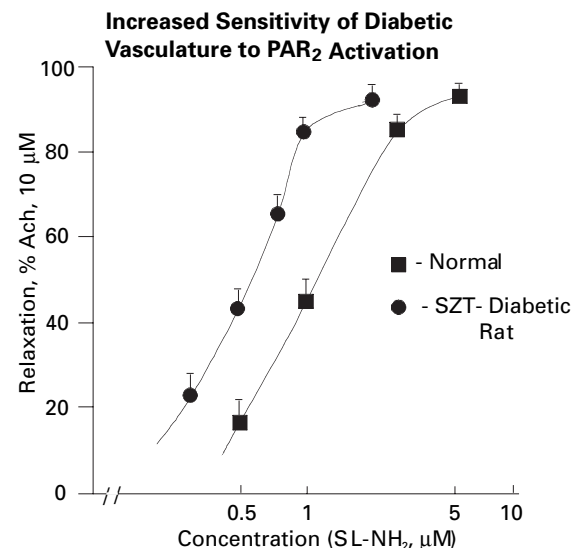
The overarching working hypothesis that can be put forward is that PARs play a key role in the body's innate defense, as a primary trigger of the inflammatory response and pain sensation due to tissue injury or remodeling caused by pathogenic processes. This hypothesis is strongly supported by the striking resistance of PAR<sub>2</sub>-deficient mice to adjuvant-induced arthritis [36].

### PARs and cardiovascular function

An isolated rat aorta tissue preparation provided one of the first bioassay systems that enabled us to predict a role for PARs in regulating vascular function [14, 15, 37]. Using the same approach, it was possible to document the ability of PAR<sub>2</sub> as well as

**Figure 2**

Vasorelaxation of rat aorta from normal and streptozotocin diabetic rats following PAR<sub>2</sub> activation. Male Sprague-Dawley rats (about 200 g) were either treated or not with streptozotocin (SZT), to render treated animals diabetic. One week after streptozotocin treatment, with hyperglycaemia established, animals were sacrificed and endothelium-intact aorta rings were monitored for PAR<sub>2</sub>-mediated vasodilatation as described previously [14], using SLIGRL-NH<sub>2</sub> (SL-NH<sub>2</sub>) as a receptor agonist. Relaxation in the normal (■) or SZT-diabetic (●) aorta rings by increasing concentrations of SL-NH<sub>2</sub> was expressed as a percentage of the relaxation caused in the same preparation by 10  $\mu$ M acetylcholine (% Ach). Values showing the shift to the left in sensitivity of the SZT-diabetic tissues represent the averages ( $\pm$  sem) for measurements done with three independent tissue preparations. (Saifeddine and Hollenberg, unpublished)



**Table 3**

Potential physiological roles for PARs.

Potential role	comment
Platelet activation, haemostasis: Thrombin-activated receptors (PARs 1, 3, 4)	PARs regulate both secretion and aggregation; PARs 1 and 4 can play separate roles
Endothelial cell function: (PARs 1, 2 and 4)	release of NO, von Willebrand factor; increased neutrophil adherence; cell migration
Vascular smooth muscle function	activation of contractility; angiogenesis?
Intestinal function: (PARs 1, 2 and 4)	regulation of motility (GI smooth muscle) and secretion (GI epithelial cell)
Myenteric neuron function	also affects GI motility and inflammatory response
Renal vascular function	regulation of flow and afferent arteriolar function
CNS neuronal and astrocyte function	up-regulation of PARs in the setting of CNS inflammation
Response to joint injury	key role for PAR <sub>2</sub> in arthritis
Tumour cell growth and metastasis	both PARs 1 and 2 may play roles, activated by tumour-derived serine proteinases and Matrix metalloproteinases (eg, MMP-1)

PAR<sub>1</sub> to activate an endothelium-dependent, nitric oxide (NO)-mediated vasorelaxation. In contrast with PAR<sub>2</sub> which does not appear to regulate vascular smooth muscle function directly, PAR<sub>1</sub>, activated by either a PAR<sub>1</sub>-selective AP such as TFLLR-NH<sub>2</sub> or by thrombin, causes a prompt vasoconstriction. In the setting of renal function, both PARs 1 and 2 can have an effect on perfusion, with PAR<sub>1</sub> activation causing a profound decrease in flow similar to angiotensin and with PAR<sub>2</sub> acting as a vasodilator to increase flow [38]. Thus, in certain settings, PARs 1 and 2 may play a bi-directional role. Although in conduit vessels like the aorta PAR activation leads primarily to an NO-mediated relaxation, in resistance vessels or in renal afferent arterioles, vasodilatation caused by PAR activation is mediated not only by NO, but also by as yet unidentified endothelium-derived relaxing factors (EDHFs) [39, 40]. The impact of PAR<sub>4</sub> activation on vascular function is not yet clear, except for its ability to play a potential role for endothelium-leukocyte interactions [41].

A potential role for PAR<sub>2</sub> in the setting of cardiovascular disease may occur in the setting of ischaemia-reperfusion in which case there can be an up-regulation of PAR<sub>2</sub> to promote vasodilatation [42]. It has also been found that PAR<sub>2</sub> is increased in human coronary atherosclerotic lesions [43]. Moreover, a preliminary assessment of aorta tissue derived from rats rendered diabetic by streptozotocin treatment indicates an increased sensitivity to the vasodilatory actions of PAR<sub>2</sub> (Figure 2). The distinct effects on blood pressure and heart rate upon activating either PAR<sub>1</sub> (both hypotension

and tachycardia) or PAR<sub>2</sub> (hypotension only, without an effect on heart rate) have been established unequivocally with the use of mice deficient in either PAR<sub>1</sub> or PAR<sub>2</sub> [44]. Thus, a generalised role for the PARs in the setting of cardiovascular pathophysiology would appear to be plausible.

#### PARs, cancer and metastasis

Since the mid 1990s, it has been suggested that the coagulation system in general and thrombin specifically may play an important role in tumour growth and metastasis [45, 46]. Not only might thrombin facilitate the ability of tumour cells to migrate through the basement membrane, but the enzyme itself has been known for some time to be a particularly potent mitogen for normal as well as tumour-derived cells, presumably acting via PAR<sub>1</sub>. A clear link has been made between the expression of PAR<sub>1</sub> in mammary tumour-derived cells and the ability of the cells to migrate in culture through a reconstituted basement membrane [47]. The ability of PAR<sub>1</sub> to subserve a role in tumour metastasis and invasion is underlined by the ability of tumour-derived matrix metalloproteinase-1 to activate the receptor and drive the process of migration and metastasis of breast carcinoma cells in a xenograft model [48]. A comparable role for PAR<sub>2</sub> in the setting of cancer would not be unexpected [49]. Given the information provided in the previous sections, it is clear that in addition to contributing to the growth and metastasis of tumour cells, PARs can potentially play a role in a wide variety of pathophysiological processes, as summarised in Table 3.

## Which endogenous proteinases regulate PAR activity?

### Thrombin and other serine proteinases

Given the physiological role established for thrombin as a member of the coagulation cascade, it is clear that this serine proteinase is a key regulator of PARs 1 and 4, with the amplification pro-

vided by PAR<sub>3</sub> [6]. That said, the tissue-localised proteinases other than thrombin that may regulate PARs, including PARs 2 and 4, which are both potentially activated by trypsin (and presumably other serine proteinases), have yet to be identified. In the

intestinal tract, it has been suggested that trypsin itself (presumably pancreatic trypsin-1 in humans) is responsible for activating PAR<sub>2</sub> on the intestinal epithelium [50]. Mast cell tryptase, which in humans might be released in the vicinity of sensory nerves, is another candidate enzyme that may regulate PAR<sub>2</sub> [51–53]. Of importance is the potential role that other serine proteinases of the coagulation cascade (Factor VIIa/Xa) may play [12], especially in terms of activating PAR<sub>2</sub>, a receptor that, as outlined above, plays a prominent role in inflammatory and nociceptive settings.

Given that PARs 1 and 2 have been shown to play an important role in models of inflammatory bowel disease [54], a major question to ask is: what intestinal proteinases at the site of inflammation

might trigger the inflammatory response? A clue to answering this question has come from a study using a model of murine infectious colitis, in which the infecting organism, *C. rodentium*, induces the production of PAR<sub>2</sub>-activating serine proteinases (members of the trypsin family and granzyme A) that in turn activate PAR<sub>2</sub> [55]. Significantly, the oral administration of a serine proteinase inhibitor (soya trypsin inhibitor) was able to attenuate the pathogen-induced PAR<sub>2</sub>-mediated colitis [55]. Thus, the identification of site-produced PAR-activating proteinases and the selective targeting of proteinase inhibitors to individual tissues may provide an interesting therapeutic modality for treating a number of inflammatory disorders that may involve PARs.

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## Proteinase signalling by mechanisms other than PARs

### Regulation of growth factor receptors

As mentioned above, one of the first indications that proteinases can activate cellular signals comparable to those of hormones came from the observations in the early 1960s that trypsin exhibits an insulin-like action in rat diaphragm tissue [1]. This hormone-like action of trypsin in striated muscle and adipocytes [3] cannot be attributed to the activation of PARs, but is rather due to the effect of trypsin on the receptor for insulin. By cleaving at a di-basic residue of the insulin receptor  $\alpha$ -subunit, trypsin generates a truncated receptor that has intrinsic signalling activity [56]. In principle, this kind of action of proteinases, either activating or disarming growth factor receptors (eg, at higher concentrations, trypsin can abolish the ability of the insulin receptor to bind insulin: [57]) can modulate cell function in a variety of settings, for example via the IGF-I receptor. Another proteolytic mechanism that can lead to the activation of a growth factor receptor involves the proteolytic generation of a growth factor agonist in the cell environment. For instance, the trans-activation of the EGF receptor can result from the metalloproteinase-mediated release from the cell surface of a receptor agonist (heparin-binding EGF) [58]. In this regard, thrombin, apart from signalling via the

PARs can also yield chemotactic-mitogenic peptides from proteolytic processing of its non-catalytic domain [59–61]. These thrombin-derived peptides cause their effects via receptors that are not PARs. Thrombin can also potentially cause its cellular effects via the activation of pro-metalloproteinase [62]. In addition to generating active peptide hormones from recognised pro-hormone precursors (eg, pro-insulin) that in turn activate receptors, novel receptor-activating hormone-like agonists can be generated from precursors in the vicinity of target receptors. For instance, interleukin-beta is generated by the interleukin-beta converting enzyme (ICE), a cysteine proteinase that also plays a role intracellularly in the apoptotic process [63]. Thus, proteinases can play a signalling role not only by receptor modulation and ligand generation, but also by regulating intracellular signalling pathways such as the one responsible for the apoptotic response. Hence, apart from activating or inactivating PARs, proteinases can play hormone-like signalling roles in a variety of cellular settings via non-PAR mechanisms. This diversity of hormone-like roles played by proteinases is exceeded only by the diversity of the proteinase families themselves.

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## Conclusions

This article has summarised the various hormone-like roles that proteinases can play, not only by activating or silencing members of a unique G-protein-coupled receptors family, the proteinase-activated receptors (PARs), but also by regulating the activity of growth factor receptors, like the one for insulin. Apart from these receptor-mediated signal pathways, proteinases can generate novel recep-

tor-activating agonists and can regulate intracellular signal transduction pathways using mechanisms that can be added to their recognised ability to generate peptide hormones from pro-hormone precursors. These signalling properties of proteinases add a novel dimension to the biological significance of this enzyme superfamily. Thus, the targeting of proteinases with tissue site-selective

and enzyme-specific inhibitors may prove of therapeutic benefit in a variety of pathophysiological settings ranging from inflammation and pain to cardiovascular disease and cancer.

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## References

- Rieser P, Rieser CH. Anabolic responses of diaphragm muscle to insulin and to other pancreatic proteins. *Proc Soc Exp Biol Med* 1964;116:669-71.
- Rieser P. The insulin-like action of pepsin and pepsinogen. *Acta Endocrinol (Copenh)* 1967;54(2):375-9.
- Kono T, Barham FW. Insulin-like effects of trypsin on fat cells. Localization of the metabolic steps and the cellular site affected by the enzyme. *J Biol Chem* 1971;246(20):6204-9.
- Rasmussen UB, Vouret-Craviari V, Jallat S, Schlesinger Y, Pages G, Pavirani A, et al. cDNA cloning and expression of a hamster alpha-thrombin receptor coupled to Ca<sup>2+</sup> mobilization. *FEBS Lett* 1991;288(1-2):123-8.
- Vu TK, Hung DT, Wheaton VI, Coughlin SR. Molecular cloning of a functional thrombin receptor reveals a novel proteolytic mechanism of receptor activation. *Cell* 1991;64(6):1057-68.
- Coughlin SR. Thrombin signalling and protease-activated receptors. *Nature* 2000;407(6801):258-64.
- Macfarlane SR, Seatter MJ, Kanke T, Hunter GD, Plevin R. Proteinase-activated receptors. *Pharmacol Rev* 2001;53(2):245-82.
- Hollenberg MD, Compton SJ. International Union of Pharmacology. XXVIII. Proteinase-activated receptors. *Pharmacol Rev* 2002;54(2):203-17.
- Ossovskaya VS, Bunnett NW. Protease-activated receptors: contribution to physiology and disease. *Physiol Rev* 2004;84(2):579-621.
- Kinlough-Rathbone RL, Rand ML, Packham MA. Rabbit and rat platelets do not respond to thrombin receptor peptides that activate human platelets. *Blood* 1993;82(1):103-6.
- Hollenberg MD, Lanionu AA, Saifeddine M, Moore GJ. Role of the amino- and carboxyl-terminal domains of thrombin receptor-derived polypeptides in biological activity in vascular endothelium and gastric smooth muscle: evidence for receptor subtypes. *Mol Pharmacol* 1993;43(6):921-30.
- Ruf W, Dorfleutner A, Riewald M. Specificity of coagulation factor signaling. *J Thromb Haemost* 2003;1(7):1495-503.
- Dulon S, Leduc D, Cottrell GS, D'Alayer J, Hansen KK, Bunnett NW, et al. Pseudomonas aeruginosa elastase disables proteinase-activated receptor 2 in respiratory epithelial cells. *Am J Respir Cell Mol Biol* 2005;32(5):411-9.
- Al-Ani B, Saifeddine M, Hollenberg MD. Detection of functional receptors for the proteinase-activated-receptor-2-activating polypeptide, SLIGRL-NH<sub>2</sub>, in rat vascular and gastric smooth muscle. *Can J Physiol Pharmacol* 1995;73(8):1203-7.
- Saifeddine M, Al-Ani B, Cheng CH, Wang L, Hollenberg MD. Rat proteinase-activated receptor-2 (PAR-2): cDNA sequence and activity of receptor-derived peptides in gastric and vascular tissue. *Br J Pharmacol* 1996;118(3):521-30.
- Ahlquist RP. A study of the adrenotropic receptors. *Am J Physiol* 1948;153: 586-600.
- Vergnolle N, MacNaughton W, Al-Ani B, Saifeddine M, Wallace JL, Hollenberg MD. Proteinase-activated receptor 2 (PAR2)-activating peptides: identification of a receptor distinct from PAR2 that regulates intestinal transport. *Proc Natl Acad Sci USA* 1998;95(13):7766-71.
- McGuire JJ, Saifeddine M, Triggler CR, Sun K, Hollenberg MD. 2-furoyl-LIGRLO-amide: a potent and selective proteinase-activated receptor 2 agonist. *J Pharmacol Exp Ther* 2004;309(3):1124-31.
- Hollenberg MD, Saifeddine M, Sandhu S, Houle S, Vergnolle N. Proteinase-activated receptor-4: evaluation of tethered ligand-derived peptides as probes for receptor function and as inflammatory agonists in vivo. *Br J Pharmacol* 2004;143(4):443-54.
- Andrade-Gordon P, Maryanoff BE, Derian CK, Zhang HC, Addo MF, Darrow AL, et al. Design, synthesis, and biological characterization of a peptide-mimetic antagonist for a tethered-ligand receptor. *Proc Natl Acad Sci USA* 1999;96(22):12257-62.
- Ahn HS, Foster C, Boykow G, Stamford A, Manna M, Graziano M. Inhibition of cellular action of thrombin by N<sup>3</sup>-cyclopropyl-7-[[4-(1-methylethyl)phenyl]methyl]-7H-pyrrolo[3,2-f]quinazoline-1,3-diamine (SCH 79797), a nonpeptide thrombin receptor antagonist. *Biochem Pharmacol* 2000;60(10):1425-34.
- Hollenberg MD, Saifeddine M. Proteinase-activated receptor 4 (PAR4): activation and inhibition of rat platelet aggregation by PAR4-derived peptides. *Can J Physiol Pharmacol* 2001;79(5):439-42.
- Covic L, Misra M, Badar J, Singh C, Kuliopulos A. Pepducin-based intervention of thrombin-receptor signaling and systemic platelet activation. *Nat Med* 2002;8(10):1161-5.
- Nystedt S, Emilsson K, Wahlestedt C, Sundelin J. Molecular cloning of a potential proteinase activated receptor. *Proc Natl Acad Sci USA* 1994;91(20):9208-12.
- Vergnolle N, Hollenberg MD, Wallace JL. Pro- and anti-inflammatory actions of thrombin: a distinct role for proteinase-activated receptor-1 (PAR1). *Br J Pharmacol* 1999;126(5):1262-8.
- Vergnolle N, Hollenberg MD, Sharkey KA, Wallace JL. Characterization of the inflammatory response to proteinase-activated receptor-2 (PAR2)-activating peptides in the rat paw. *Br J Pharmacol* 1999;127(5):1083-90.
- Corvera CU, Dery O, McConalogue K, Gamp P, Thoma M, Al-Ani B, et al. Thrombin and mast cell tryptase regulate guinea-pig myenteric neurons through proteinase-activated receptors-1 and -2. *J Physiol* 1999;517 ( Pt 3):741-56.
- Steinhoff M, Vergnolle N, Young SH, Tognetto M, Amadesi S, Ennes HS, et al. Agonists of proteinase-activated receptor 2 induce inflammation by a neurogenic mechanism. *Nat Med* 2000;6(2):151-8.
- de Garavilla L, Vergnolle N, Young SH, Ennes H, Steinhoff M, Ossovskaya VS, et al. Agonists of proteinase-activated receptor 1 induce plasma extravasation by a neurogenic mechanism. *Br J Pharmacol* 2001;133(7):975-87.
- Vergnolle N, Bunnett NW, Sharkey KA, Brussee V, Compton SJ, Grady EF, et al. Proteinase-activated receptor-2 and hyperalgesia: A novel pain pathway. *Nat Med* 2001;7(7):821-6.
- Vergnolle N, Wallace JL, Bunnett NW, Hollenberg MD. Protease-activated receptors in inflammation, neuronal signaling and pain. *Trends Pharmacol Sci* 2001;22(3):146-52.
- Asfaha S, Brussee V, Chapman K, Zochodne DW, Vergnolle N. Proteinase-activated receptor-1 agonists attenuate nociception in response to noxious stimuli. *Br J Pharmacol* 2002;135(5):1101-6.
- Vergnolle N. Modulation of visceral pain and inflammation by protease-activated receptors. *Br J Pharmacol* 2004;141(8):1264-74.

- 34 Boven LA, Vergnolle N, Henry SD, Silva C, Imai Y, Holden J, et al. Up-regulation of proteinase-activated receptor 1 expression in astrocytes during HIV encephalitis. *J Immunol* 2003; 170(5):2638–46.
- 35 Noorbakhsh F, Vergnolle N, McArthur JC, Silva C, Vodjgani M, Andrade-Gordon P, et al. Proteinase-Activated Receptor-2 Induction by Neuroinflammation Prevents Neuronal Death during HIV Infection. *J Immunol* 2005;174(11):7320–9.
- 36 Ferrell WR, Lockhart JC, Kelso EB, Dunning L, Plevin R, Meek SE, et al. Essential role for proteinase-activated receptor-2 in arthritis. *J Clin Invest* 2003;111(1):35–41.
- 37 Muramatsu I, Laniyonu A, Moore GJ, Hollenberg MD. Vascular actions of thrombin receptor peptide. *Can J Physiol Pharmacol* 1992;70(7):996–1003.
- 38 Gui Y, Loutzenhiser R, Hollenberg MD. Bidirectional regulation of renal hemodynamics by activation of PAR1 and PAR2 in isolated perfused rat kidney. *Am J Physiol Renal Physiol* 2003;285(1):F95–104.
- 39 McGuire JJ, Hollenberg MD, Bennett BM, Triggle CR. Hyperpolarization of murine small caliber mesenteric arteries by activation of endothelial proteinase-activated receptor 2. *Can J Physiol Pharmacol* 2004;82(12):1103–12.
- 40 Wang X, Hollenberg MD, Loutzenhiser R. Redundant signaling mechanisms contribute to the vasodilatory response of the afferent arteriole to proteinase-activated receptor-2. *Am J Physiol Renal Physiol* 2005;288(1):F65–75.
- 41 Vergnolle N, Derian CK, D'Andrea MR, Steinhoff M, Andrade-Gordon P. Characterization of thrombin-induced leukocyte rolling and adherence: a potential proinflammatory role for proteinase-activated receptor-4. *J Immunol* 2002;169(3):1467–73.
- 42 Napoli C, Cicala C, Wallace JL, de Nigris F, Santagada V, Caliendo G, et al. Protease-activated receptor-2 modulates myocardial ischemia-reperfusion injury in the rat heart. *Proc Natl Acad Sci USA* 2000;97(7):3678–83.
- 43 Napoli C, de Nigris F, Wallace JL, Hollenberg MD, Tajana G, De Rosa G, et al. Evidence that protease activated receptor 2 expression is enhanced in human coronary atherosclerotic lesions. *J Clin Pathol* 2004;57(5):513–6.
- 44 Damiano BP, Cheung WM, Santulli RJ, Fung-Leung WP, Ngo K, Ye RD, et al. Cardiovascular responses mediated by protease-activated receptor-2 (PAR-2) and thrombin receptor (PAR-1) are distinguished in mice deficient in PAR-2 or PAR-1. *J Pharmacol Exp Ther* 1999;288(2):671–8.
- 45 Nierodzik ML, Chen K, Takeshita K, Li JJ, Huang YQ, Feng XS, et al. Protease-activated receptor 1 (PAR-1) is required and rate-limiting for thrombin-enhanced experimental pulmonary metastasis. *Blood* 1998;92:3694–3700.
- 46 Henrikson KP, Salazar SL, Fenton JW 2nd, Pentecost BT. Role of thrombin receptor in breast cancer invasiveness. *Br J Cancer* 1999;79:401–406.
- 47 Even-Ram S, Uziely B, Cohen P, Grisaru-Granovsky S, Maoz M, Ginzburg Y, et al. Thrombin receptor overexpression in malignant and physiological invasion processes. *Nat Med* 1998; 4(8):909–14.
- 48 Boire A, Covic L, Agarwal A, Jacques S, Sherif S, Kuliopulos A. PAR1 is a matrix metalloprotease-1 receptor that promotes invasion and tumorigenesis of breast cancer cells. *Cell* 2005; 120(3):303–13.
- 49 Shi X, Gangadharan B, Brass LF, Ruf W, Mueller BM. Protease-activated receptors (PAR1 and PAR2) contribute to tumor cell motility and metastasis. *Mol Cancer Res* 2004;2(7):395–402.
- 50 Kong W, McConalogue K, Khitin LM, Hollenberg MD, Payan DG, Bohm SK, et al. Luminal trypsin may regulate enterocytes through proteinase-activated receptor 2. *Proc Natl Acad Sci USA* 1997;94(16):8884–9.
- 51 Corvera CU, Dery O, McConalogue K, Bohm SK, Khitin LM, Caughey GH, et al. Mast cell tryptase regulates rat colonic myocytes through proteinase-activated receptor 2. *J Clin Invest* 1997;100(6):1383–93.
- 52 Mirza H, Schmidt VA, Derian CK, Jesty J, Bahou WF. Mitogenic responses mediated through the proteinase-activated receptor-2 are induced by expressed forms of mast cell alpha- or beta-tryptases. *Blood* 1997;90(10):3914–22.
- 53 Molino M, Barnathan ES, Numerof R, Clark J, Dreyer M, Cumashi A, et al. Interactions of mast cell tryptase with thrombin receptors and PAR-2. *J Biol Chem* 1997;272(7):4043–9.
- 54 Vergnolle N. Clinical relevance of proteinase activated receptors (pars) in the gut. *Gut* 2005;54(6):867–74.
- 55 Hansen KK, Sherman PM, Cellars L, Andrade-Gordon P, Pan Z, Baruch A, et al. A major role for proteolytic activity and proteinase-activated receptor-2 in the pathogenesis of infectious colitis. *Proc Natl Acad Sci USA* 2005;102:8363–8.
- 56 Shoelson SE, White MF, Kahn CR. Tryptic activation of the insulin receptor. Proteolytic truncation of the alpha-subunit releases the beta-subunit from inhibitory control. *J Biol Chem* 1988;263(10):4852–60.
- 57 Cuatrecasas P. Perturbation of the insulin receptor of isolated fat cells with proteolytic enzymes. *J Biol Chem* 1971;246(21): 6522–31.
- 58 Prenzel N, Zwick E, Daub H, Leserer M, Abraham R, Wallasch C, et al. EGF receptor transactivation by G-protein-coupled receptors requires metalloproteinase cleavage of proHB-EGF. *Nature* 1999;402(6764):884–8.
- 59 Bar-Shavit R, Kahn A, Mudd MS, Wilner GD, Mann KG, Fenton JW 2nd. Localization of a chemotactic domain in human thrombin. *Biochemistry* 1984;23(3):397–400.
- 60 Bar-Shavit R, Kahn AJ, Mann KG, Wilner GD. Identification of a thrombin sequence with growth factor activity on macrophages. *Proc Natl Acad Sci USA* 1986;83(4):976–80.
- 61 Glenn KC, Frost GH, Bergmann JS, Carney DH. Synthetic peptides bind to high-affinity thrombin receptors and modulate thrombin mitogenesis. *Pept Res* 1988;1(2):65–73.
- 62 Lafleur MA, Hollenberg MD, Atkinson SJ, Knauper V, Murphy G, Edwards DR. Activation of pro-(matrix metalloproteinase-2) by thrombin is membrane-type-MMP-dependent in human umbilical vein endothelial cells and generates a distinct 63 kDa active species. *Biochem J* 2001; 357(Pt 1):107–15.
- 63 Nicholson DW, Ali A, Thornberry NA, Vaillancourt JP, Ding CK, Gallant M, et al. Identification and inhibition of the ICE/ CED-3 protease necessary for mammalian apoptosis. *Nature* 1995;376(6535):37–43.