

Genetics of familial cardiomyopathies and arrhythmias

Dagmar I. Keller^{a,b}, Lucie Carrier^a, Ketty Schwartz^a

^a INSERM U523, Hôpital Pitié-Salpêtrière, Paris, France

^b Cardiology, University Hospital, Basel, Switzerland

Summary

Molecular cardiology has become an important tool in understanding the aetiology, pathogenesis and development of familial cardiomyopathies and arrhythmias. The knowledge of genotype-phenotype correlations in certain pathologies has changed the concepts of therapy. In monogenic diseases, genetic testing offers a new complementary diagnostic approach. A genetic test can be used to confirm a clinically determined diagnosis, predict prognosis in a clinically affected patient, or provide options for therapy in patients and in clinically unaffected relatives of a patient with the disease producing mutation. In pure forms of familial hypertrophic cardiomyopathy mutations in several genes coding for sarcomeric proteins have been identified, indicating wide locus heterogeneity. Various disease genes are implicated in familial dilated cardiomyopathy in the pure form or in combination with other diseases. In the long QT syndrome and Brugada syndrome, mutations in ion channel genes can cause the dis-

ease; one of those genes is also implicated in progressive cardiac conduction defect. In other familial diseases like the arrhythmogenic right ventricular cardiomyopathy, anyone of the numerous chromosomal loci can be involved, but only one gene has been identified so far. The same gene is also involved in catecholaminergic polymorphic ventricular tachycardia. From genotype-phenotype studies, correlations between gene-mutations and the clinical course of the disease have become clear. As only a few families with the same mutations have been studied, data have to be considered as preliminary and any conclusion must be regarded as tentative. This emphasizes the need to study genotype-phenotype correlations in a large number of families.

Key words: molecular cardiology; inherited; cardiomyopathies; arrhythmias; gene mutations; genotype-phenotype-correlations

Introduction

During the last decade, molecular cardiology has provided important new insights into the mechanisms responsible for inherited cardiomyopathies and arrhythmias. As a result of the ability to detect the disease-causing genes and their proteins, it has become obvious that multiple cellular components are involved in the different pathologies. The genetically determined cardiomyopathies and arrhythmias are inherited as an autosomal dominant trait, but for some, an autosomal recessive inheritance has also been identified. In monogenic diseases a mutation in a single gene is sufficient to cause the disease phenotype, whereas double or compound heterozygous mutations, homozygous mutations or gene modifiers can influence the phenotype dramatically. By analysing families with the same pathology it was apprehended that most of the mutations are private mu-

tations, indicating that most of the families have a different mutation.

If a gene-test is considered, genetic counselling of the patient is necessary, and informed consent has to be obtained. It is crucial to determine the phenotype of the family in a sporadic case or in the individual if a disease is caused by mutations in a single gene. The strategy of testing depends on the characteristics of the disease gene, like the gene-size and available samples. If several genes are known to be involved, the choice of the targeted gene should be determined by factors like the established genotype-phenotype-correlations. Some genes can be excluded by linkage analysis, which can be performed in large families. Theoretically all genes involved in a pathology can be screened, but in a routine diagnostic setting it is time-consuming and technically impossible. At

present, mutations in disease genes can be detected after gene amplification by polymerase chain reaction (PCR), followed by a mutation screening technique (single strand conformation polymorphism [SSCP], denaturing high performance liquid chromatography [DHPLC] or other) and finally by sequencing. These techniques are highly specific if different conditions are analysed and if the PCR primers largely surround the exons and important intronic sites. The sensitivity depends on several factors, particularly, one cannot exclude the implication of another not yet identified gene and/or the fact that the PCR primers may hide a point mutation.

In some familial cardiomyopathies and arrhythmias chromosomal loci and genes are known, while in others, loci have been discovered but the disease genes have not yet been identified. This review will focus on inherited cardiomyopathies and arrhythmias for which at least one responsible disease gene is known and genotype-phenotype-correlations studies are, at least in part, available. It is important to know that the range of inherited monogenic and multigenic cardiovascular diseases is even wider.

Familial cardiomyopathies

Hypertrophic cardiomyopathy

Hypertrophic cardiomyopathy (HCM) is characterized by a left and/or right ventricular hypertrophy that is usually asymmetric and involves the interventricular septum. Morphological changes include hypertrophy and disarray of myocytes and fibrosis. Arrhythmias and sudden cardiac death (SCD) are common [1], and HCM often results in heart failure. The familial type of primary hypertrophic cardiomyopathy (FHC) includes a group of HCM lacking a defined aetiology and is characterized by a high rate of morbidity and mortality. FHC is an autosomal dominant disease with incomplete penetrance. Its prevalence is 1/500 in young adults [2]. Eleven genes have been identified in the aetiology of the pure form of FHC (table 1). They all encode proteins of the cardiac sarcomere. Recently, mutations in the *PRKAG2* gene, coding for the γ 2-subunit of the AMP-activated protein-kinase, were found to be responsible for familial Wolff-Parkinson-White-Syndrome (WPW) with cardiac hypertrophy [3]

or for WPW with conduction defect and absence of cardiac hypertrophy [4]. Around 150 mutations have been identified [5], and most of them are found in genes *MYH7*, *MYBPC3* and *TNNT2*. By systematic screening of nine of the eleven genes, recent data have shown that 88% of genotyped families present a mutation in the *MYBPC3* and *MYH7* genes ([6], AHA 2001).

Mutations leading to FHC are missense or frameshift mutations. The molecular mechanisms have not been completely elucidated. Missense mutations result in stable mutant proteins that act by a dominant negative effect on the structure or function of the sarcomere (poison peptide hypothesis). In contrast, frameshift mutations result in unstable truncated proteins, and therefore act as "null-allele" leading to haploinsufficiency of the "wilde-type" protein (haploinsufficiency hypothesis) [7].

The FHC phenotype is first influenced by factors varying the penetrance, like age and gender [8]. Second, the phenotype depends on the

Table 1
Familial hypertrophic cardiomyopathy genes, proteins and mutations.

Gene	locus	protein	mutations	frequency
<i>MYH7</i>	14q11.2–q12	β -myosin heavy chain (β -MyHC)	70	>35–50%
<i>MYH6</i>	14q11.2–q12	α -myosin heavy chain (α -MyHC)	1	?
<i>MYL3</i>	3p21.2–p21.2	ventricular essential myosin light chain (MLC-1s/v)	2	<1%
<i>MYL2</i>	12q23–q24.3	ventricular regulatory myosin light chain (MLC-2s/v)	8	<1%
<i>ACTC</i>	15q14	α -cardiac actin (α -cAct)	5	?
<i>TNNT2</i>	1q32	cardiac troponin T (cTnT)	14	15–20%
<i>TNNI3</i>	19p13.4	cardiac troponin I (cTnI)	8	<1%
<i>TNNC1</i>	3p21.3	cardiac troponin C (cTnC)	1	?
<i>TPMI</i>	15q22.1	α -tropomyosin (α -TM)	5	<5%
<i>MYBPC3</i>	11p11.2	Cardiac myosin-binding protein C (cMyBP-C)	30	>15–20%
<i>TTN</i>	2q24.3	Titin	1	?
<i>PRKAG2</i>	7q36	AMP-activated protein kinase (AMPK)	2	?

Eleven FHC genes code for sarcomeric proteins: genes *MYH7*, *MYH6*, *MYL2* and *MYL3* code for the thick filament proteins, genes *TNNC1*, *TNNI3*, *TNNT2*, *TPMI* and *ACTC* for the thin filament proteins, and gene *MYBPC3* codes for the myosin-binding protein C. The titin-protein, coded by *TTN*, is the third filament of the sarcomere. Gene *PRKAG2* codes for the γ 2-subunit of the AMP-activated protein kinase.

The last column indicates the frequency of the mutations found in the different genes.

responsible mutation. In the *MYH7* gene several mutations are associated with a high risk of SCD: Arg403Gln, Arg719Trp, Arg453Cys and Arg723Gly. In contrast, hearts with the mutations Gly256Glu, Val606Met and Leu908Val have a good prognosis as regards arrhythmic events [9, 10]. Mutations in the *MYBPC3* gene are associated with mild hypertrophy in young patients, late onset of symptoms and good prognosis before the age of forty [11, 12]. The penetrance and clinical presentation of mutations in *TNNT2* vary widely: some mutations lead to subclinical hypertrophy associated with high SCD-risk [13, 14], and others are completely penetrant but without a high risk for arrhythmic events [15]. Third, the phenotype depends on the complexity of the genotype. About 8% of the families have a complex genotype with homozygous, double or compound heterozygous mutations [6]. A homozygous Arg869Gly mutation is associated with early onset of severe left ventricular hypertrophy, systolic dysfunction and atrial fibrillation [16]. Double heterozygous mutations in the *MYH7* and *MYBPC3* genes lead to significant hypertrophy, but are not strictly associated with a high SCD-risk [17]. Finally, about 25% of genotypically affected patients do not develop a FHC phenotype. This could be explained by environmental and/or genetic factors. Modifier poly-

morphisms have been identified in the genes coding for the angiotensin converting enzyme [18, 19], endothelin [20] and the angiotensin II type 1 receptor [21].

Mutations associated with a high SCD risk are important when considering therapeutical options, like the implantation of a cardioverter/defibrillator. Further studies in large and numerous families and animal models will help to identify the underlying mechanisms of the pathogenesis of FHC.

Dilated cardiomyopathy

Dilated cardiomyopathy (DCM) is characterized by left ventricular dilation with impaired contraction, often with involvement of the right ventricle. It represents a leading cause for cardiac transplantation due to heart failure. The prevalence is nearly 40/100,000 [22]. The 5-year mortality rate varies between 15 and 50%, SCD accounts for 30% of deaths in DCM. At least 25% of the DCM are familial [23]. Familial DCM is clinically and genetically heterogeneous. Autosomal dominant DCM, pure or in association with other pathologies, is the most common segregated form, whereas autosomal recessive and X-linked inheritances are rare. Table 2 gives an overview of the DCM inheritance patterns, genes and their

Table 2
Familial dilated cardiomyopathy phenotypes, genes and proteins.

Inheritance	phenotype	locus	gene	protein
Autosomal dominant	pure DCM	9q12-q13	?	
	pure DCM	1q32	?	
	pure DCM	2q24.3-q31	<i>TTN</i>	titin
	pure DCM	6q12-q16	?	
	pure DCM	2q35	<i>DES</i>	desmin
	pure DCM	5q33	<i>SGCD</i>	δ-sarcoglycan
	pure DCM	15q11-qter	<i>ACTC</i>	actin
	pure DCM, early onset	14q11.2	<i>MYH7</i>	β-myosin heavy chain
	pure DCM, early onset	1q32	<i>TNNT2</i>	cardiac troponin T
Autosomal dominant +	DCM + CD	1q21	<i>LMNA</i>	lamin A/C
	DCM + CD	2q14-q22	?	
	DCM + CD + SND	3p22-p25	?	
	DCM + MVP	10q21-q23	?	
	DCM + hearing loss	6q23-q24	<i>EYA4</i>	eyes absent 4
	DCM + CD + LGMD	6q22-q23	?	
	DCM + CD + MD (AD-EDMD)	1q21	<i>LMNA</i>	lamin A/C
	DCM + CD + LGMD (LGMD1B)	1q21	<i>LMNA</i>	lamin A/C
Autosomal recessive	LGMD +/- cardiomyopathy	17q21	<i>SGCA</i>	α-sarcoglycan
	LGMD + severe cardiomyopathy	4q12	<i>SGCB</i>	β-sarcoglycan
	LGMD + cardiomyopathy (Brazil)	5q33	<i>SGCD</i>	δ-sarcoglycan
X-linked	Pure DCM	Xp21.3	<i>DYS</i>	dystrophin
	DCM lethal in infancy	Xq28	<i>TAZ</i>	tafazzin
	DCM + myopathy (Barth-Syndrome)	Xq28	<i>TAZ</i>	tafazzin
	DCM + CD + MD (XL-EDMD)	Xq28	<i>EMD</i>	emerin

Abbreviations: DCM, dilated cardiomyopathy; CD, conduction defect; SND, sinus node dysfunction; MVP, mitral valve prolapse; LGMD, limb girdle muscular dystrophy; MD, muscular dystrophy; EDMD, Emery-Dreifuss muscular dystrophy; AD, autosomal dominant; XL, X-linked.

proteins, and the resulting phenotypes (adapted from [24]). Thirteen genes have been identified, coding for proteins of the cytoskeleton and interacting elements, sarcolemma and sarcomere. Pure DCM is inherited as an autosomal dominant trait and six disease genes have been identified: *TTN*, *DES*, *SGCD*, *ACTC*, *MYH7* and *TNNT2*. Autosomal dominant DCM can be associated with other cardiac and/or muscle pathologies or hearing loss; several loci, but only two genes, *LMNA* and *EYA4*, have been identified. Autosomal recessive DCM, a rare disease due to mutations in the genes coding for sarcoglycans, is always associated with limb gir-

dle muscular dystrophy. In the X-linked DCM, a broad range of clinical pictures is possible due to mutations in genes *DYS*, *TAZ* and *EMD*.

Due to the wide clinical and genetic heterogeneity and the lack of genotype-phenotype correlation studies in representative populations, the impact of a genetic test in DCM is still limited. Studies of the molecular defects of DCM and the disease genes in large populations will provide important insights into the mechanisms of heart failure and may lead to the modification of treatment options.

Familial arrhythmias

Congenital long QT syndrome

Congenital long QT syndrome (LQTS) is characterized by abnormal ventricular repolarization with QTc prolongation >440 ms, and high risk of Torsades de pointes and malignant ventricular tachyarrhythmias. LQTS is an autosomal-dominant or -recessive disease in which the sites of dysfunction are the ion channels, which consist of proteins allowing much greater ion flow during the cardiac action potential. The disease prevalence is about 1/5,000. The autosomal-dominant Romano-Ward syndrome is the most common form [25, 26]. A second type, the Jervell-Lange-Nielsen syndrome, which is associated with congenital deafness, is transmitted as an autosomal-recessive trait and has a poor prognosis [27]. Six chromosomal loci and five disease genes have been identified (table 3). Almost 70% of the mutations are missense mutations, and 87% of the mutations are found in genes *KCNQ1* and *KCNH2* [28]. This frequency may change in the future because those two genes were first identified and therefore extensively screened. *KCNQ1* (*KvLQT1*) codes for the α -subunit of the slowly activating delayed rectifier potassium ion channel $I_{Ks\alpha}$ and *KCNE1* for the β -subunit; *KCNH2* (*HERG*) codes for the α -subunit of the rapidly activating delayed rectifier potassium ion channel $I_{Kr\alpha}$ and *KCNE2* for the β -subunit. *SCN5A* is the gene coding for a cardiac sodium ion channel I_{Na} . The gene of *LQT4* has not

been identified yet. Functional consequences of mutations in LQTS genes are a loss of the channel function in the genes encoding the I_{Ks} and I_{Kr} cardiac potassium channels [29], and additional sodium channel activity or gain of function in *SCN5A* mutations [30]. Due to studies performed on the mutations of the three major genes, the QT-interval, and T-wave pattern of the surface ECG, gene-ECG correlations have been established: generally, mutations in *KCNQ1* lead to a broad T-wave, and mutations in *KCNH2* to a low T-wave amplitude with double peak in some cases. Mutations in *SCN5A* are characterized by a very delayed onset of the T-wave [31].

The identification of gene mutations in the LQTS can modify the specific treatment. Beta-blockers are efficient in *LQT1* and *LQT2*, whereas mexiletine, a sodium channel blocker, is a treatment option in *LQT3* [32, 33]. Pacing is important in *LQT3* to avoid tachyarrhythmic events induced by bradycardia. In contrast *LQT2* patients do not benefit from pacing [34]. Triggers associated with the induction of Torsades de pointes and ventricular arrhythmias have been described: physical exercise especially swimming in *LQT1*, auditory stimuli during sleep or rest in *LQT2*, and sleep or rest in *LQT3* [35]. In the presence of certain disease genes, modifications in life-style to avoid triggers are possible. About 10% of mutation carriers present with a normal ECG [36]. In these

Table 3
LQTS genes, proteins and frequency of mutations.

LQT	gene	locus	protein	frequency of mutations
LQT1	<i>KCNQ1</i> (<i>KvLQT1</i>)	11p15.5	Cardiac potassium ion channel $I_{Ks\alpha}$	42%
LQT2	<i>KCNH2</i> (<i>HERG</i>)	7q35–q36	Cardiac potassium ion channel $I_{Kr\alpha}$	45%
LQT3	<i>SCN5A</i>	3p21	Cardiac sodium ion channel I_{Na}	8%
LQT4	?	4q25–q27		
LQT5	<i>KCNE1</i>	21q22.1–22.2	Cardiac potassium ion channel $I_{Ks\beta}$ subunit <i>KCNQ1</i>	3%
LQT6	<i>KCNE2</i>	21q22.1	Cardiac potassium ion channel $I_{Kr\beta}$ subunit <i>KCNH2</i>	2%

patients the value of a beta-blocker treatment is unclear.

Most of the patients with drug induced LQTS do not have ion channel defects, but they seem to carry a predisposition for malignant arrhythmias [37]. The value of genetic diagnosis is unclear; those patients should avoid any LQTS-provoking drugs.

Gene *SCN5A*: Brugada syndrome and progressive cardiac conduction defect

Brugada syndrome belongs, as the LQTS, to the group of idiopathic ventricular fibrillation, characterized by a primary electrical disease without underlying structural heart pathology. The Brugada syndrome is defined by a right bundle branch block, a right precordial ST-segment elevation and SCD, mainly during sleep [38]. No data are available on the prevalence of the Brugada syndrome, but a study showed that there is a high prevalence (0.1%) of the typical ECG pattern in an apparently healthy population [39]. *SCN5A* is the only gene so far known to be involved in the disease [40]. *SCN5A* mutations are found in about 20% of the patients with clinical Brugada syndrome [41]. Recently, a second locus was found on chromosome 3, indicating locus heterogeneity in this disease [42].

Mutations in *SCN5A* may also cause the *LQT3* and the progressive cardiac conduction defect found in the Lenegre-Lev-disease [43]. Progressive cardiac conduction defect is characterized by continuous alteration of cardiac conduction through the His-Purkinje system, associated with right or left bundle branch block and QRS-complex widening, finally leading to complete atrio-ventricular block.

The mechanisms by which mutations in *SCN5A* lead to different diseases are unclear, yet. Basically, *SCN5A* mutations implicated in *LQT3* cause a gain of function of the channel, whereas the ones implicated in the Brugada syndrome cause a loss of channel function [44].

Gene *RyR2*: catecholaminergic polymorphic ventricular tachycardia and arrhythmogenic right ventricular cardiomyopathy

Catecholaminergic polymorphic ventricular tachycardia (C-PMVT) is a disease of the group of idiopathic ventricular fibrillation. Affected patients experience polymorphic ventricular extrasystoles and tachycardia during exercise or stress, which can degenerate in malignant ventricular arrhythmias and cardiac death. Gene *RyR2*, which codes for the human cardiac ryanodin 2-receptor protein, is implicated in C-PMVT [45]. This protein induces the release of Ca^{2+} from the sarcoplasmic reticulum into the cytosol. A second gene, *CASQ2*, has been identified in the context of C-PMVT [46]. The encoded calsequestrin protein is an internal constituent of the sarcoplasmic reticulum.

Arrhythmogenic right ventricular cardiomyopathy (ARVC) is characterized by a fibro-fatty replacement of the right ventricular myocardium with possible involvement of the left ventricle. Monomorphic ventricular tachyarrhythmias with left bundle branch block pattern are typical. The prevalence of ARVC is probably higher than 6/10,000; it is the major cause of SCD in young athletes. Mutations in the *RyR2* gene have been identified in ARVC2. In addition, five chromosomal loci are known [47, 48]. Naxos disease is the autosomal recessive form associated with skin and hair abnormalities. Mutations in the gene coding for plakoglobin, a cell adhesion protein, have been found in this disease [49, 50].

The impact of the gene-test in C-PMVT and ARVC is unclear. Only a few families have been studied so far. Larger population studies are needed to determine genotype-phenotype-correlations, in order to help identify individuals at risk.

Perspectives of genetic studies

For optimal patient care and definition of disease groups, genotype-phenotype correlation studies in numerous large families are needed. The identification of mutations in the major gene is important, but other relevant factors like gene modifiers are also important because they influence the phenotype and clinical course. For certain inherited diseases like FHC, mutations with a high SCD risk have been identified. In the LQTS, identification of mutations profoundly modifies the medical treatment and lifestyle. For other diseases, genotype-phenotype-correlations are not yet available.

We are grateful to Professor Saul Winegrad for his helpful comments on the manuscript.

Correspondence:

Dagmar Keller, MD

INSERM U523, Institut de Myologie

Bâtiment Joseph Babinski

Groupe Hospitalier Pitié-Salpêtrière

47, boulevard de l'Hôpital

F-75651 Paris Cedex 13

E-Mail: d.keller@myologie.chups.jussieu.fr

References

- Report of the 1995 World Health Organization/International Society and Federation of Cardiology Task Force on the Definition and Classification of Cardiomyopathies. *Circulation* 1996;93:841–2.
- Maron BJ, Gardin JM, Flack JM, Gidding SS, Kurosaki TT, Bild DE. Prevalence of hypertrophic cardiomyopathy in a general population of young adults. Echocardiographic analysis of 4111 subjects in the CARDIA Study. Coronary Artery Risk Development in (Young) Adults. *Circulation* 1995;92:785–9.
- Gollob MH, Green MS, Tang AS, Gollob T, Karibe A, Ali Hassan AS, et al. Identification of a gene responsible for familial Wolff-Parkinson-White syndrome. *N Engl J Med* 2001;344:1823–31.
- Gollob MH, Seger JJ, Gollob TN, Tapscott T, Gonzales O, Bachinski L, et al. Novel PRKAG2 mutation responsible for the genetic syndrome of ventricular preexcitation and conduction system disease with childhood onset and absence of cardiac hypertrophy. *Circulation* 2001;104:3030–3.
- Fung DC, Yu B, Littlejohn T, Trent R J. An online locus-specific mutation database for familial hypertrophic cardiomyopathy. *Hum Mutat* 1999;14:326–32.
- Richard P, Charron P, Carrier L, Ledeuil L, Dubourg O, Desnos M, et al. Distribution of disease genes in 102 genotyped families with hypertrophic cardiomyopathy. *Circulation* 2001;104(suppl.):II-521.
- Vignier N, Perrot A, Schulte HD, Richard P, Sebillon P, Schwartz K, et al. Cardiac myosin-binding protein C and familial hypertrophic cardiomyopathy: from mutations identification to human endomyocardial proteins analysis. *Circulation* 2001;104(suppl.):II-1.
- Charron P, Carrier L, Dubourg O, Tesson F, Desnos M, Richard P, et al. Penetrance of familial hypertrophic cardiomyopathy. *Genet Couns* 1997;8:107–14.
- Marian AJ, Roberts R. Molecular genetic basis of hypertrophic cardiomyopathy: genetic markers for sudden cardiac death. *J Cardiovasc Electrophysiol* 1998;9:88–99.
- Enjuto M, Francino A, Navarro-Lopez F, Viles D, Pare JC, Ballesta AM. Malignant hypertrophic cardiomyopathy caused by the Arg723Gly mutation in beta-myosin heavy chain gene. *J Mol Cell Cardiol* 2000;32:2307–13.
- Niimura H, Bachinski LL, Sangwatanaroj S, Watkins H, Chudley AE, McKenna W, et al. Mutations in the gene for cardiac myosin-binding protein C and late-onset familial hypertrophic cardiomyopathy. *N Engl J Med* 1998;338:1248–57.
- Charron P, Dubourg O, Desnos M, Bennaceur M, Carrier L, Camproux AC, et al. Clinical features and prognostic implications of familial hypertrophic cardiomyopathy related to cardiac myosin-binding protein C gene. *Circulation* 1998;97:2230–6.
- Moolman JC, Corfield VA, Posen B, Ngumbela K, Seidman C, Brink PA, et al. Sudden death due to troponin T mutations. *J Am Coll Cardiol* 1997;29:549–55.
- Thierfelder L, Watkins H, MacRae C, Lamas R, McKenna W, Vosberg HP, et al. Alpha-tropomyosin and cardiac troponin T mutations cause familial hypertrophic cardiomyopathy: a disease of the sarcomere. *Cell* 1994;77:701–12.
- Forissier JF, Carrier L, Farza H, Bonne G, Bercovici J, Richard P, et al. Codon 102 of the cardiac troponin T gene is a putative hot spot for mutations in familial hypertrophic cardiomyopathy. *Circulation* 1996;94:3069–73.
- Richard P, Charron P, Leclercq C, Ledeuil C, Carrier L, Dubourg O, et al. Homozygotes for a R869G mutation in the beta-myosin heavy chain gene have a severe form of familial hypertrophic cardiomyopathy. *J Mol Cell Cardiol* 2000;32:1575–83.
- Richard P, Isnard R, Carrier L, Dubourg O, Donatien Y, Mathieu B, et al. Double heterozygosity for mutations in the beta-myosin heavy chain and in the cardiac myosin binding protein C genes in a family with hypertrophic cardiomyopathy. *J Med Genet* 1999;36:542–5.
- Marian AJ, Yu QT, Workman R, Greve G, Roberts R. Angiotensin-converting enzyme polymorphism in hypertrophic cardiomyopathy and sudden cardiac death. *Lancet* 1993;342:1085–6.
- Tesson F, Dufour C, Moolman JC, Carrier L, al-Mahdawi S, Chojnowska L, et al. The influence of the angiotensin I converting enzyme genotype in familial hypertrophic cardiomyopathy varies with the disease gene mutation. *J Mol Cell Cardiol* 1997;29:831–8.
- Brugada R, Kelsey W, Lechin M, Zhao G, Yu QT, Zoghbi W, et al. Role of candidate modifier genes on the phenotypic expression of hypertrophy in patients with hypertrophic cardiomyopathy. *J Investig Med* 1997;45:542–51.
- Osterop AP, Kofflard MJ, Sandkuijl LA, ten Cate FJ, Krams R, Schalekamp MA, et al. AT1 receptor A/C1166 polymorphism contributes to cardiac hypertrophy in subjects with hypertrophic cardiomyopathy. *Hypertension* 1998;32:825–30.
- Durand JB, Abchee AB, Roberts R. Molecular and clinical aspects of inherited cardiomyopathies. *Ann Med* 1995;27:311–7.
- Keeling PJ, Gang Y, Smith G, Seo H, Bent SE, Murday V, et al. Familial dilated cardiomyopathy in the United Kingdom. *Br Heart J* 1995;73:417–21.
- Marcelis C, et al. Dilated Cardiomyopathy in Cardiovascular Genetics for Clinicians (Doevendans PA and Wilde AA, eds), Kluwer Academic Publishers, 2001, p 155–67.
- Romano C, Gemme G, Pongiglione R. Aritmie cardiache rare in eta pediatrica. *Clin Pediatr* 1963;45:656–83.
- Ward OC. A new familial cardiac syndrome in children. *J Irish Med Assoc* 1964;54:103–6.
- Jervell A, Lange-Nielsen F. Congenital deaf-mutism, functional heart disease with prolongation of the Q-T interval and sudden death. *Am Heart J* 1957;54:59–68.
- Splawski I, Shen J, Timothy KW, Lehmann MH, Priori S, Robinson JL, et al. Spectrum of mutations in long-QT syndrome genes KVLQT1, HERG, SCN5A, KCNE1, and KCNE2. *Circulation* 2000;102:1178–85.
- Sanguinetti MC, Curran ME, Spector PS, Keating MT. Spectrum of HERG K⁺-channel dysfunction in an inherited cardiac arrhythmia. *Proc Natl Acad Sci U S A* 1996;93:2208–12.
- Clancy CE, Rudy Y. Linking a genetic defect to its cellular phenotype in a cardiac arrhythmia. *Nature* 1999;400:566–9.
- Roden DM, Lazzara R, Rosen M, Schwartz PJ, Towbin J, Vincent GM. Multiple mechanisms in the long-QT syndrome. Current knowledge, gaps, and future directions. The SADS Foundation Task Force on LQTS. *Circulation* 1996;94:1996–2012.
- Moss AJ, Zareba W, Hall WJ, Schwartz PJ, Crampton RS, Benhorin J, et al. Effectiveness and limitations of beta-blocker therapy in congenital long-QT syndrome. *Circulation* 2000;101:616–23.
- Wilde AA, Roden DM. Predicting the long-QT genotype from clinical data: from sense to science. *Circulation* 2000;102:2796–8.
- Schwartz PJ, Priori SG, Locati EH, Napolitano C, Cantu F, Towbin JA, et al. Long QT syndrome patients with mutations of the SCN5A and HERG genes have differential responses to Na⁺ channel blockade and to increases in heart rate: implications for gene-specific therapy. *Circulation* 1995;92:3381–6.
- Schwartz PJ, Priori SG, Spazzolini C, Moss AJ, Vincent GM, Napolitano C, et al. Genotype-phenotype correlation in the long-QT syndrome: gene-specific triggers for life-threatening arrhythmias. *Circulation* 2001;103:89–95.
- Zhang L, Timothy KW, Vincent GM, Lehmann MH, Fox J, Giuli LC, et al. Spectrum of ST-T-wave patterns and repolarization parameters in congenital long-QT syndrome: ECG findings identify genotypes. *Circulation* 2000;102:2849–55.
- Roden DM. Mechanisms and management of proarrhythmia. *Am J Cardiol* 1998;82:491–571.
- Brugada P, Brugada J. Right bundle branch block, persistent ST segment elevation and sudden cardiac death: a distinct clinical and electrocardiographic syndrome. A multicenter report. *J Am Coll Cardiol* 1992;20:1391–6.
- Hermida JS, Lemoine JL, Aoun FB, Jarry G, Rey JL, Quiret JC. Prevalence of the Brugada syndrome in an apparently healthy population. *Am J Cardiol* 2000;86:91–4.
- Chen Q, Kirsch GE, Zhang D, Brugada R, Brugada J, Brugada P, et al. Genetic basis and molecular mechanism for idiopathic ventricular fibrillation. *Nature* 1998;392:293–6.
- Priori SG, Napolitano C, Gasparini M, Pappone C, Della Bella P, Giordano U, et al. Natural history of Brugada syndrome: insights for risk stratification and management. *Circulation* 2002;105:1342–7.
- Weiss R, Barmada MM, Nguyen T, Seibel JS, Cavlovich D, Kornblit CA, et al. Clinical and molecular heterogeneity in the Brugada syndrome: a novel gene locus on chromosome 3. *Circulation* 2002;105:707–13.

- 43 Schott JJ, Alshinawi C, Kyndt F, Probst V, Hoorntje TM, Hulsbeek M, et al. Cardiac conduction defects associate with mutations in SCN5A. *Nat Genet* 1999;23:20-1.
- 44 Veldkamp MW, Viswanathan PC, Bezzina C, Baartscheer A, Wilde AA, Balseer JR. Two distinct congenital arrhythmias evoked by a multidysfunctional Na(+) channel. *Circ Res* 2000;86:E91-7.
- 45 Laitinen PJ, Brown KM, Piippo K, Swan H, Devaney JM, Brahmabhatt B, et al. Mutations of the cardiac ryanodine receptor (RyR2) gene in familial polymorphic ventricular tachycardia. *Circulation* 2001;103:485-90.
- 46 Lahat H, Pras E, Olender T, Avidan N, Ben-Asher E, Man O, et al. A missense mutation in a highly conserved region of CASQ2 is associated with autosomal recessive catecholamine-induced polymorphic ventricular tachycardia in Bedouin families from Israel. *Am J Hum Genet* 2001;69:1378-84.
- 47 Tiso N, Stephan DA, Nava A, Bagattin A, Devaney JM, Stanchi F, et al. Identification of mutations in the cardiac ryanodine receptor gene in families affected with arrhythmogenic right ventricular cardiomyopathy type 2 (ARVD2). *Hum Mol Genet* 2001;10:189-94.
- 48 Li D, Ahmad F, Gardner MJ, Weilbaecher D, Hill R, Karibe A, et al. The locus of a novel gene responsible for arrhythmogenic right-ventricular dysplasia characterized by early onset and high penetrance maps to chromosome 10p12-p14. *Am J Hum Genet* 2000;66:148-56.
- 49 Protonotarios N, Tsatsopoulou A, Patsourakos P, Alexopoulos D, Gezerlis P, Simitsis S, et al. Cardiac abnormalities in familial palmoplantar keratosis. *Br Heart J* 1986;56:321-6.
- 50 McKoy G, Protonotarios N, Crosby A, Tsatsopoulou A, Anastakis A, Coonar A, et al. Identification of a deletion in plakoglobin in arrhythmogenic right ventricular cardiomyopathy with palmoplantar keratoderma and woolly hair (Naxos disease). *Lancet* 2000;355:2119-24.

The many reasons why you should choose SMW to publish your research

What Swiss Medical Weekly has to offer:

- SMW's impact factor has been steadily rising, to the current 1.537
- Open access to the publication via the Internet, therefore wide audience and impact
- Rapid listing in Medline
- LinkOut-button from PubMed with link to the full text website <http://www.smw.ch> (direct link from each SMW record in PubMed)
- No-nonsense submission – you submit a single copy of your manuscript by e-mail attachment
- Peer review based on a broad spectrum of international academic referees
- Assistance of our professional statistician for every article with statistical analyses
- Fast peer review, by e-mail exchange with the referees
- Prompt decisions based on weekly conferences of the Editorial Board
- Prompt notification on the status of your manuscript by e-mail
- Professional English copy editing
- No page charges and attractive colour offprints at no extra cost

Editorial Board

Prof. Jean-Michel Dayer, Geneva
 Prof. Peter Gehr, Berne
 Prof. André P. Perruchoud, Basel
 Prof. Andreas Schaffner, Zurich
 (Editor in chief)
 Prof. Werner Straub, Berne
 Prof. Ludwig von Segesser, Lausanne

International Advisory Committee

Prof. K. E. Juhani Airaksinen, Turku, Finland
 Prof. Anthony Bayes de Luna, Barcelona, Spain
 Prof. Hubert E. Blum, Freiburg, Germany
 Prof. Walter E. Haefeli, Heidelberg, Germany
 Prof. Nino Kuenzli, Los Angeles, USA
 Prof. René Lutter, Amsterdam, The Netherlands
 Prof. Claude Martin, Marseille, France
 Prof. Josef Patsch, Innsbruck, Austria
 Prof. Luigi Tavazzi, Pavia, Italy

We evaluate manuscripts of broad clinical interest from all specialities, including experimental medicine and clinical investigation.

We look forward to receiving your paper!

Guidelines for authors:

http://www.smw.ch/set_authors.html

Impact factor Swiss Medical Weekly



All manuscripts should be sent in electronic form, to:

EMH Swiss Medical Publishers Ltd.
 SMW Editorial Secretariat
 Farnsburgerstrasse 8
 CH-4132 Muttenz

Manuscripts: submission@smw.ch
 Letters to the editor: letters@smw.ch
 Editorial Board: red@smw.ch
 Internet: <http://www.smw.ch>